Study of Some Boswellia Carterii Contents and Effect of their Extracts as Antioxidants and Antibacterial

Heyam A. Hashim

Department of Chemistry, College of Science, Al-Mustansiriyah University, Baghdad, Iraq, hiamchem@gmail.com

Mustafa T. Mohammed

Department of Chemistry, College of Science, Al-Mustansiriyah University, Baghdad, Iraq.

Mohammed Z. Thani

Department of Chemistry, College of Science, Al-Mustansiriyah University, Baghdad, Iraq.

Abstract

Boswellia carterii leaves are one of the most popular herbal medicines. The purpose of this study was to use Boswellia carterii leaf extracts to find out how many free amino acids are in a sample. Phenolic compounds, particularly flavonoids, were estimated in the three types of extracts using the high-performance liquid chromatography device (HPLC) and many other important biologically active factors due to their benefits to human health and antioxidant and antibacterial effects. Aqueous extracts, ethanol and ethyl acetate showed five types of amino acids, the highest of which was Glutamic acid. In addition to containing good numbers and quantities of phenolic compounds, many of which contain multiple biological properties such as antioxidant and antibacterial, this research has been carried out. An antioxidant analysis showed that ethanol extract showed more antioxidant properties than aqueous extract and ethyl acetate extract. However, compared to vitamin C measurement the three extract sections were able to inhibit a good ratio of 1, 1- diphenyl 2-picrylhydrazyl free radical (DPPH) with a good excellent ratio balance. The maximum activity of DPPH root rickets for vitamin C (38.5) ppm, ethanol extract (69.5) ppm, ethyl acetate extract(115) ppm and aqueous extraction (125) ppm were concentrated. The antibacterial activity showed a maximum inhibition area of 24 mm for Staphylococcus aureus as Shigella flexneri at a concentration of 500 ppm .

Keywords: Boswellia carterii, Amino acids, Phenolic compounds, Antioxidants, Antibacterial.

INTRODUCTION

The Boswellia genus is made up of around 20 species; however, this number is in dispute.1 Boswellia species are well known for producing frankincense, an aromatic oleogum resin Frankincense has been used in traditional medicine and cultural and religious ceremonies for thousands of years. More

recently essential oils derived from frankincense resins have become[1] Among the various species of Boswellia, only a few are traded in a significant amount: B. sacra, B. carteri, B. frereana, B. papyrifera, and B. serrata. The highest quality frankincense resins are produced by B. carteri trees [2] Studies have also demonstrated that these

extracts can improve blood flow, lower the risk of clotting, support capillary walls and their flexibility, and protect neurons from damage that could result from oxygen deprivation[3] Boswellia carterii leaves contain large amounts of protein, lipids, carbohydrates, vitamins, organic acids, polyphenols, beta carotene, flavonoids, and terpenoids [4]. In addition to the well-known terpene lactones and flavonol glycosides, amino acids from Ginkgo Biloba leaves are frequently used in meals, drinks, and medications [5]. Chemical compounds known as amino acids (AAs) have both amino and acid groups[6]. The fundamental building ingredients that support life are amino acids. It is well known how they contribute to the production of protein synthesis, this metabolic rewiring and support for numerous immune cell functions depend on crucial amino acid metabolism. [7][8] .Many plants contain a broad collection of secondary metabolites called flavonoids. It has a flavone structure, in which the C ring of the pyran ring (which may have hydroxyl, methoxy, methyl, or isoamyl substituents) connects the two benzene rings (A and B rings)[9] Plants receive their color, tastes, and medicinal effects from flavonoids [10] The main sources of flavonoids are fruits and vegetables [11]. Plants are shielded from harmful environmental factors by flavonoids [12], which also contain biological features like increased blood circulation, lowered cholesterol, UV protection, angiogenesis suppression, antibacterial, and antiinflammatory Activities[9] They have consequently attracted a lot of interest. To determine if they can aid in treating a variety of acute and chronic human illnesses, they have been put to the test in numerous epidemiological and experimental research [14]. In vitro and animal studies have demonstrated the anti-inflammatory and

immunomodulatory activities of flavonoids [15] as well as their powerful anticancer properties [14]. [16] [17]. The oxidative effects of free radicals and other oxidants in biological systems can be mitigated by antioxidants [18] Based on their existence in the human body, antioxidants are classified as endogenous or exogenous[19][20] Due to their special chemical and biological characteristics, they are advantageous to a variety of human products [21]. Because they can shield the body against oxidative damage to biological macromolecules including lipid, protein, and nucleic acid, which is predominantly brought on by secondary metabolism, the hunt for new and natural antioxidants from dietary plants is gaining traction[22].

MATERIAL AND METHODS

Sample Collection:

Boswelliacarterii leaves were collected from Sulaymaniyah governorate, Iraq, in September 2022. The leaves were then washed with deionized water, dried in shade for several days at room temperature, and ground as a powder The conventional Soxhlet extraction method was applied to samples according to [24] for obtaining aqueous and alcohol and ethyl acetate extract.

Determination of Amino Acids and Phenols concentrations:

All of the reference materials were made up entirely (99%) of Samarra Pharmaceuticals using an HPLC system (Sykamn s3210, Germany) and a C18 column (4.6mm 150 mm, 5m). Mobile phase (acetonitrile: H2O) (5: 95) for amino acids, the flow rate of 2 mL/min, and the detection system using fluorescence (340nm-450nm) are the corresponding values [25]. For the following parameters: flow rate (1.2 mL/min), detection fluorescence system (260nm-310nm) and phenols mobile phase (H2O: acetonitrile)(70:30) respectively [26]

Antibacterial Activity studies:

Many microorganisms were employed to bacteria; bacteria from selected create microorganisms were tested for antibacterial activity using a spreading tablet; and Boswellia extracts were tested for antimicrobial activity using a well-chosen deployment method. [27][28] According to the McFarland method, samples were loaded on cellulose tablets in diameter (5mm) bacterial insulation was developed, and 0.5 was transported 1 ml of bacterial and bacterial filaments equivalent to (1.5 108) cells/ml to the dish that contains the center of Müller Hanton's den. The study included testing inkind water, alcohol and ethyl acetate extracts from the Boswellia plant in inhibiting the growth of bacteria causing diseases positive for Kram dye outside the living body until the implant is absorbed, disseminate, and let the dish at room temperature for 15-20 minutes. At temperature of 37 C, bacteria are grown on plates, extract-loaded tablets, and other media. For each type of bacteria utilized, the diameters of the tablet regions around the chips are measured for a whole day. Determination of free radical scavenging activity

The DPPH was used to assess the free radical scavenging capacity of Boswellia carterii

preparations. The extract was assessed at various concentrations (12.5, 25, 50, 100, and 200 g/mL). An addition of a (150L) methanol solution comprising varying quantities of sample solution was made to 3 mL of daily generated methanol and DPPH solution The sample's scavenging activity was determined by comparing its absorbance to that of the DPPH reference solution The following formula was used to evaluate the radical scavenging activity percentage of the DPPH: % Antiradical activity percentage equals (Ao - AC) /Ao× **100** Where Ao is the absorbance of the control and Ac is the absorbance of the sample or standard in the present of the control. The results of three independent trials were averaged and expressed as a percentage of mean radical scavenging activity [29].

RESULTS AND DISCUSSION

The results revealed that B. carterii leaf extracts were high in free amino acids, especially those considered essential, such as (Arginine, Valine, and Glycine,), in addition to (Cysteine, Alanine, and Glutamic acid)The total amount of amino acids in the sample ranged from (10.25 to 14.00) μ g /gm in aqueous extract, while it was from (13.59 to 16.28) μ g /gm in the alcoholic extract, and in the ethylacetate extract it was from(8.22 to 15.98) μ g /gm according to the Table (1) and Fig.(1)

 Table 1: Concentration of amino acids in the aqueous, alcohol extracts and Ethyl acetate

 extract of the Boswellia carterii

Aqueous extract				Ethyl acetate extract			
Amino acid	Retain. Time [min]	Amount [μg / gm]	Retain. Time [min]	Retain. TimeAmountRetain. Time[min][µg /gm][min]			
Alanine	3.00	11.25	3.09	13.59	-	-	
Arginine			-	-	5.24	8.22	

Cysteine	-	-	-	-	-	-
Glutamic acid	11.62	14.00	11.65	16.28	11.71	15.98
Glycine	12.67	12.59	12.54	14.11	-	-
Proline	15.08	10.25	15.03	13.69	15.03	10.46

Figure 1: HPLC analysis for(A) Amino acid in the aqueous extract(B) Amino acid in ethanol extract (C) Amino acid in ethyl acetate extract







acids serve as vital Amino structural constituents and a source of energy. An amino group, a carboxylic acid, and a unique carbon structure are all present in every amino acid. Twenty amino acids are required by humans: the majority of them can be produced naturally, but nine of them are "essential," meaning they must be ingested The components of proteins that are either ketogenic, glycogenic, or both, recycle as needed, and serve as the building blocks for intermediates in the Krebs cycle and other metabolites, making amino acids essential energy sources [30] Amino acids (AA) are used of product protein and a variety of lowmolecular-weight molecules with physiological important. Nutritionally, amino acids are essential for maximal growth breastfeeding and good health, well-being, and reproduction [31] In the human body, specific amino acids can be changed into other amino acids, proteins, glucose, fatty acids, or ketones Neurotransmitters, chemical [32]. or messengers in the nervous system, are Alanine play a direct role in gluconeogenesis; transamination [34]. precursors to nucleic acids, which are parts of DNA [33]. Alanine has a direct function in gluconeogenesis; transamination [34] Arginine contains antioxidants, hormone secretion, ammonia detoxification, expression, gene and immunological function-regulating capabilities [35] Cysteine is a disulfide bond in proteins that allows sulfur to be transferred [36]. Glutamine: The production of arginine connects the urea and Krebs cycles. Isoleucine Synthesis of glutamine and alanine; regulation of protein synthesis and gene expression; activator of glutamate.; production of tyrosine; neurological development [37]. Tyrosine is the source of dopamine, epinephrine, norepinephrine, and thyroxin [38] The HPLC analysis of Boswellia carterii Extracts looked for important antioxidant phenolic compounds in this study We utilized several a variety of standard solutions for phenols. The results demonstrated that B.carterii intriguing focus extracts because it contains a high number of phenol which each compound was calculated by comparing the area of the pick of the standard substance with the size of the picked choice for the desired combination and according to the subsequent equation: $C(\text{sample}) = [A (\text{sample}) \times A (\text{standard})] / C$ average In the aqueous extract. the concentration of (Glumamic acid) was the highest and had the best separation of the peak at retention time (11.62min) according to the peak of standard substances, and the presence of all phenolic compounds at range retention time (2-11min), Figure (2) and table (2). In the alcohol extract. the concentration of (Glumamic acid) too was the highest and had the best separation of the peak at retention time (11.65 min) according to the rise of standard substances, and the presence of all phenolic compounds at the range retention time (2-11min), Figure (2) and table (2). In the ethylacetate extract, the concentration of (Glumamic acid)) was the highest and had the best separation of the peak at retention time (11.71min) according to the peak of standard substances, and the presence of all phenolic compounds at range retention time (2-11min), Figure (2) and table (2).

phenolic compounds	Result Aqueus Extract		Result in Alcohol Extract			Result in et	Retentio n Time [Min]			
	Area[m V.s]	conc.[µ g.mL- 1]	Reten tion	Area[mV. s]	conc.[µ g.mL- 1]	Retenti on	Area[mV. s]	conc.[µ g.mL-1]	Retenti on	Standar d
Apigenin	9652.58	48.457	11.32	932.02	14.338	11.36	-	-	-	11.38
P-coumaric acid	-	-	3.90	3652.19	36.521	3.95	1625.49	-	-	7.90
Ferulic acid	2305.98	25.036	8.72	1263.58	12.365	8.74	2013.69	21.862	8.72	8.72
Vanillic acid	1365.28	24.191	4.20	-	-	-	-	-	-	4.20
Gallic acid	2698.59	33.726	5.92	2140.88	26.750	5.90	598.28	7.477	5.92	5.91
Qurcetin	-	-	-	2564.85	29.005	2.19	-	24.254	2.15	2.15
Kaempfero l	580.44	46.492	3.90		62.917	3.95	-	-	-	3.90

Table 2: Concentration of phenolic compounds in the aqueous and alcohol extracts of the Boswellia carterii

Figure 2: HPLC analysis for(A) Phenolic compounds in the aqueous extract. (B) Phenolic compounds in ethanol extract (C) Phenolic compounds in ethyl acetate extract





Figure 3: HPLC analysis for standards phenolic compound







Figure 4: HPLC analysis for standards phenolic compound





 Table 3: Values the standard deviation of the aqueous, alcohol extracts and ethyl acetate of the Boswellia carterii

Sample No.					S2	S 3	S4
		S.aureus	Mean	-	-	-	-
	Gram- positive		Sd	-	-	-	-
		S.epidermidis	Mean	-	-	-	-
			Sd	-	-	-	-
Aqueous Extract	Gram- Negative Fungi	E .coli	Mean	-	-	-	-
			Sd	-	-	-	-
		Klebsiella sp.	Mean	-	-	-	-
			Sd	-	-	-	-
		C.albicans	Mean	-	-	-	-
			Sd	-	-	-	-
	Gram- positive Gram- Negative Fungi	S.aureus	Mean	20	9	22	-
			Sd	16	-	17	-
		S.epidermidis	Mean	10	-	9	-
Alcohol Extract			Sd	12	10	12	-
		E .coli	Mean	17	15	14	-
			Sd	1.44	1.56	1.34	-
		Klebsiella sp.	Mean	-	-	-	-
			Sd	-	-	-	-
		C.albicans	Mean	21	8	22	-
			Sd	1.12	1.52	1.94	-
Ethyl acetate Extract		S.aureus	Mean	-	-	-	-

Gram-		Sd	-	-	-	-
positive	S.epidermidis	Mean	-	-	-	-
		Sd	-	-	-	-
Gram-	E .coli	Mean	-	-	-	-
Negative		Sd	-	-	-	-
	Klebsiella sp.	Mean	-	-	-	-
		Sd	-	-	-	-
Fungi	C.albicans	Mean	-	-	-	-
		Sd	-	-	-	-

Figure 5: Inhibition zones caused by Boswellia carterii extracts Against Staphylococcus aureus, Staphylococcus



Microorganisms can be inhibited through a variety of mechanisms, including cell membrane damage. protein synthesis and disruption of inhibition, biological processes and cell membranes by particular enzymes. The presence of compounds like -Pinene and -Pinene (pinene-type monoterpene hydrocarbons), which are involved in the disruption of the membrane cell wall by the lipophilic compounds, has an impact on the nature of the bacterial cell wall [45] Additionally, bacteria like Escherichia coli are resistant to antibiotics because gram-negative bacteria's cell walls have a high lipid content (up to 20%) compared to gram-positive bacteria's lipid content of 0-2%. Positive gram-positives, on the other hand, The antibacterial substances present in extracts are

more sensitive to microorganisms [40]. According to research, Boswllia alcohol extract has more antibacterial action against gram-positive bacteria than gram-negative bacteria that are resistant to each of the three extracts water and ethanol [46]. Free radical activity The DPPH radical scavenging scavenging assay is a sensitive antioxidant assay substrate that is unaffected by the polarity of the substrate A hydrogen radical or an electron can be taken up by the stable free radical DPPH to create a stable diamagnetic molecule The extracts' high phenolic content may explain for their potent antioxidant properties. Based on these results, antioxidant activity against DPPH radicals was shown by both the extracts and the mixed sample three extractions showed composite action antioxidant efficacy against DPPH radicals. The fascinating anti-oxidant stress activity of gingko leaf extracts is therefore revealed by this exploratory study, suggesting its potential use as a medicinal source for the treatment and prevention of disease. It has therapeutic and pharmaceutical properties that can treat diseases brought on by free radicals. Fig (6) Data indicated that the three extracts contain a good percentage of antioxidants when compared to vitamin C, a potent and efficient antioxidant [47].

Figure 6: DPPH Free radical scavenging activities of Boswellia carterii extracts compared with Vitamin C.



A: Vitamin C (IC50=32.909 ppm), B: Ethanol Extract (IC50= 42.421ppm), C:

Aqueous Extract(IC50=53.64ppm)

D:Ethyl Acetate Extract (IC50 =IC50=77.078ppm) the outcomes demonstrated that water extracts, ethanol and ethyl acetate from carterii boswellio leaves had DPPH radical scavenging action. According to their diverse concentrations of 10 g/ml to 60 g/ml, the ethanol extract showed the highest percentage of scavenging activity. Boswellia carterii leaf extracts' strong DPPH scavenging activity is consistent with reports [48] The increasing intensity of the color is directly proportional to the inhibition of DppH[49] and phenols. This high content of flavonoids and phenols is the primary driver of their antioxidant activities. Ethanol also showed more extraction of phenolic content than extraction aqueous and ethyl acetate extraction, so ethanol extract showed the best antioxidant properties. The increasing

concentration of the extract is increasingly preventing DPPH activity. Ethanol extracts also had a clear and apparent effect on bacterial inhibition compared to aqueous and ethyl acetate extracts.

CONCLUSIONS

This study established that Boswellia carterii leaf extracts contain significant levels of phenols, flavonoids, and amino acids. The abundance of flavonoids and phenols is the primary driver of their antioxidant activities. Ethanol also showed 99% more extraction of phenolic content than aqueous extraction, so ethanol extract showed the best antioxidant properties. The extract's concentration is limiting DPPH activity more and more as it grows. In comparison to aqueous extracts, ethanol extracts also appeared to have a clear influence on bacterial inhibition.

ACKNOWLEDGMENT

Like to thank the Deanship of the Faculty of Science Head of chemistry and all the staff in the Chemistry Department of the Faculty of Science for their support during my studies.

Reference

- Ojha, P. K., Poudel, D. K., Rokaya, A., Satyal, R., Setzer, W. N., & Satyal, P. (2022). Comparison of Volatile Constituents Present in Commercial and Lab-Distilled Frankincense (Boswellia carteri) Essential Oils for Authentication. Plants, 11(16), 2134..
- Johnson, S., DeCarlo, A., Satyal, P., Dosoky, N. S., Sorensen, A., & Setzer, W. N. (2019). The chemical composition of Boswellia occulta oleogum resin essential oils. Natural Product Communications, 14(7), 1934578X19866307.
- [3] DeCarlo, A., Johnson, S., Poudel, A., Satyal, P., Bangerter, L., & Setzer, W. N.

(2018). Chemical Variation in Essential Oils from the Oleo - gum Resin of Boswellia carteri: A Preliminary Investigation. Chemistry & biodiversity, 15(6), e1800047.

- [4] Huang, K., Chen, Y., Liang, K., Xu, X., Jiang, J., Liu, M., & Zhou, F. (2022). Review of the chemical composition, pharmacological effects, pharmacokinetics, and quality control of Boswellia carterii. Evidence-Based Complementary and Alternative Medicine..,.
- [5] Gafurdjanov, B., Berdiev, E., & Xoliyorov, U. (2021, December). Study on the breeding ginkgo (ginkgo biloba l.) in Tashkent oasis. In IOP Conference Series: Earth and Environmental Science (Vol. 939, No. 1, p. 012058). IOP Publishing.
- [6] Hosain, N. A., Ghosh, R., Bryant, D. L., Arivett, B. A., Farone, A. L., & Kline, P. C. (2019). Isolation, structure elucidation, and immunostimulatory activity of polysaccharide fractions from Boswellia carterii frankincense resin. International journal of biological macromolecules, 133, 76-85.
- [7] Kelly, B., & Pearce, E. L. (2020). Amino assets: how amino acids support immunity. Cell metabolism, 32(2), 154-175.
- [8] Mutar, Y. S., Al-Rawi, K. F., & Mohammed, M. T. (2021). Moringa oleifera: Nutritive importance and its medicinal application, as a Review. Egyptian Journal of Chemistry, 64(11), 6827-6834.
- [9] Liu, J., Mu, T., Sun, H., & Fauconnier, M. L. (2019). Optimization of ultrasonic– microwave synergistic extraction of flavonoids from sweet potato leaves by response surface methodology. Journal of

Food Processing and Preservation, 43(5), e13928.

- [10] Scarano, A., Chieppa, M., & Santino, A. (2018). Looking at flavonoid biodiversity in horticultural crops: A colored mine with nutritional benefits. Plants, 7(4), 98.
- [11] Liu, J., Wang, X., Yong, H., Kan, J., & Jin, C. (2018). Recent advances in flavonoid-grafted polysaccharides: Synthesis, structural characterization, bioactivities and potential applications. International Journal of Biological Macromolecules, 116, 1011-1025..
- [12] Chongsrimsirisakhol, O., & Pirak, T. (2022). Total polyphenol content and antioxidant properties of cold brew coffee extracts as affected by ultrasound treatment and their application in low fat pork sausage. International Journal of Food Properties, 25(1), 813-826..
- [13] Nabavi, S. M., Šamec, D., Tomczyk, M., Milella, L., Russo, D., Habtemariam, S., ... & Shirooie, S. (2020). Flavonoid biosynthetic pathways in plants: Versatile targets for metabolic engineering. Biotechnology advances, 38, 107316.
- [14] Rodríguez-García, C., Sánchez-Quesada, C., & Gaforio, J. J. (2019). Dietary flavonoids as cancer chemopreventive agents: An updated review of human studies. Antioxidants, 8(5), 137.
- [15] Yahfoufi, N., Alsadi, N., Jambi, M., & Matar, C. (2018). The immunomodulatory and anti-inflammatory role of polyphenols. Nutrients, 10(11), 1618.
- [16] Abotaleb, M., Samuel, S. M., Varghese, E., Varghese, S., Kubatka, P., Liskova, A., & Büsselberg, D. (2018). Flavonoids in cancer and apoptosis. Cancers, 11(1), 28..
- [17] Chirumbolo, S., Bjørklund, G., Lysiuk, R., Vella, A., Lenchyk, L., & Upyr, T.

(2018). Targeting cancer with phytochemicals via their fine tuning of the cell survival signaling pathways. International journal of molecular sciences, 19(11), 3568..

- [18] Gulcin, İ. (2020). Antioxidants and antioxidant methods: An updated overview. Archives of toxicology, 94(3), 651-715.
- [19] Yadav, A., Kumari, R., Yadav, A., Mishra, J. P., Srivatva, S., & Prabha, S. (2016). Antioxidants and its functions in human body-A Review. Res. Environ. Life Sci, 9(11), 1328-1331.
- [20] Khalisyaseen, O., & Mohammed, M. T. (2021). Identification Of Some Antioxidant Active Compounds In True Cinnamon (Cinnamomm Zeylanicum) Bark Extract. NVEO-Natural Volatiles & Essential Oils Journal| NVEO, 7565-7577.
- [21] Fu, P. P., Xia, Q., Hwang, H. M., & Paresh, C. (2014). Ray, and Hongtao Yu." Mechanisms of Nanotoxicity: Generation of Reactive Oxygen Species.". Journal of food and drug analysis, 22, 64-75..
- [22] Patel, V. B., Watson, R. R., & Preedy, V. R. (Eds.). (2011). Nuts and seeds in health and disease prevention. Academic Press.
- [23] More, M. P., Motule, A. S., Dongare, P. N., Patinge, P. A., Jawarkar, R. D., Bakal, R. L., & Manwar, J. V. (2021). Pharmacognosy, phytochemistry, pharmacology and clinical application of Ginkgo biloba. GSC Biological and Pharmaceutical Sciences, 16(2), 229-240.
- [24] Ulusoy, H. İ., Acıdereli, H., & Tutar, U.
 (2017). Optimization of extraction parameters for fat soluble vitamins and major element analysis in Polygonum cognatum Meissn plant (Madimak). Journal of the Turkish Chemical Society Section A: Chemistry, 4(1), 165-178.

- [25] Sedehi, S., Tabani, H., & Nojavan, S. (2018). Electro-driven extraction of polar compounds using agarose gel as a new membrane: determination of amino acids in fruit juice and human plasma samples. Talanta, 179, 318-325..
- [26] Li, Y., Wang, R., Lin, Y., Han, B., Wang, B., & Wang, S. (2020). Qualitative and quantitative analysis of phenolic acid glycosides in Ginkgo biloba L. leaf, G. biloba leaf extract and its injection. Biomedical Chromatography, 34(12), e4964.
- [27] Sati, P., Dhyani, P., Bhatt, I. D., & Pandey, A. (2019). Ginkgo biloba flavonoid glycosides in antimicrobial perspective with reference to extraction method. Journal of traditional and complementary medicine, 9(1), 15-23.
- [28] Orabi, M., Abdulsattar, J. O., & Nasi, Z.
 O. (2022). Phytochemical Profile, Antimicrobial, Antioxidant Activity and Cyclooxygenase 2 Inhibitory Properties of Nutmeg (Myristica Fragrans) Seeds Extract. Egyptian Journal of Chemistry, 65(1), 317-326.
- [29] Szewczyk, A., Kwiecień, I., Grabowski, M., Rajek, K., Cavò, E., Taviano, M. F., & Miceli, N. (2021). Phenylalanine increases the production of antioxidant phenolic acids in Boswellia carterii cell cultures. Molecules, 26(16), 4965..
- [30] Kandasamy, P., Gyimesi, G., Kanai, Y., & Hediger, M. A. (2018). Amino acid transporters revisited: New views in health and disease. Trends in biochemical sciences, 43(10), 752-789.
- [31] Hou, Y., Yin, Y., & Wu, G. (2015). Dietary essentiality of "nutritionally nonessential amino acids" for animals and humans. Experimental Biology and Medicine, 240(8), 997-1007.

- [32] Marczuk, J., Brodzki, P., Brodzki, A., & Kurek, Ł. (2018). The concentration of free amino acids in blood serum of dairy cows with primary ketosis. Polish journal of veterinary sciences, 21(1), 149-156.
- [33] Farina, M., & Aschner, M. (2017). Methylmercury-induced neurotoxicity: focus on pro-oxidative events and related consequences. Neurotoxicity of Metals, 267-286.
- [34] Ramadhani, R., & Prijanti, A. R. (2019). Alanine amino transferase (ALT) specific activities in long term systemic hypoxic rat brain tissues. Acta Biochimica Indonesiana, 2(2), 75-81.
- [35] Birmani, M. W., Raza, A., Nawab, A., Tang, S., Ghani, M. W., Li, G., ... & An, L. (2019). Importance of arginine as immune regulator in animal nutrition. Int. J. Vet. Sci. Res, 5, 1-10.
- [36] Pedre, B., Barayeu, U., Ezeriņa, D., & Dick, T. P. (2021). The mechanism of action of N-acetylcysteine (NAC): The emerging role of H2S and sulfane sulfur species. Pharmacology & therapeutics, 228, 107916.
- [37] Rofifah, D. (2020). 済無 No Title No Title No Title. Paper Knowledge. Toward a Media History of Documents, 3(3), 12-26.
- [38] Salamanca, N., Giráldez, I., Morales, E., de La Rosa, I., & Herrera, M. (2020). Phenylalanine and tyrosine as feed additives for reducing stress and enhancing welfare in gilthead seabream and meagre. Animals, 11(1), 45.
- [39] Jucá, M. M., Cysne Filho, F. M. S., & de Almeida, J. C. (2020). D. d. S. Mesquita, JR d. M. Barriga, KCF Dias, TM Barbosa, LC Vasconcelos, LKAM Leal and JE Ribeiro. Nat. Prod. Res, 34, 692-705.

- [40] Ashrafizadeh, M., Ahmadi, Z., Farkhondeh, T., & Samarghandian, S. (2020). Autophagy regulation using luteolin: new insight into its anti-tumor activity. Cancer Cell International,
- [41] Spagnuolo, C., Moccia, S., & Russo, G. L. (2018). Anti-inflammatory effects of flavonoids in neurodegenerative disorders. European journal of medicinal chemistry, 153, 105-115.
- [42] Bessa, C., Francisco, T., Dias, R., Mateus, N., Freitas, V. D., & Pérez-Gregorio, R. (2021). Use of polyphenols as modulators of food allergies. From chemistry to biological implications. Frontiers in Sustainable Food Systems, 5, 623611.
- [43] S. Ramapatruni, S. N. Narayanan, S. Mittal, A. Joshi, and K. Joshi, "Please provide feedback Please support the ScholarWorks @ UMBC repository by emailing scholarworks-group@umbc.edu and telling us what having access to this work means to you and why it 's important to you . Thank you . Anomaly Detection Models for Smart," 2019.
- [44] Lalani, S., & Poh, C. L. Flavonoids as antiviral agents for enterovirus A71 (EV-A71). Viruses. 2020; 12: 184..
- [45] Liu, Y. Y., Yao, W. M., Wu, T., Xu, B. L., Chen, F., & Cui, L. (2011). Captopril improves osteopenia in ovariectomized rats and promotes bone formation in osteoblasts. Journal of bone and mineral metabolism, 29, 149-158.
- [46] Pereira, S., Santos, R. S., Moreira, L., Guimarães, N., Gomes, M., Zhang, H., ... & Azevedo, N. F. (2021). Lipoplexes to deliver oligonucleotides in gram-positive and gram-negative bacteria: towards treatment of blood infections. Pharmaceutics, 13(7), 989.

- [47] Zhou, X., Qi, Y., & Chen, T. (2017). Long-term pre-treatment of antioxidant Ginkgo biloba extract EGb-761 attenuates cerebral-ischemia-induced neuronal damage in aged mice. Biomedicine & Pharmacotherapy, 85, 256-263.
- [48] Cao, J., Chen, L., Li, M., Cao, F., Zhao, L., & Su, E. (2018). Efficient extraction of proanthocyanidin from Ginkgo biloba leaves employing rationally designed deep eutectic solvent-water mixture and evaluation of the antioxidant activity. Journal of pharmaceutical and biomedical analysis, 158, 317-326..
- [49] Kingsley, N. O., Ikechukwu, U. K., Chinwe, O. A. N., & Okechukwu, O. D. (2019). Investigation of the antioxidant activity of aqueous and ethanol leaf extracts of ginkgo biloba from South-East Nigeria. International Journal of Plant Science and Ecology, 5, 31-36.