# A mitogenomic phylogeny and genetic history of Amphioctopus fangsiao (d'Orbigny 1839-1841) from China

Lashari P.<sup>1</sup>,<sup>3</sup>; Wei Ch.; Gong L.<sup>1</sup>; Liu L.<sup>1</sup>; Jiang L.<sup>1</sup>; Liu B.<sup>1</sup>; Muhammad F.<sup>2</sup>; Laghari M.Y.<sup>3</sup>; Lashari Kh.H.<sup>3</sup>; Waryani B.<sup>3</sup>; Hlaing N.N.S.<sup>4</sup>; Yingying Ye<sup>1</sup>; Lü Z.<sup>1\*</sup>

Received: March 2019

Accepted: November 2019

## Abstract

Phylogeny and genetic diversity of *Amphioctopus fangsiao* were assessed by sequence analysis of complete mitochondrial genomes, sequenced from 15 individuals of nine populations. The whole mtDNA genomes size were ranging from 15977 to 15990 bp. Data revealed 1642 polymorphic sites and 1023 parsimony informative sites. The phylogenetic analysis based on neighbor joining tree disclosed two clades. It consisted of four (Dalian, Yantai, Qingdao and Nantong) and five populations (Shanghai, Zhoushan, Xiamen, Dongshan and Zhanjiang). Genetic differentiation coefficient (FST) was recorded higher i.e 0.61476. While, the AMOVA analysis showed that 61.48% of the genetic variation existed between the two clades. However, only 38.52% of the genetic variation existed within each clade. In further, the net genetic distance between the two groups was 0.030. The possible reason of differentiation is quaternary glacial period and Yangtze River.

**Keywords:** *Amphioctopus fangsiao*, mtDNA, genetic differentiation, phylogeny, populations.

<sup>1-</sup>National Engineering Research Center of Marine Facilities Aquaculture, College of Marine Sciences and Technology, Zhejiang Ocean University, No.1, Haida South Road, Lincheng Changzhi Island, Zhoushan, Zhejiang, 316022 P.R. China

<sup>2-</sup>Center of Excellence in Marine Biology, University of Karachi, Main University Road Karachi 75270. Sindh, Pakistan

<sup>3-</sup>Department Fresh Water Biology & Fisheries, Allama I.I.Kazi Campus, University of Sindh, Jamshoro-76080, Sindh, Pakistan.

<sup>4-</sup>Biotechnology Research Departments, Ministry of Education, Tazoe Gate, Kyaukse, Union of Myanmar

Corresponding author's Emial: nblzmnb@163.com

## Introduction

Amphioctopus fangsiao belongs to family Octopodidae. distributes It west Pacific widely in Ocean. especially in Yellow Sea and Bohai Sea (Dong, 1988; Lu et al., 2012). A. fangsiao considered as a potential species for aquaculture (Wang et al., 2015), due to its rapid growth rate and high nutritional value. Tons of its fisheries have annually been harvested alone in Shandong province. However, it has been reported that its natural population is decreasing because of over-exploitation (Zhang et al., 2009). Therefore, it is intuitive to have its resource management and conservation. Until now, few studies are available to elucidate A. fangsiao genetic aspects (Gao et al., 2002; Zhang et al., 2009, and Lü et al., 2010, Muhammad et al. 2019). Gao et al. (2002) and Zhang et al. (2009) used allozyme and amplified fragment length polymorphism (AFLP). Lü et al. (2011, 2010) examined several populations Α. fangsiao using mitochondrial 16S rRNA and the COI gene and Muhammad et al. (2019), investigated using three mtDNA genes (ATPase 6, ND2 and ND5). All the earlier studies showed certain level of differentiation. The whole mtDNA genome approach of investigation was not carried earlier.

*A. fangsiao* is one of the marine species that spawn life time in marine ecosystem. The fluctuating physical properties of marine ecosystem influence organisms which restrict gene flow over large geographic distances

(Maltagliati et al., 2002, Hellberg et al., 2002, Fernandez et al., 2011). Unlike other marine organisms, having high range of dispersal capacity, large population sizes, high fecundity and planktonic mood of dispersal which helps in extensive gene flow (Bohonak 1999, Zid et al., 2012), the Octopus species like Octopus minor, O. vulgaris and A. fangsiao show limited dispersal capacity both at larval and adult stages (Oosthuizen et al., 2004; Kim et al., 2009; Kang et al., 2012; Lü et al., 2013, De Luca et al., 2016, 2015, Melis et al., 2018). A. fangsiao adults are either slow mover or like sessile, crawling away on the sea bed or burrowing deep in mud. It has benthos-attached eggs, spent only a few days as planktonic larval stage. Therefore, it has weak dispersal and limited gene flow. resulting in the genetic differentiation among populations.

Assessment of genetic diversity within and among populations has immense importance, which deliver a possible genetic source for future adaptation (Xu, 2012) and in area of evolutionary biology (Hao et al., 2015, Ortego et al., 2015). In natural population its level is determined by the interplay of mutation, migration, drift and selection (Harrison, 1991, Vellend and Geber 2005, Wethington et al., 2007). The role of each of these forces depends on the life history of the mating species. It's system and dispersal abilities, as well as extrinsic factors such as the landscape matrix, its history and anthropogenic actions (Gow, et al., 2004; Husemann et al., 2012). In the present study, we analyzed data of fifteen complete mitochondrial genes gained, from fifteen individuals collected from nine localities across the Chinese coast. The sequences were examined to understand the genetic diversity and phylogeny of A. fangsiao and reveal the possible issues impacting phylogeographic pattern. The the results of the present investigation provide profound information for the future management and conservation of this species.

#### Materials and methods

Samples were collected from nine locations (Fig. 1). Thereafter, they were preserved ethanol in 95% and transported to the laboratory. The total genomic DNA was isolated from muscle tissues using a standard protocol. A set of 20 PCR primers were used to amplify overlapping fragments that covered the whole mitochondrial genome of the Amphioctopus fangsiao. Primarily, primers were designed on the basis of complete mtDNA sequence Gene Bank under accession number NC\_007896.1 (Akasaki et al., 2006). specific Later on, primers were designed according to newly obtained sequences to facilitate primer walking. Thermocycler conditions were as follows: denaturation at 94 °C for 5 min, 35 cycles 94°C for 30s, annealing at 50°C for 30s, extension at 72°C for 30s, and the final extension at  $72^{\circ}C$  for 7 min. Electrophoresis was performed on a 1.2% agarose gel and was sequenced

using the same PCR primers. Sequences were aligned using CLUSTALX 1.83, Thompson et al. (1997) was used for alignment and editing of sequences and BioEdit 7.0.1, (Hall, 1999). Using CG View Server to draw the mtDNA ring diagram. The neutrality test of haplotype nucleotide and diversity indices their variances as well (Tajima's D, Fu and Li's D\*, Fu and Li's F\*) were calculated using DnaSP version 5.0 software package, (Librado and Rozas, 2009). Genetic distance calculation and cluster analysis were carried out by using Mega 6.0 software. The phylogenetic tree was constructed by UPGMA model and the bootstrap (repetition number 1000) was used to check the branch trust. Octopus Aegina (NC\_029702.1), (Zhang et al., 2015), variabilis (NC\_015896.1), Octopus (Cheng et al., 2012) Octopus vulgaris (NC 006353.1), (Yokobori et al., 2004), Octopus bimaculoides (NC 028547.1), Domínguez et al. (2015) mtDNA were the out group used as in the analysis. Phylogeography The haplotype networking was created using NETWORK software version 5.0.0.1 (Bandelt et al., 1999). The genetic structure of the population was assessed FST using the statistic, and its significance repeated using was Arlequin 3.5 for 1 000 cycles. The relationship between (1-FST) / 2-FST and the geographical distance of the sample is plotted to determine whether population the genetic structure conforms to the geo-isolation model (Slatkin, 1993).



Figure 1: Map showing collection sites of Amphioctopus fangsiao specimens.

The genetic structure of the population was further detected using the molecular variation analysis AMOVA method in Arlequin 3.5 software. Homoplasic sites were exposed by the the of phylogeny use network investigation as applied in the Network 4.6 program.

The historical dynamics of the population was carried out using two methods: (1) using Tajima's D and Fu's Fs test to determine whether the neutral hypothesis was established, Tajima's D and Fu's Fs neutral test results were negative and significantly deviated from neutral, (2) The use of nucleotide distribution mismatch (mismatch distribution) analysis to test the existence of group expansion; unpaired distribution test and neutral hypothesis test were used Arlequin3.01 software. The differentiation time of the CO1 gene and cytochrome b gene were calculated by molecular clock theory. The evolution rate of CO1 gene was calculated by 0.5% to 1. 4%/ million years (MY), and the evolution rate of cytochrome b gene were calculated by 2.15% to 2.6% / MY, Zhao *et al.* (2013).

## Results

of 15 complete mtDNA A total of Amphioctopus fangsiao genomes were characterized (Gene Bank Accession No (MF 029678-MF 029691). The mtDNA genome sizes range from 15977 to 15990 bp, with the G+C content ranging from 22.39% to 22.93%. The mtDNA genome contains all 37 genes as typically present in mollusks, which include cytochrome c oxidase subunits I-III (COI, COII, COIII), ATPase subunit 6 and 8 (ATP6, ATP8) and NADH dehydrogenase subunits 1-6 and 4L (ND1- 6, ND4L); 22 tRNA genes, two rRNA genes. Eight of 22 tRNA genes (tRNA-Lys, tRNA-Ala, tRNA-Arg, tRNA-Asn, tRNA-Ile, tRNA-Ser, tRNA-Asp, and tRNA-Thr) were located in the light chain and the remainders were found in the other chain (Fig. 2). In the A. fangsiao mtDNAs, 1642 positions were variable and 1023 were parsimony-informative. The complete mitochondrial genomes of A. fangsiao and other related species 0. bimaculatus. such as 0 conispadiceus, A. fangsiao and O. vulgaris were compared to estimate intraspecific nucleotide diversity (Pi) and Rozas, 2009). The (Librado analysis showed that gene by gene Pi values were highly variable (Table 1). Nucleotide diversity in A. fangsiao varies from ~0.2% in ATP8 genes to 6.5% in the ATP6 genes, with highest among protein-coding genes value of Pi (~1%) detected in ATP6 gene (Table 1).



Figure 2: Neighbor-joining tree inferred from complete mitochondrial genomes of Amphioctopus fangsiao, Octopus bimaculatus, Octopus variabilis, Octopus vulgaris and Amphiotopus aegina. The representatives of other four related species were used as outgroups. The highly significant statistical supports are listed in the order ML/NJ (≥95%/N75%). Statistically supported mtDNA clades are designated by Latin letters. Scale bar indicates replacements per site. Key: SH: Shanghai, ZS: Zhoushan, XM: Xiamen, DS: Dongshan, ZJ: Zhanjiang, QD: Qingdao, NT: Nantong, YT: Yantai.

ole 1: Nucleonde diversity (PI) of midNA genes in Amphiociopus jangsia							
	Locus	Length (bp)	Position	Pi			
	CO3	780	1-780	0.02862			
	ND3	351	1115-1465	0.03305			
	ND2	1038	1533-2570	0.03035			
	CO1	1533	2542-4074	0.02865			
	CO2	687	4078-4764	0.02756			
	ATP8	156	4831-4986	0.01600			
	ATP6	705	4989-5681	0.06513			
	ND5	1719	5729-7465	0.03964			
	ND4	1344	7533-8858	0.04104			
	ND4L	237	8873-9124	0.01584			
	CYTB	1140	9314-10445	0.02484			
	ND6	513	10438-10950	0.04702			
	ND1	942	11026-11967	0.03907			

 Table 1: Nucleotide diversity (Pi) of mtDNA genes in Amphioctopus fangsiao.

There were 15 haplotypes among all the 15 mtDNA genomes. The genetic distances across the mtDNA genomes ranged from 0.007 to 0.059 (Table 2). The phylogenetic trees were generated using NJ and ML methods, where O. aegina, O. variabilis, O. vulgaris, O. bimaculoides used as outgroup (Fig. 3) it revealed two clades A (Dalian, Yantai, Qingdao, Nantong populations) and B (Shanghai, Zhoushan. Xiamen. Dongshan and Zhanjiang populations) (Table 3). The haplotype network analysis also revealed two cladded and none of the haplotype was shared (Fig. 4). The AMOVA analysis showed that 61.48% of the genetic variation existed between the two clades, while only 38.52% of the genetic variation existed within each clade. FST of the two clades reached 0.61476. The net genetic distance between the two groups was 0.030.

The molecular clocks showed that the genetic distances between the two clades of COI gene were 0.027, and the genetic distances between the two clades of Cytb gene was 0.215.The differentiation time was estimated to be about  $119 \times 104 \sim 540 \times 104$  million years ago before the Tertiary period (according to the fish and shrimp and crab's COI gene 0.5% to 1.4% per million years of evolution rate inferred (Knowlton and Weigt, 1998), the evolution rate of the cytochrome b gene was calculated by 2.15% to 2.6%/ million years).

In the present study we projected the ratio of the number of non-synonymous substitutions per non-synonymous sites (KA) to the number of synonymous substitutions per synonymous sites (KS) of (CYTB), and (COX3) gene, (NADH3) gene, (NADH2) gene, (COX1) gene, COX2) gene, ATP synthase F0 subunit 8 (ATP-8) gene, ATP synthase F0 subunit 6 (ATP-6) gene and found low KA/KS values in both clades (KA/KS =0.0126 and 0.9827 in clades A and B, respectively), representing the influence of negative selection.

Table 2: Genetic distances between pairwise populations of Amphioctopus fangsiao.									
	DL	QD	YT	NT	SH	ZS	XM	DS	ZJ
DL									
QD	0.012								
YT	0.008	0.007							
NT	0.007	0.006	0.001						
SH	0.044	0.043	0.038	0.037					
ZS	0.053	0.051	0.047	0.045	0.019				
XM	0.049	0.048	0044	0.043	0.030	0.037			
DS	0.059	0.057	0.053	0.050	0.031	0.041	0.031		
ZJ	0.054	0.052	0.048	0.046	0.029	0.037	0.031	0.026	



Figure 3: Mitochondrial genome map of Amphioctopus fangsiao.

Table 3: Geographical distri	bution of Amphioctopi	us fangsiao sam	oles used in this study
	The second se	J	

Sequence ID	Site ID	Province	Country	Clade	GenBank Accession No
DL	DL Dalian Liaoning		China	А	MF029678
QD	D Qingdao Shandong		China	А	MF029684
YT	Yantai	Shandong	China	А	MF029690
NT1	Nantong	Jiangsu	China	А	MF029680
NT19	Nantong	Jiangsu	China	А	MF029681
NT21	Nantong	Jiangsu	China	А	MF029682
NT26	Nantong	Jiangsu	China	А	MF029683
SH1	Shanghai	Shanghai	China	В	MF029685
SH21	Shanghai	Shanghai	China	В	MF029686
SH22	Shanghai	Shanghai	China	В	MF029687
SH23	Shanghai	Shanghai	China	В	MF029688
ZS	Zhoushan	Zhejiang	China	В	MF029692
XM	Xiamen	Fujian	China	В	MF029689
DS	Dongshan	Fujian	China	В	MF029679
ZJ	Zhanjiang	Guangdong	China	В	MF029691



Figure 4: Median-joining network of mtDNA haplotypes observed in 15 *Amphioctopus fangsiao* specimens. This network illustrates the relationships between two major clades A and B. Median vectors are shown by red dots.

## Discussion

Amphioctopus fangsiao is a potential species for aquaculture (Wang et al., 2015; Feng et al., 2017) and overexploitation causes its population decline (Zhang et al., 2009). Therefore, basic information of phylogeny and population history counts valuable. A few investigations are in account for its population genetics, by utilizing AFLP, microsatellite and Allozyme (Gao et al., 2002, Zhang et al., 2009, and Feng et al., 2017) and detailed genetic structure (Muhammad et al., 2019). These earlier investigations excluding Muhammad et al. (2019) are limited to northern part of the coast and do not provide complete profile of population genetic of this economically important species. However, our previous prediction of population differentiations of Α. present fangsiao be validated by investigation (Lü et al., 2010, 2011). Here, we are describing the phylogeny

and population history of nine populations of A. fangsiao (Fig. 1) using complete mtDNA from 15 individuals. Molecular markers separated north (Dalian, Nantong, Qingdao, Yantai) and south populations (Huizhou. Shanghai, Xiamen and Zhenjiang), which is in accordance with our previous results of CO1 and 16S rDNA (Lü et al., 2010, 2011), and differ with AFLP results where northern populations Dalian, Qingdao were placed in one clade while Yantai and Lianyungang were reported to be another clade. This pattern of separation is like Octopus minor populations where south and north populations are distinctly distributed into two clades (Lü et al., 2013).

The genetic distance between the two clades was 0.030, and the genetic differentiation index  $F_{ST}$  reached 0.61476. The higher  $F_{ST}$  values illustrate lower level of gene flow (N<sub>m</sub>)

genetic differentiation and higher among populations (Hedrick 2005; Ye et al., 2015). The standard values of  $F_{ST}$ are illustrated as 0.05 defines negligible genetic differentiation while greater than 0.25 defines high genetic analyzed differentiation within the population (Weir and Cockerham, 1996). Based on given standard obtained results showed greater differentiation among fangsiao Α. populations. The higher genetic diversity was also reported in other species of the area, such as; genetic diversity between seven populations of Octopus variabilis was (0.91) based on the 12S rRNA and COIII gene (Xu et al., 2011). Kang et al. (2012) reported variations in Octopus minor populations of different localities ranging from (0.109 to 0.999). The genetic divergence of Maoricolpus roseus, found less than (1%) using mtDNA 16S and CO1 gene, Kirsten et al. (2015). Muhammad et al. (2018) investigated eight populations ranged (0.011 to 0.992). Comparison to said studies, present investigation showed moderate level of differentiation. There are two concepts to illustrate the genetic variations in A. fangsiao populations. It might be due to life style such as lack of planktonic larval stages and limited migration range (Lv et al., 2013). These factors might limit the dispersal ability and resulting reduction of gene flow. Nevertheless, this explanation does not support in case of clade A where no signification differentiation found within the group. The probable reasons

for genetic differentiation are results of various factors including geographic isolation. current. life history characteristics, (Gao et al., 2016), Islands and Gulfs also contribute in gene flow complications, the early glacier activities, where sea level climatic fluctuations encountered during the Pleistocene period and caused gene flow restriction in marine organisms (Imbrie et al., 1992). Besides it. the Changjiang River might influence in genetic structure of A. fangsiao populations along China coast (Lü et al., 2011). It is generally believed that the marine hydrological conditions. the ecological characteristics of the species and the life history can cause the differentiation of marine life (Muss et al., 2001; Yong et al., 2009). An intriguing possibility is that this type of environmentally driven genetic structure in Ocean species promotes allopatric speciation, whereby genetically different population glacial divergence during cvcles (France and Kocher 1996, Zardus et al., 2006, Schüller 2011). A number of divergent forces effect genetic variation, including geographic separation, existing and life history features (Gao et al., 2016).

Estimation of the differentiation time of the octopus group using different molecular clocks showed that the genetic distances between the two clades of COI gene was 0.027, and the genetic distances between the two clades of Cytb gene was 0.215, so the differentiation time was estimated to be about  $119 \times 104 \sim 540 \times 104$  million years ago (MYA) before the Tertiary period (according to the fish and shrimp and crabs' COI gene 0.5% to 1.4%/ MYA of evolution rate inferred (Knowlton and Weigt, 1998), the evolution rate of cytochrome b gene was calculated by 2.15% to 2.6%/ MYA), corresponding to the early Pleistocene period the genetic differentiation between both populations has been attributed to the repeated Pleistocene glaciations (1.25-0.7 MYA), (Mark *et al.*, 2005, Clark *et al.*, 2006).

There were 15 haplotypes among all the 15 mitochondrial DNA genomes. The genetic distances across the mtDNA genomes ranged from 0.007 between NT and YT, to 0.059 between north-south region (Dongshan located in the south of China and Dalian located in the north of China), with an average genetic distance of 0.057. To test whether there is a signature selection in the Α. fangsiao mitochondrial genome, we assessed the ratio of the number of non-synonymous substitutions per non-synonymous sites (KA) to the number of synonymous substitutions per synonymous sites (KS) and found low KA/KS values in both clades (KA/KS = 0.0126 and 0.9827 in clades А and B. respectively), representing the influence of negative selection. Negative selection plays an important role in maintaining the longterm stability of biological structures by removing deleterious mutations, Ana M Pérez O'Brien et al. (2014). However,

further studies are required to confirm differentiation of *A. fangsiao*.

## Conclusion

Among populations main divergence factor is the ice age quaternary glacial period. Yangtze River acts as a physical barrier to larval dispersal and variation between both populations. This lays the foundation for the future development and utilization of the octopus resources in China.

# Acknowledgements

This research was supported by the National Natural Science Foundation of China (NSFC) (41576131)and Talented Yong Scientist Program (PAK-18-024). We pay thank to whose reviewers constructive comments helped us to improve this manuscript.

# Referenced

- Akasaki, T., Nikaido, M., Tsuchiya, K., Segawa, S., Hasegawa, M. and Okada, N., 2006. Extensive mitochondrial gene arrangements in coleoid Cephalopoda and their phylogenetic implications. Molecular **Phylogenetics** & 648. Evolution, 3. https://doi.org/10.1016/j.ympev.200 5.10.018T
- AnaMPérezO'Brien,A.M.P.,Utsunomiya,Y.T.,Mészáros,G., BickhartD.M.,<sup>4</sup>LiuG.E.,VanTassellC.P., Sonstegard,T.S., DaSilva,M.V.B,Garcia J.F. and Sölkner J.,

**2014.** Assessing signatures of selection through variation in linkage disequilibrium between taurine and indicine cattle. *Genetic Selection Evolution*, 1, 19. Doi: 10.1186/1297-9686-46-19.

- Bandelt, H., Forster, P. and Rohl, A., 1999. Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology Evolution*, 16, 37-48.
- Bohonak, A.J., 1999. Dispersal, Gene Flow, and Population Structure. *The Quarterly Review of Biology*, 74, 21-45.
- Cheng, R., Zheng, X., Lin, X., Yang, J. and Li, Q., 2012. Determination of the complete mitochondrial DNA sequence of Octopus minor. *Molecular Biology Reports*, 4, 3461-3470.

https://doi.org/10.3109/19401736.20 14.928866

Clark, P., Archer, D., Pollard, D., Joel, D., Rial, J., Brovkin, V., Alan, C. and Martin, N.G., 2006. The middle Pleistocene transition: characteristics, mechanisms and implications for long-term changes in atmospheric pCO<sub>2</sub>. *Quaternary Science Reviews. Elsevier*, 23-24, 3150-3184.

https://doi.org/10.1016/j.quascirev.2 006.07.008

De Luca, D., Catanese, G., Fiorito, G. and Procaccini, G., 2015. A new set of pure microsatellite loci in the common octopus *Octopus vulgaris* Cuvier, 1797 for multiplex PCR assay and their cross-amplification in O. maya Voss & Solis Ramirez, 1966. *Conservation Genetics Resources*, 1, 299–301.

- De Luca, S., Ziko, I., Sominsky, L., Nguyen, J.C.D., Dinan T., Miller, A.A. and Jenkins T.A. and Spencer S.J., 2016. Early life overfeeding impairs spatial memory performance by reducing microglial sensitivity to learning. *Journal of Neuro inflammation*, 13, 112.
- Domínguez, J., Munguiavega, A., Ceballosvázquez, B., Garcíarodriguez, F. and Arellanomartinez, M., 2015. The complete mitochondrial genome of Verrill, 1883 from the Gulf of California. Mitochondrial DNA 1-2.
- **Dong, Z., 1988.** Fauna Sinica: Phylum Mollusca. Class Cephalopoda. Science Press, Beijing, 174-176.
- Feng, Y., Liu, W., Xin, X., Yang, J., Wang, W., Wei, X., Liu, X. and Sun, G., 2017. Construction of a normalized full-length **cDNA** Library of Cephalopod Amphioctopus fangsiao and development of microsatellite Journal markers. of Ocean University of China, 5, 897–904. https://link.springer.com/article/10.1 007/s11802-017-3291-y
- Fernandez, A., John, B., Julia, S. and Wellner, B.H., 2011. Timescale dependence of glacial erosion rates: A case study of Marinelli Glacier, Cordillera Darwin, southern Patagonia. *Journal of Geophysical Research*, 116, F01020.

- France, S. and Kocher, T., 1996. Geographic and bathymetric patterns of mitochondria116S rRNA sequence divergence among deepsea amphipods, *Eurythenes gryllus*. *Marine Biology*, 126, 633. https://doi.org/10.1007/BF003 51330
- Gao, Q., Wang, Z., Wang, R. and Zheng, X., 2002. Allozyme variation in five populations of Octopus ocellatus. *Transaction of Oceanology & Limnology*, 4, 46-51. https://europepmc.org/abstract/cba/3 78970
- Gao, X., Zheng, X., Bo, Q. and Li, Q., 2016. Population genetics of the common long-armed Octopus minor Octopus (Sasaki, 1920) (Cephalopoda: Octopoda) in Chinese waters microsatellite based on analysis. Biochemical Systematics Ecology, 129-136. and 66, https://doi.org/10.1016/j.bse.2016.03 .014
- Gow, J., Noble, L., Rollinson, D., Mimpfoundi, R. and Jones, C., 2004. Breeding system and demography shape population genetic structure across ecological and climatic zones in the African freshwater snail, Bulinus forskalii (Gastropoda, Pulmonata), intermediate host for schistosomes. *Molecular Ecology*, 13, 3561–3573. https://doi.org/10.1111/ j.1365-294X.2004.02339.x
- Hall, T., 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows

95/98/NT. Nucleic Acids Symposium, 41, 95-98.

- Hao, Z., Liu, Y., Nazaire, M., Wei, X. and Wang, X., 2015. Molecular phylogenetics evolutionary and history of sect. Quinquefoliae (Pinus): implications for Northern Hemisphere biogeography Molecular *Phylogenetics* and Evolution, 87, 65–79. https://doi.org/10.1016/j.ympev.201 5.03.013
- Harrison, R., 1991. Molecular changes at speciation. Annual Review of Ecology and Systematics, 22, 281– 308.

https://doi.org/10.1146/annurev.es.2 2.110191.001433

Hedrick, P. and Goodnight, C., 2005. A standardized genetic differentiation measure. *Evolution*, 8, 1633–8. https://doi.org/ 10.1111/j.0014-

3820.2005.tb01814.x

- Hellberg, M., Ronald, S. Burton, J., Neigel, E. and Palumbi, S.R., 2002. Genetic assessment of connectivity among marine populations. *Bulletin Marine Science*, 70, 273-290.
- Husemann, M,. Ray, J., King, R., Hooser, E. and Daneley, P., 2012. Comparative biogeography reveals differences in population genetic structure of five species of stream fishes. *Biological Journal of the Linnean Society*, 107, 867–885 https://doi.org/10.1111/ j.1095-8312.2012.01973.x
- Imbrie, J., Boyle, E.A., Clemens, S.C., Duffy, A., Howard, W.R., Kukla,

G., Kutzbach, J., Martinson, D.G., McIntyre, A., Mix, A.C., Molfino, B., Morley, J.J., Peterson, L.C., Pisias, N.G., Prell, W.L., Raymo, Shackleton, N.J. **M.E.** and Toggweiler, J.R., 1992. On the structure and origin of major cycles. glaciations I. Linear responses to Milankovitch forcing. 7. Paleoceanography, 701-738. https://doi.org/10.1029/92PA02253

- Kang, J., Park, J. and Choi, T., 2012. Genetic differentiation of octopuses from different habitats near the Korean Peninsula and eastern China based on analysis of the mDNA cytochrome C oxidase 1 gene. *Genetic and Molecular Research*, 4, 3988–3997.
- Kim, J.I., Oh, T.Y., Seo, Y.I. and Cho, E.S., 2009. Population genetic structure of Octopus minor Sasaki from Korea and China based on a partial sequencing of mitochondrial 16S rRNA. *Korean Journal of Life Science*, 19, 711-719.
- Kirsten, M. and Hamish, G., 2015. New Zealand screw shells *Maoricolpus roseus* (Gastropoda: Turritellidae): two species, two subspecies or a single variable species. *Molluscan Research*, 2, 123-127.

https://doi.org/10.1080/13235818.20 14.977838

Knowlton, N. and Weigt, L., 1998. New dates and new rates for divergence across the Isthmus of Panama. Proc R Soc Lond, B, 265, 2257–2263.

- Librado, P. and Rozas, J., 2009. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics*, 11, 1451-1452.
- Lü, Z., Li, H., Wu, C., Fan, Z. and Zhang, J., 2010. Genetic variation of Octopus ocellatus populations in China's coastal waters based on the COI gene analysis. Acta Oceanologica Sinica, 1, 130–138. (In Chinese with English abstract).
- Lü, Z., Li, H., Wu, C.W., Fan, Z.J. and Zhang, J.S., 2011. Population genetics of Octopus ocellatus in coastal waters of China based on 16S rDNA sequence. Journal of Fishery Sciences of China, 18 (1), 29-37. https://10.3724/SP.J.1118.2011.00029
- Lü, C., Zheng, X. and Lin, X., 2012.
  Diversity of Cephalopoda from the waters of the Chinese mainland and Taiwan. In: Proceeding of the 1st Mainland and Taiwan Symposium on Marine Biodiversity Studies. Ocean Press, Beijing, 76-87.
- Lü, Z.M., Liu, L.Q., Li, H., Wu, C.N. and Zhang, J.S., 2013. Deep phylogeographic break among Octopus variabilis populations in China: Evidence from mitochondrial and nuclear DNA analyses. Biochemial Systematics and Ecology, 51, 224–231.

https://doi.org/10.1016/j.bse.2013.09 .003

Lv, Z., Li, H., Wu, C., Fan, Z. and Zhang, J., 2013. Population genetic structure of *Octopus ocellatus* in coastal waters of China based on 16S rDNA sequence. *Journal of Fishery Sciences of China*, 1, 29-37. Doi:10.3897/zookeys.775.24258

- Maltagliati, F., 2002. Genetic brackish-water monitoring of populations: the Mediterranean tooth carp *Aphanius* fasciatus (Cyprinodontidae) as a model. Marine Ecology Progress Series, 235, 257-262.
- Mark, A., Maslin, A. and Ridgwell, J., 2005. Mid-Pleistocene revolution and the 'eccentricity myth', Geological Society, London, Special Publications, 247, 19-34.
- Melis, R., Vacca, L., Cuccu, D., Mereu, M., Cau, A., Follesa, M. and Cannas, R., 2018. Genetic population structure and phylogeny of the common octopus *Octopus vulgaris* Cuvier, 1797 in the Western Mediterranean Sea through nuclear and mitochondrial markers. *Hydrobiologia*, 807, 277–296.
- Muhammad, F., Liu, L., Lü, Z., Gong,
  L., Du, X., Shafi, M., Waryani, B.
  and Kaleri, H., 2018. Genetic
  diversity of Octopus minor
  (SASAKI, 1920) inferred by
  mitochondrial NADH Dehydrogenase
  Subunit 2 gene. *The Journal of*Animal & Plant Sciences, 5, 1364-1371.

http://www.thejaps.org.pk/ docs/v-28-05/16.pdf

Muhammad, F., Chen, W., Liu, L.,
Gong, L., Du, X., Shafi, M. and Lü,
Z.M., 2019. Genetic structure of *Amphioctopus fangsiao* (Mollusca,
Cephalopoda) in Chinese waters

inferred from variation in three mtDNA genes (ATPase 6, ND2, and ND5). *Hydrobiologia*, 1, 111–119. https://doi.org/10.1007/s10750-019-03981-9

- Muss, A., Robertson, D., Stepien, C., Wirtz, P. and Bowen, B., 2001. Phylogeography of Ophioblennius: the role of ocean currents and geography in reef fish evolution. *Evolution*, 3, 561-572. https://doi.org/10.1111/ j.0014-3820.2001.tb00789.x
- Oosthuizen, A., Meesbah, J. and Shaw, P., 2004. Genetic analysis of the Octopus vulgaris population on the coast of South Africa. South African Journal of Science, 100.
- Ortego, J., Noguerales, V., Gugger, P. and Sork, V., 2015. Evolutionary and demographic history of the Californian scrub white oak species complex: an integrative approach. *Molecular Ecolology*, 24, 6188– 6208.

https://doi.org/10.1111/mec.13457

- Schüller, M., 2011. Evidence for a role of bathymetry and emergence in speciation in the genus Glycera (Glyceridae, Polychaeta) from the deep Eastern Weddell Sea. *Polar Biology*, 34, 549-564, Doi: 10.1007/s00300-010-0913-x
- **Slatkin, M., 1993.** Isolation by distance in equilibrium and non-equilibrium populations. *Evolution*, 47, 264-279.
- Thompson, J., Gibson, T., Plewniak,F., Jeanmougin, F. and Higgins, D.,1997. The ClustalX windows interface: flexible strategies for

multiple sequence alignment aided by quality analysis tools. Nucleic Acids Res. *Nucleic Acids Research*, 25, 4876-4882.

- Vellend, M. and Geber, M., 2005. Connections between species diversity and genetic diversity. *Ecology Letters*, 8, 767–781. https://doi.org/10.1111/ j.1461-0248.2005.00775.x
- Wang, W., Dong, G., Yang, J., Zheng, X., Wei, X. and Sun, G., 2015. The development process and seasonal changes of the gonad in *Octopus* ocellatus Gray off the coast of Qingdao, Northeast China. Fisheries Science, 81, 309-319. https://doi.org/10.1007/s12562-014-

0846-3.

- Weir, B.S. and Cockerham, C.C., 1996. Genetic data analysis II: Methods for discrete population genetic data. Sinauer Associates Inc., Sunderland, MA.
- Wethington, A., Zavodna, M., Smith, M., Oliveira, G., Lewis, F. and Minchella, D., 2007. Population genetic structure of *Biomphalaria* glabrata in a schistosomiasisendemic region in Brazil. Journal of Molluscan Studies, 73, 45–52. https://doi.org/10.1093/mollus/ey1028
- Xu, M., Li, B., Guo, Z., Lu, C. and Wu, Z.C. 2011. Genetic diversity of seven populations of Octopus variabilis in China's coastal waters based on the 12S rRNA and COIII gene analysis. Oceanologia et Limnologia Sinica, 42, 387–396.

- Xu, X., Wu, X. and Yu, Z., 2012. Comparative studies of the complete mitochondrial genomes of four Paphia clams and reconsideration of subgenus Neotapes (Bivalvia: Veneridae). *Gene*, 1, 17–23. https://doi.org/10.1016/j.gene.2011.1 2.002
- Ye, Y., Wu, C. and Li, J., 2015. Genetic population structure of Macridiscus multifarious (Mollusca: **Bivalvia**) on the basis of mitochondrial markers: strong population structure in a species with a short planktonic larval stage. PloS ONE, 10(12), e 0146260. https://doi.org/10.1371/journal.pone. 0146260
- Yokobori, S., Fukuda, N., Nakamura, M., Aoyama, T. and Oshima, T., 2004. Long-Term Conservation of Six Duplicated Structural Genes in Cephalopod Mitochondrial Genomes. *Molecular Biology & Evolution*, 11, 2034-2046.

https://doi.org/10.1093/molbev/msh22

- Yong, L., Liu, R., Ye, L., Liang, J., Xuan, F. and Xu, Q., 2009. Genetic differentiation between populations of swimming crab *Portunus trituberculatus* along the coastal waters of the East China Sea. *Hydrobiologia*, 1, 125-137. Doi: 10.1007/s10750-008-9570-2
- Zardus, J., Etter, R., Chase, M., Rex,M. and Boyle, E., 2006.Bathymetric and geographicpopulation structure in the pan-Atlantic deep-sea bivalveDeminucula atacellana (Schenck,

1939). *Molecular Ecolology*, 15, 639-651. Doi:10.1111/j.1365-294X.2005.0282.x

Zhang, L., Yang, J., Liu, X., Wang, W., Sun, G., Liu, L. and Zheng, X., 2009. The genetic diversity of Octopus ocellatus by AFLP markers. Oceanologia Et Limnologia Sinica, 6, 803-807.

https://www.cabdirect.org/cabdirect/ abstract/20103104993

Zhang, X., Zheng, X., Ma, Y. and Li, Q., 2015. Complete mitochondrial genome and phylogenetic relationship analyses of *Amphioctopus aegina* (Gray, 1849) (Cephalopoda: Octopodidae). *Mitochondrial DNA*, 6-1. https://doi.org/10.3109/19401736.20 15.1106522

- Zhao, Y., Zhang, Y. and Li, X., 2013.
  Molecular Phylogeography and Population Genetic Structure of an Endangered Species *Pachyhynobius shangchengensis* (hynobiid Salamander) in a Fragmented Habitat of Southeastern China. *Plos One*, 8(10), e78064.
- Zid, F., Knittweis, L., Aurelle, D., Nafkha, Ch., Ezzeddine, S., Fiorentino, F., Ghmati, H., Ceriola, L., Jarboui, O. and Maltagliati, F., 2012. Genetic structure of Octopus vulgaris (Cephalopoda, Octopodidae) in the central Mediterranean Sea inferred from the mitochondrial COIII gene. C. R. Biologies, 335, 625–636.