

Bioinformatic analysis of important miRNA and Gene network analysis of APC gene, a key regulator of head and neck cancer

¹N Vishal Prakasam, ²Sathish Sankar*, ³S Smiline Girija

¹Department of Microbiology, Saveetha Dental College and Hospitals Saveetha Institute of Medical and Technical Sciences Saveetha University, Chennai-600 077, India

^{2*}Professor, Department of microbiology Saveetha Dental College and Hospitals Saveetha Institute of Medical and Technical Sciences Saveetha University, Chennai-600 077, India

Introduction:-

Every individual in this world is cursed with getting cancer and this curse is dependent on a structure called the genome. A genome is all genetic information of an organism which consists of a nucleotide sequence of DNA (or RNA in RNA virus) A nuclear genome includes protein-coding genes and non-coding genes, the other functional regions of the genome (Noncoding DNA), and any junk DNA. mutations to these nucleotide sequences leads to an abnormality in production of protein and leads to cancer, one such cancer is cancer that arises in the head and neck. Head and neck cancer is the most common type of cancer and is ranked seventh worldwide . These types of cancers comprise a diverse group of tumors affecting the upper aerodigestive tract (1) and comprise a heterogeneous group of cancers, most common type is squamous cell carcinoma (SCC), which arise from the epithelial lining of the oral cavity, sinonasal tract, pharynx, larynx and salivary glands etc. Most HNC are already at an advanced

stage when it is diagnosed, which significantly reduces the survival rate, even after curative treatment. A Major risk factor includes tobacco smoking or chewing, excessive consumption of alcohol, paan masala (Gutkha), gamma and ultraviolet radiation, overexposure to sunlight, a family history of cancer and increasing age. Total incidence in the globe continues to rise with more than million cases annually and more than 10000 new cases in India every year, an increase of 20% in the last decade. However prognosis remains poor, with a 28-67% chance of survival at five years, depending upon the stage at presentation of the case.

APC is a Biallelic mutation of the *APC* gene that occurs in 45%–80% of colorectal cancers (2–4)and is observed in the earliest detectable lesions (5,6). The *APC* locus was originally identified based on its link to familial adenomatous polyposis coli (FAP), an inherited syndrome of cancer predisposition (7–9). Inherited mutations in the *APC* gene cause affected individuals to

develop hundreds to thousands of adenomatous polyps, resulting in the onset of CRC typically before the age of 40 (10)Individuals with FAP inherit a loss-offunction mutation in a single allele of APC, followed by an additional acquired mutation in the second allele of APC in the adenomas and adenocarcinomas that develop (11). Thus, the acquisition of biallelic APC mutations represents an early and rate-limiting step in all FAP-associated and most sporadic colorectal tumors.In studies of colorectal cancer as a whole, APC mutational status does not strongly correlate with outcome (12)Nevertheless, APC mutations exhibit an interesting pattern of differential distribution in the recognized subtypes of colorectal cancer. APC mutations correlate strongly with a large subset of colorectal cancers associated with intermediate prognosis.On the other hand, APC mutations occur infrequently within a smaller subset derived from sessile serrated adenomas and associated with microsatellite instability and good prognosis. This latter subset exhibits a relatively high proportion of activating mutations in the gene encoding β -catenin (*CTNNB1*) that are mutually exclusive of APC mutations(13). Interestingly, CTNNB1 mutations are significantly more prevalent in small adenomas than in large adenomas or adenocarcinomas, whereas APC mutations are well-represented across all stages of tumorigenesis. It has recently emerged that APC mutational status has value as a predictive marker of poor prognosis in Stage III colorectal cancer raising the possibility that APC mutations not only initiate colorectal cancer development, but drive clinical phenotypes relevant to progression and metastasis as well.

Materials and Methods

Target scan prediction Prediction of microRNA targets for APC gene in humans was carried out using Targetscan Human software program. Targetscan is a database that allows the user to filter miRNAs and its targets that are associated with determined diseases, and we filtered the results that were associated with GC. MiRTarBase is a database that predicts targets that have already been validated by molecular biology techniques. Broadly conserved, conserved and poorly conserved mRNA families were searched for the presence of 8mer, 7mer, and 6mer sites matching each miRNA seed region. Predicted regulatory targets of the TP53 gene

were identified using the program with default settings. The targets of the conserved miRNA. hsa-miR-122-5p miRNA with a miTG score of ≥ 0.99 were identified using the Dianalab software program. Gene network analysis of APC was carried out by STRING database online server program. Important gene interactions with a combined score of ≥ 0.99 were considered and listed.

miRDB prediction

Target mRNAs were predicted using miRDB online server program. miRNA targets with a target score of more than 95 were considered for further analysis. The target details and the predicted genes for the miRNA hsa- miR-3922-5p were carried out.(14–17)

GenenetworkanalysisGene network analysis of APC was carriedout by STRING database online serverprogram. This software was used in order todetermine the various links that APC haswith other genes. This database allows the

user to filter out the desired gene links for the user's benefits.

MiRDB prediction for APC gene. Gene 324 is predicted to be targeted by 184 miRNAs in miRDB. Selected target score for the following data from MiRDB is >95

Target Rank	Target Score	miRNA Name	MiRNA sequence	Important genes
1	99	<u>hsa-miR-</u> <u>3120-3p</u>	5' - cacagcaaguguagacaggca - 3'	PXYLP1, SCAF4, TGIF2, APBB2. , ETV5
2	99	<u>hsa-miR-</u> <u>561-3p</u>	5' - caaaguuuaagauccuugaagu - 3'	ARIH1, MED14, ACTR2, ACVR2A, C16orf72
3	99	<u>hsa-miR-</u> <u>6853-3p</u>	5' - uguucauuggaacccugcgcag - 3'	GRIK2, APC, CA13, DIP2C, PARP12
4	98	<u>hsa-miR-</u> 5696	5' - cucauuuaaguagucugaugcc - 3'	AHCTF1, DNAJB14, TENT5A, APIP, RNFT1
5	97	<u>hsa-miR-</u> 6504-3p	5' - cauuacagcacagccauucu - 3'	MS12, SCAMP1, ARIH1, CDC73, ATAD2B
6	97	<u>hsa-miR-</u> 3942-5p	5' - aagcaauacuguuaccugaaau - 3'	SLBP, CXCL16, BHMT, CDK6, RB1
7	97	<u>hsa-miR-</u> <u>582-5p</u>	5' - uuacaguuguucaaccaguuacu - 3'	GABPB1, PRDM12, ABTB2, TNKS2, BCL11B
8	97	<u>hsa-miR-</u> 10399-5p	5' - aauuacagauugucucagaga - 3'	MS12, ARIH1, CDC73, ATAD2B, TBC1D19
9	97	<u>hsa-miR-</u> 4703-5p	5' - <mark>uagcaauacaguacaaauauagu</mark> - 3'	BHMT, SLBP, CXCL16, OTOGL, TRIM43B
10	96	<u>hsa-miR-</u> 153-3p	5' - uugcauagucacaaaagugauc - 3'	UNC5C, KCNQ4, SERTAD2, KLF5, HEY2

Results:-

Sathish Sankar.et.al., Bioinformatic analysis of important miRNA and Gene network analysis of APC gene, a key regulator of head and neck cancer

11	95	<u>hsa-miR-</u> 4282	5' - uaaaauuugcauccagga - 3'	TMOD2, C21orf91, ZDHHC21, RPS6KA6, RBM27
12	95	<u>hsa-miR-</u> <u>30e-3p</u>	5' - cuuucagucggauguuuacagc - 3'	CDC73, ZEB2, NUFIP2, PCLO, LRRTM2

MiRNA (hsa-miR-3120-3p)with a target score of 99 is further detailed.

Mature miRNA:

miRNA Name: hsa-miR-3120-3p. miRNA Sequence: 5' - cacagcaaguguagacaggca - 3' (length = 21).

Precursor miRNA:

Precursor Name: hsa-mir-3120.

There are 1253 predicted targets for hsamiR-3120-3p in miRDB out of which the selected target score for selected miRNA is >=98

Target Rank	Target Score	miRNA Name
1	100	hsa-miR-3120-3p
2	99	hsa-miR-3120-3p
3	99	hsa-miR-3120-3p
4	99	hsa-miR-3120-3p
5	99	hsa-miR-3120-3p
6	99	hsa-miR-3120-3p
7	99	hsa-miR-3120-3p
8	99	hsa-miR-3120-3p
9	99	hsa-miR-3120-3p
10	98	hsa-miR-3120-3p
11	98	hsa-miR-3120-3p
12	98	hsa-miR-3120-3p
13	98	hsa-miR-3120-3p

14	98	hsa-miR-3120-3p
15	98	hsa-miR-3120-3p





ROB01, APBB2, PXYLP1, ARIH1, GRIK2 were some of the genes found to have interactions of miRNA associated with APC gene.

Figure1: The following image represents the gene network analysis of APC gene. Which helps us in understanding the various links that APC have towards other genes.

Link between APC and other genes

Adenomatous polyposis coli protein; Tumor suppressor. Promotes rapid degradation of CTNNB1 and participates in Wnt signaling as a negative regulator. APC activity is correlated with its phosphorylation state. Activates the GEF activity of SPATA13 and ARHGEF4. Plays

a role in hepatocyte growth factor (HGF)induced cell migration. Required for MMP9 up-regulation via the JNK signaling pathway in colorectal tumor cells. Acts as a mediator of ERBB2-dependent stabilization of microtubules at the cell cortex.

DLH 1

Disks large homolog 1; Essential multidomain scaffolding protein required for normal development (By similarity). Recruits channels, receptors and signaling molecules to discrete plasma membrane domains in polarized cells. May play a role in adherens junction assembly, signal transduction. cell proliferation, synaptogenesis and lymphocyte activation. Regulates the excitability of cardiac

2023

myocytes by modulating the functional expression of Kv4 channels. Functional regulator of Kv1.5 channel; Belongs to the MAGUK family.

CTNNB1

beta-1; Key Catenin downstream component of the canonical Wnt signaling pathway. In the absence of Wnt, forms a complex with AXIN1, AXIN2, APC, CSNK1A1 and GSK3B that promotes phosphorylation on N-terminal Ser and Thr residues and ubiquitination of CTNNB1 via BTRC and its subsequent degradation by the proteasome. In the presence of Wnt ligand, CTNNB1 is not ubiquitinated and accumulates in the nucleus, where it acts as a coactivator for transcription factors of the TCF/LEF family, leading to activate Wnt responsive genes.

CSNK1A1

Casein kinase I isoform alpha; Casein kinases are operationally defined by their preferential utilization of acidic proteins such as caseins as substrates. It can phosphorylate a large number of proteins. Participates in Wnt signaling. Phosphorylates CTNNB1 at 'Ser-45'. May phosphorylate PER1 and PER2. May play a role in segregating chromosomes during mitosis. May play a role in keratin cytoskeleton disassembly and thereby, it may regulate epithelial cell migration.

ARHGEF4

Rho guanine nucleotide exchange factor 4; Acts as guanine nucleotide exchange factor (GEF) for RHOA, RAC1 and CDC42 GTPases. Binding of APC may activate RAC1 GEF activity. The APC-ARHGEF4 complex seems to be involved in cell migration as well as in E-cadherinmediated cell-cell adhesion. Required for MMP9 up-regulation via the JNK signaling pathway in colorectal tumor cells. Involved in tumor angiogenesis and may play a role in intestinal adenoma formation and tumor progression.

GSK3B

Glycogen synthase kinase-3 beta; Constitutively active protein kinase that acts as a negative regulator in the hormonal control of glucose homeostasis, Wnt signaling and regulation of transcription factors and microtubules. by phosphorylating and inactivating glycogen synthase (GYS1 or GYS2), EIF2B, CTNNB1/beta-catenin, APC, AXIN1, DPYSL2/CRMP2, JUN, NFATC1/NFATC. MAPT/TAU and MACF1. Requires primed phosphorylation of the majority of its substrates.

AXIN1

Axin-1; Component of the beta-catenin destruction complex required for regulating CTNNB1 levels through phosphorylation and ubiquitination, and modulating Wntsignaling. Controls dorsoventral patterning via two opposing effects; down-regulates CTNNB1 to inhibit the Wnt signaling pathway and ventralized embryos, but also dorsalizes embryos by activating a Wntindependent JNK signaling pathway. In Wnt signaling, probably facilitates the phosphorylation of CTNNB1 and APC by GSK3B. Likely to function as a tumor suppressor gene.

AXIN2

Axin-2; Inhibitor of the Wnt signaling pathway. Down-regulates beta-catenin. Probably facilitate the phosphorylation of beta- catenin and APC by GSK3B.

Discussion:

Various gene links have been identified from our study in relation to the APC gene . The most common cancer associated with APC was observed to be colorectal cancer however since genetic information gets transferred over each multiplication. There is a possibility that APC will be associated with head and neck cancers as well. It has been shown that the location of germline mutations in the APC gene is the most striking source of variability affecting the number of polyps that will develop in the colon of FAP patients (18). Other researchers has also found a correlation between the severity of the disease in FAP patients who donated their affected embryos for our FAP-hESC lines and the 3D structure of their derived colon organoids and the location of their germline APC mutations(19). It was previously suggested that a stable truncated APC protein may act in a dominant negative fashion to inactivate APC transcribed from the WT allele. Homodimerization of APC at the amino-terminus implies a possible dominant negative mode of action for mutant APC in heterozygous cells, in which shorter proteins can functionally inactivate the full-length WT protein(19–21) The first 170 amino acids are sufficient for APC homodimerization in-vitro, an association which requires the 45 amino acids only. FAP3 carries a heterozygous frameshift mutation that is predicted to be in amino acid 60, presumably resulting in lack of function of the mutated allele ('first hit'), but the normal allele expresses the APC since the 'second hit' didn't occur yet. Consequently, in the absence of a gain-offunction effect, the patient presents only mild disease, and the corresponding complexed organoids derived in vitro are similar to those of WT hESCs expressing

the two normal alleles of the APC. Therefore, we hypothesis that while the APC germline mutations in FAP1 and FAP2 may result in a truncated protein that can cause a dominant-negative effect on the normal allele, the germline mutated protein translated in FAP3 is likely too short to interfere with the normal APC protein.

In recent years, gene expression, as well as other molecular profiling, has been used as biological tools for diagnosis of cancer (22) These molecular approaches also have the potential to explore molecular mechanisms of the disease and to pave the road to fighting targeted cancer drugs and advanced treatments (23). APC is one the most important elements of the Wntsignaling pathway; its activation is a general characteristic of solid tumors such as bladder, prostate, and renal tumors. Epigenetic down regulation of Wnt pathway inhibitors may contribute to aberrant activation of the Wnt signaling pathway (24). Hypermethylation of APC promoter can silence gene expression by interfering with the binding of transcription factors to the promoter

In a previous study OCD cases reported herein showed pathogenic mutations in *CTNNB1* and *APC* genes and strong β catenin accumulation in the cytoplasm and in the nuclei, consistent with Wnt-signaling activation. The molecular profile of OCD may help in its diagnosis, as well as in the identification of potential molecular targets for therapy in the future.

In previous research Loss of heterozygosity at APC and MCC genes was investigated in 57 and 40 unrelated primary oral leukoplakia (a precancerous lesion) and squamous cell carcinomas (SCC), respectively, by polymerase chain reaction (25-34). In these samples, most of the leukoplakia patients had a tobacco smoking habit whereas the majority of cancer patients had tobacco chewing habit. LOH at APC gene was observed in 4 of 16 (25%) and 1 of 29 (3%) informative tumor and leukoplakia DNAs from tobacco chewers, respectively. LOH at MCC gene was not detected either in tumor or in leukoplakia DNAs. This infrequent LOH at APC gene of pre-cancer and cancer tissues suggests that it may not be an early event in oral carcinogenesis in these patients.

Conclusion:

In conclusion the Functional gene - miRNA interaction may contribute to growth of APC.These findings indicate miRNAs act together to promote tumour progression.This could help develop miRNA targeted therapy and study tumorigenesis process of head and neck cancer.

Reference:

- Zackrisson B, Mercke C, Strander H, Wennerberg J, Cavallin-ståhl E. A Systematic Overview of Radiation Therapy Effects in Head and Neck Cancer [Internet]. Vol. 42, Acta Oncologica. 2003. p. 443–61. Available from: http://dx.doi.org/10.1080/0284186031 0014886
- Network TCGA, The Cancer Genome Atlas Network. Comprehensive molecular characterization of human colon and rectal cancer [Internet]. Vol. 487, Nature. 2012. p. 330–7. Available from:

http://dx.doi.org/10.1038/nature11252

3. Powell SM, Zilz N, Beazer-Barclay Y,

Bryan TM, Hamilton SR, Thibodeau SN, et al. APC mutations occur early during colorectal tumorigenesis [Internet]. Vol. 359, Nature. 1992. p. 235–7. Available from: http://dx.doi.org/10.1038/359235a0

- Mori Y, Nagse H, Ando H, Horii A, Ichii S, Nakatsuru S, et al. Somatic mutations of the *APC* gene in colorectal tumors: mutation cluster region in the *APC* gene [Internet]. Vol. 1, Human Molecular Genetics. 1992. p. 229–33. Available from: http://dx.doi.org/10.1093/hmg/1.4.229
- Roberto A, Radrezza S, Mosconi P. Transparency in ovarian cancer clinical trial results: ClinicalTrials.gov versus PubMed, Embase and Google scholar [Internet]. Vol. 11, Journal of Ovarian Research. 2018. Available from: http://dx.doi.org/10.1186/s12048.018

http://dx.doi.org/10.1186/s13048-018-0404-1

 Jen J, Johnson C, Levin B. Molecular approaches for colorectal cancer screening [Internet]. Vol. 10, European Journal of Gastroenterology & Hepatology. 1998. p. 213–8. Available from: http://dx.doi.org/10.1097/00042737-

199803000-00004

- Groden J, Thliveris A, Samowitz W, Carlson M, Gelbert L, Albertsen H, et al. Identification and characterization of the familial adenomatous polyposis coli gene [Internet]. Vol. 66, Cell. 1991. p. 589–600. Available from: http://dx.doi.org/10.1016/0092-8674(81)90021-0
- Traboulsi EI, Maumenee IH, Krush AJ, Giardiello FM, Stefan Levin L, Hamilton SR. Pigmented Ocular Fundus Lesions in the Inherited

Gastrointestinal Polyposis Syndromes and in Hereditary Nonpolyposis Colorectal Cancer [Internet]. Vol. 95, Ophthalmology. 1988. p. 964–9. Available from: http://dx.doi.org/10.1016/s0161-6420(88)33093-9

- 9. Joslyn G, Carlson M, Thliveris A, Albertsen H, Gelbert L, Samowitz W, et al. Identification of deletion mutations and three new genes at the familial polyposis locus [Internet]. Vol. 66, Cell. 1991. p. 601–13. Available from: http://dx.doi.org/10.1016/0092-8674(81)90022-2
- Utsunomiya J, Lynch HT. Hereditary Colorectal Cancer: Proceedings of the Fourth International Symposium on Colorectal Cancer (ISCC-4) November 9–11, 1989, Kobe Japan. Springer Science & Business Media; 2013. 575 p.
- 11. Luongo C, Dove WF. Somatic genetic events linked to the apc locus in intestinal adenomas of the min mouse [Internet]. Vol. 17, Genes, Chromosomes and Cancer. 1996. p. 194–8. Available from: http://dx.doi.org/10.1002/1098-2264(199611)17:3<194::aidgcc2870170302>3.0.co;2-e
- van den Broek E, Krijgsman O, Sie D, Tijssen M, Mongera S, van de Wiel MA, et al. Genomic profiling of stage II and III colon cancers reveals APC mutations to be associated with survival in stage III colon cancer patients. Oncotarget. 2016 Nov 8;7(45):73876–87.
- Morin PJ, Sparks AB, Korinek V, Barker N, Clevers H, Vogelstein B, et al. Activation of β-Catenin-Tcf Signaling in Colon Cancer by

Mutations in β-Catenin or APC [Internet]. Vol. 275, Science. 1997. p. 1787–90. Available from: http://dx.doi.org/10.1126/science.275. 5307.1787

- Otoukesh B, Abbasi M, Gorgani HOL, Farahini H, Moghtadaei M, Boddouhi B, et al. MicroRNAs signatures, bioinformatics analysis of miRNAs, miRNA mimics and antagonists, and miRNA therapeutics in osteosarcoma. Cancer Cell Int. 2020 Jun 17;20:254.
- 15. Figure 13: miRNA gene regulatory network analysis [Internet]. Available from: http://dx.doi.org/10.7717/peerj.8907/fi g-13
- 16. Srivastava S, Singh N, Srivastava G, Sharma A. miRNA mediated gene regulatory network analysis of Cichorium intybus (chicory) [Internet]. Vol. 3, Agri Gene. 2017. p. 37–45. Available from: http://dx.doi.org/10.1016/j.aggene.201 6.11.003
- 17. Institute NC, National Cancer Institute. APC Gene Mutation [Internet]. Definitions. 2020. Available from: http://dx.doi.org/10.32388/pr62y3
- 18. Pouya F, Shariatpanahi AM, Ghaffarzadegan K, Yazdi SAT. Golmohammadzadeh H, Soltani G, et al. A novel large germ line deletion in adenomatous polyposis coli(APC)gene associated with familial adenomatous polyposis [Internet]. Vol. 6, Molecular Genetics & Genomic Medicine. 2018. 1031-40. Available from: p. http://dx.doi.org/10.1002/mgg3.479
- Preisler L, Habib A, Shapira G, Kuznitsov-Yanovsky L, Mayshar Y, Carmel-Gross I, et al. Heterozygous APC germline mutations impart predisposition to colorectal cancer. Sci

Rep. 2021 Mar 4;11(1):5113.

20. Gebert JF, Dupon C, Kadmon M, Hahn M, Herfarth C, von Knebel Doeberitz M, et al. Combined Molecular and Clinical Approaches for the Identification of Families with Familial Adenomatous Polyposis Coli [Internet]. Vol. 229, Annals of Surgery. 1999. p. 350–61. Available from: http://dx.doi.org/10.1007/00000658

http://dx.doi.org/10.1097/00000658-199903000-00008

- 21. Gebert J, Dupon C, Kadmon M, Tandara A, Herfarth C, von Knebel Doeberitz M. Genetic analysis of familial adenomatous polyposis (FAP) families: Lessons and implications [Internet]. Vol. 33, European Journal of Cancer. 1997. p. S11–2. Available from: http://dx.doi.org/10.1016/s0959-8049(97)84420-6
- 22. Pinyol M, Hernández L, Martínez A, Cobo F, Hernández S, Beà S, et al. INK4a/ARFLocus Alterations in Human Non-Hodgkin's Lymphomas Mainly Occur in Tumors with Wild-Type p53 Gene [Internet]. Vol. 156, The American Journal of Pathology. 2000. p. 1987–96. Available from: http://dx.doi.org/10.1016/s0002-9440(10)65071-7
- 23. Vijver MJ van de, van de Vijver MJ, He YD, van 't Veer LJ, Dai H, Hart AAM, et al. A Gene-Expression Signature as a Predictor of Survival in Breast Cancer [Internet]. Vol. 347, New England Journal of Medicine. 2002. p. 1999–2009. Available from: http://dx.doi.org/10.1056/nejmoa0219 67
- 24. Costa VL, Henrique R, Ribeiro FR, Carvalho JR, Oliveira J, Lobo F, et al. Epigenetic regulation of Wnt signaling

pathway in urological cancer [Internet]. Vol. 5, Epigenetics. 2010. p. 343–51. Available from: http://dx.doi.org/10.4161/epi.5.4.1174 9

- 25. Aldhuwayhi, Sami, Sreekanth Kumar Mallineni, Srinivasulu Sakhamuri, Amar Ashok Thakare, Sahana Mallineni, Rishitha Sajja, Mallika Sethi, Venkatesh Nettam, and Azher Mohiuddin Mohammad. 2021. "Covid-19 Knowledge and Perceptions Among Dental Specialists: А **Cross-Sectional** Online Questionnaire Survey." Risk Management and Healthcare Policy 14 (July): 2851-61.
- 26. Dua, Kamal, Ridhima Wadhwa, Gautam Singhvi, Vamshikrishna Rapalli, Shakti Dhar Shukla, Madhur D. Shastri, Gaurav Gupta, et al. 2019. "The Potential of siRNA Based Drug Delivery in Respiratory Disorders: Recent Advances and Progress." Drug Development Research 80 (6): 714-30.
- 27. Gan, Hongyun, Yaqing Zhang, Qingyun Zhou, Lierui Zheng, Xiaofeng Xie, Vishnu Priya Veeraraghavan, and Surapaneni Krishna Mohan. 2019. "Zingerone Induced Caspase-Dependent Apoptosis in MCF-7 Cells and Prevents 7,12-Dimethylbenz(a)anthracene-Induced Mammary Carcinogenesis

in Experimental Rats." Journal of Biochemical and Molecular Toxicology 33 (10): e22387.

28. Jayaraj, Gifrina, Pratibha Ramani, Herald J. Sherlin, Priya Premkumar, and N. Anuja. 2015. "Inter-Observer Agreement in Grading Oral Epithelial Dysplasia – A Systematic Review." Journal of Oral and Maxillofacial Surgery, Medicine, and Pathology. https://doi.org/10.1016/j.ajoms.201 4.01.006.

- 29. Li, Zhenjiang, Vishnu Priya Veeraraghavan, Surapaneni Krishna Mohan, Srinivasa Rao Bolla, Hariprasath Lakshmanan, Subramanian Kumaran. Wilson Aruni, et al. 2020. "Apoptotic Induction and Anti-Metastatic Activity of Eugenol Encapsulated Chitosan Nanopolymer on Rat Glioma C6 Cells via Alleviating the MMP Signaling Pathway." Journal of Photochemistry and Photobiology B: Biology. https://doi.org/10.1016/j.jphotobiol .2019.111773.
- 30. Markov. Alexander. Lakshmi Thangavelu, Surendar Aravindhan, Angelina Olegovna Zekiy, Mostafa Jarahian, Max Stanley Chartrand, Yashwant Pathak, Faroogh Marofi, Somayeh Shamlou, and Ali Hassanzadeh. 2021. "Mesenchymal Stem/stromal Cells as a Valuable Source for the Treatment of Immune-Mediated Disorders." Stem Cell Research & Therapy 12 (1): 192.
- Mohan, Meenakshi, and Nithya Jagannathan. 2014. "Oral Field Cancerization: An Update on Current Concepts." Oncology Reviews 8 (1): 244.
- Neelakantan, Prasanna,
 Deeksha Grotra, and Subash
 Sharma. 2013. "Retreatability of 2
 Mineral Trioxide Aggregate-Based

Root Canal Sealers: A Cone-Beam Computed Tomography Analysis." Journal of Endodontia 39 (7): 893– 96.

- 33. Paramasivam, Arumugam, Jayaseelan Vijayashree
 Priyadharsini, Subramanian
 Raghunandhakumar, and Perumal
 Elumalai. 2020. "A Novel COVID-19 and Its Effects on Cardiovascular
 Disease." Hypertension Research:
 Official Journal of the Japanese
 Society of Hypertension.
- 34. Sheriff, K. Ahmed Hilal, K. Ahmed Hilal Sheriff, and Archana Santhanam. 2018. "Knowledge and Awareness towards Oral Biopsy among Students of Saveetha Dental College." Research Journal of Pharmacy and Technology. https://doi.org/10.5958/0974-360x.2018.00101.4.