

Metabolic Abnormalities of Obesity Induced by Monosodium Glutamate and Their Effect on Male Sex Hormones in Rats

Sherif. Y. Saleh

Department of Biochemistry physiotherapy portsaid University, portsaid , Egypt

Elsayed H. Eltamany

Chemistry Department, Faculty of science, Suez Canal University, Ismailia, Egypt.

Eman. A. Elshamy

*Chemistry Department, Faculty of science, Suez Canal University, Ismailia, Egypt,
nimissy@gmail.com*

Abstract

This study aimed to identify the potentially negative impacts of monosodium glutamate (MSG) on male rat sex hormones and several biochemical markers. There were two primary groups made up of thirty male Sprague Dawley rats. The first group (n=15) was given distilled water (serves as negative control). The other group (n=15) received 15 mg/kg B.W. of MSG orally throughout a three-month period. According to results the amount of total testosterone (TT) significantly dropped. In addition, Triacylglycerol (TAG), total cholesterol (TC), glycated haemoglobin (HbA1c), insulin, leptin, tumour necrosis factor alpha (TNF- α), interleukin-6 (IL-6), liver enzymes, renal function, follicle-stimulating hormone (FSH), and luteinizing hormone (LH) were significantly higher than before. prolactin (PRL) levels did not alter, though. It can be said that MSG has severe effects on the liver, kidneys, lipid profile, and may be a factor in male infertility.

Keywords: *obesity; sex hormones; monosodium glutamate*

INTRODUCTION

Obesity is a global problem which increases at an alarming rate in the world during the last three decades. It is a complex, multifactorial chronic disease involving environmental, genetic, physiological, metabolic, behavioral, and psychological components (1). Obesity is closely associated with the development of metabolic abnormalities including glucose intolerance, insulin resistance (IR), type 2 diabetes (T2D), cardiovascular diseases (CVD), metabolic syndrome, hyperlipidemia, chronic inflammation and male hypogonadism (2). An essential part of controlling inflammatory processes is played by fat tissue. Increased production of cytokines that cause a low-grade inflammatory response has been

linked to increased visceral fat, and this could eventually lead to insulin resistance (3). Particularly, the establishment of insulin resistance in obesity and type 2 diabetes is stimulated by the production of numerous hormones and cytokines by adipose tissue which have side effects on skeletal muscle and the liver (4). In the regulation of insulin sensitivity, tumour necrosis factor type alpha (TNF- α) and adiponectin have shown an inverse association; whereas TNF- α contributes significantly to insulin resistance, adiponectin promotes insulin sensitivity. Other cytokines are also thought to be involved in this process (5). Interleukine-6 (IL-6) also rises with obesity and is strongly associated with endothelial dysfunction and sub-clinical

inflammation. Leptin has also been linked to disorders like insulin resistance syndrome (6) that include obesity, type 2 diabetes, dyslipidemia, and cardiovascular disease. Obesity and metabolic disorder in males are related to hypogonadism as a common feature. Thus, luteinizing hormone (LH) and follicle-stimulating hormone (FSH) are secreted from the pituitary as a result of the pulsatile release of hypothalamic gonadotropin-releasing hormone (GnRH) (7). The primary regulating hormones that affect the testicular cells are these gonadotropins. Inhibin synthesis is stimulated by FSH action on sertoli cells, which also indirectly controls spermatogenesis. On the other hand, inhibin works directly to downregulate pituitary FSH release through a negative feedback mechanism. LH encourages Leydig cells to increase testosterone production and steroidogenesis. Moreover Spermatogenesis is mediated by testosterone through its nuclear receptors in the sertoli cell (8).

Monosodium glutamate (MSG) (E621) is a common salt of glutamic acid widely used as a flavor enhancer at home as well as in food industry (9). Therefore, most canned food and fast food such as flavored flakes, ready meals, canned soups, mutton meat, bottled soy or Eastern sauces, tested and frozen tuna containing different concentrations of MSG (10).

MSG is harmful to both humans and test subjects' animals (11).it seems to have symptoms like headaches, flushing, numbness, dizziness, and weakness (12). High dosages of MSG have been linked to a number of diseases, including urticaria, cardiac arrhythmia, neuropathy, asthma, and atopic dermatitis (12). Additionally, it has neurotoxic effects that result in damage to brain cells, endocrine issues, retinal degeneration, and some

pathological conditions like addiction, stroke, epilepsy, brain trauma, neuropathic pain, schizophrenia, anxiety, and depression, as well as Alzheimer's disease, Parkinson's disease, Huntington's disease, and amyotrophic lateral sclerosis (13).

High doses of MSG administered to newborns or maternal are associated with obesity in animal models (14), probably through their effects on immature neurological mechanisms that control food intake and energy expenditure (15). Similarly, increased serum triglyceride, fasting glucose, and insulin levels are associated with increasing dietary MSG intake in rodent models, which are indicators of metabolic disorders (16).

The various mechanisms that MSG may cause male reproductive dysfunctions include spermatogenic alteration leading to low sperm count, high sperm abnormality, reduced live sperm, oxidative damage, histological alteration, and gonadotropin imbalance that ultimately results in male reproductive abnormalities (17).

Material and methods

From the Suez Canal University, Faculty of Veterinary Medicine's laboratory animal household, 30 healthy male Sprague-Dawley rats (weighing 120–150 g) were taken. Rats were housed in separate metal animal cages, five to a cage, with controlled humidity (55–60%), temperature (20–24° C), and dietary requirements. Rats had unlimited access to food and water (18). Two primary groups of rats were randomly assigned; the initial group, designated as Control one (group A) (n=15), distilled water was delivered orally once per day for three months. MSG is the second group (group B) (n=15), got MSG via stomach intubation once daily for three months at a dose of 15 mg/kg B.W. The weekly body weight was

used for adjust the MSG dosage. The study method's protocol was followed in accordance with the recommendations of the faculty of medicine's ethics committee at Suez Canal University in Egypt. The rats were slaughtered at the conclusion of the third month, and samples of blood were taken from the medial canthus of the eyes of the two groups.

Blood Samples

Blood was collected from two samples. There were two divisions of the samples. The first division was placed in chemically free tubes with anticoagulant (EDTA) for measuring glycated haemoglobin. While the second division was collected in centrifuge tubes without anticoagulant for serum separation, The blood samples were centrifuged at 3000 rpm for 15 minutes after clotting, and the clear serum was carefully aspirated into chemically free and clean tubes for the analysis of biochemical parameters.

Biochemical studies

Total cholesterol and triacylglycerol were determined using the procedures described by Allain et al. (19) and Siedel et al. (20), respectively. Fasting blood glucose (FBG), glycated hemoglobin (HbA1C) (%), and fasting blood insulin (FINS) according to Tietz (21), Zander et al. (22) and Sapin (23), respectively. HOMA-IR calculation in accordance with Simental-Mendía et al. (24). serum ALT and AST assay in accordance with ECCLS (25), and creatinine and urea assay in accordance with Jaffé (26) and Rock (27), respectively. Leptin, tumour necrosis factor alpha (TNF- α), and interleukin-6 (IL-6) levels were measured using the techniques described by Landt et al. (28), Feldmann and Maini (29), and Taga and

Kishimoto (30), respectively. total testosterone was determined according to Thienpont et al (31). Follicle Stimulating Hormone (FSH), Luteinizing hormone (LH) and Prolactin hormone (PRL) were determined according to Bablok et al (32).

Statistical analysis

Independent sample t-test was used for statistical analysis using Windows SPSS, version 20. Results were showed as mean \pm SE. $P < 0.001$ was regarded as statistically highly significant, and $P < 0.05$ as statistically significant.

Results and discussion

Obesity is associated with an array of illnesses, including T2D, IR, dyslipidemia, and cardiovascular disease (CVD). It is a major contributing factor to the emergence of metabolic disorders like insulin resistance, metabolic syndrome, glucose intolerance, low-grade inflammation, and male infertility (33). Obesity is described as having extra body fat or adipose tissue as a result of consuming too much food and/or using less energy (34). It is defined by an abnormal, excessive storage of energy in the form of fat in adipose tissue (35), which is caused by a malfunction of the brain's satiety centre, an imbalance between energy intake and usage, and hereditary differences (36).

Due to MSG's dual functions as a taste stimulation and a taste bud neuromodulator, use of the substance has expanded globally in recent years. MSG's use as a flavouring agent has been linked in numerous studies to the development of the metabolic syndrome and obesity in both humans and animals (37).

Table 1. Effects of MSG on body weight, total cholesterol, triacylglycerol and glucose homeostasis

Parameters	Control group (group A)	MSG group (group B)
Average body weight (BW/g)	190 ^b ±6.7	342 ^a ±19.24 ^{**}
Total Cholesterol (mg/dl)	77.00 ^b ±1.96	143.75 ^a ±8.98 ^{**}
Triacylglycerol (mg/dl)	54.00 ^b ±4.10	161.75 ^a ±15.73 ^{**}
Fasting blood glucose (mg/dl)	76.25 ^b ±5	139.25 ^a ±3.48 ^{**}
Glycated hemoglobin HbA1c (%)	4.43 ^b ±0.22	6.63 ^a ±0.13 ^{**}
Fasting blood insulin (ng /ml)	6.25 ^b ±0.32	19.25 ^a ±0.48 ^{**}
HOMA –IR	1.00 ^b ±0.08	5.85 ^a ±0.25 ^{**}

Represented data mean ± SEM. ^{**}Highly significant difference at p<0.001

The results in Table1 showed that group B exhibited significantly high increase (p<0.001) in body weight, total cholesterol, triacylglycerol, Fasting blood glucose, HbA1c, Fasting blood insulin and HOMA –IR when compared to the group A. MSG stimulated orosensory receptors, which led to an increase in body weight and also had a beneficial impact on the appetite centre, which led to an increase in weight due to an increase in food palatability (38).MSG may be neurotoxic by increasing energy intake, causing damage to the hypothalamic arcuate nucleus, which is essential for the control of metabolic balance and insulin secretion and action and impair leptin action by disrupting the hypothalamus signalling cascade, leading to leptin resistance associated with obesity (39). By making food more palatable and interfering with the leptin-mediated hypothalamic signalling flow, MSG use disrupts the body's sense of energy balance and ultimately results in obesity (40).

High significant increase in total cholesterol and triacylglycerol (P< 0.001) might be attributed to the lipid peroxidation of cell membranes, release of free fatty acids into the circulation from adipose tissue, a rise in acetyl CoA concentrations, which increases cholesterol synthesis (41).

Highly significant rise in the levels of insulin, HBA1C, and blood sugar in addition to HOMA-IR demonstrating that type 2 diabetes, obesity, hyperglycemia, hyperlipidemia, and insulin resistance are all caused in mice by parenteral MSG administration. The increase in blood sugar levels can be attributed to the liver's acceleration of glycogenolysis and gluconeogenesis as well as a temporary loss of the pancreas' endocrine functions (42).

The possibility of a worsening of glucose tolerance in rats after MSG administration has been raised. Even under the hyperinsulinemic conditions seen in mice treated with MSG, the altered glucose tolerance may be attributable to impaired cellular insulin sensitivity. Cells may

switch to pathways that promote gluconeogenesis in hyperinsulinemic circumstances to make up for the increased insulin secretion (43). Hugues et al. (44) also demonstrated the occurrence of hyperinsulinemia/hyperglycemia in MSG-

treated rats. Despite the fact that insulin secretion increased to a higher degree, they discovered that glutamate did not lower blood glucose levels. They said glutamate was the cause of insulin resistance.

Table 2. Effects of MSG on kidney function tests and liver enzymes

Parameters	Control group (group A)	MSG group (group B)
Alanine transaminase (ALT) (U/L)	38.75 ^b ±1.11	71.25 ^a ±3.12*
Aspartate transaminase (AST) (U/L)	121.75 ^b ±5.45	140.75 ^a ±3.25*
Creatinine (mg/dl)	0.50 ^b ±0.06	0.62 ^a ±0.06*
Urea (mg/dl)	20.50 ^b ±2.1	36.00 ^a ±1.22*

Represented data mean ± SEM. * significant difference at p<0.05

The results in Table2 showed that group B exhibited a significant increase in serum ALT and AST activity than group A (P<0.05). This could be attributable to hepatocellular impairment and liver function alterations, which led to the release of more intracellular enzymes into the blood (45).

Elevation level of creatinine and urea can be attributed to impairment of kidney function which may be brought on by the impact of MSG on kidney tissues (46). Additionally, blood levels of urea and creatinine elevated when the kidney's capacity to filter bodily fluid decreased (47).

Table 3. Effects of MSG on Leptin, TNF- α and Interleukin-6

Parameters	Control group (group A)	MSG group (group B)
<i>Leptin</i> (pg/ml)	881.25 ^b ±13.90	1435.00 ^a ±25.33**
<i>TNF-α</i> (pg/ml)	187.50 ^b ±5.95	395.00 ^a ±10.41**
<i>Interleukin-6</i> (IL-6) (pg/ml)	86.25 ^b ±4.03	248.25 ^a ±13.53**

Represented data mean ± SEM. **Highly significant difference at p<0.001

Table3. showed that MSG-obese rats had a high significant levels of leptin (P<0.001) than control rats (group A). This is in agreement with Perello et al. (48) who found that rodents given monosodium L-glutamate (MSG), which destroys between 80% and 90% of the neurons in the arcuate nucleus of the hypothalamus at neonatal age develop neuroendocrine and

metabolic abnormalities, leading to a phenotype of adiposity characterised by GH deficiency, hyperinsulinemia, and hyperleptinemia because of leptin resistance.

Compared to rats in control groups, MSG-obese animals showed a highly significant rise in TNF- α and IL-6 levels (P<0.001). This is in

line with other studies that have demonstrated that persistent pro-inflammatory pathway activation in insulin target cells might result in obesity linked with IR. Therefore, it has been found that people with IR and diabetes have high levels of IL-6, TNF- α , and C-reactive

protein, among other pro-inflammatory mediators (CRP). Additionally, obese, diabetic, and insulin resistant animals have higher levels of TNF- α in their blood and adipose tissue (49).

Table 4. Effects of MSG on sex hormones

Parameters	Control group (group A)	MSG group (group B)
Total Testosterone (ng/mL)	3.38 ^a ±0.18	1.58 ^b ±0.25*
Follicle-Stimulating Hormones (FSH) (mIU/mL)	2.93 ^b ±0.17	6.38 ^a ±0.44*
Luteinizing Hormones (LH) (mIU/mL)	2.25 ^b ±0.25	3.50 ^a ±0.29*
Prolactin(PRL) (mIU/mL)	4.90 ^a ±0.47	4.18 ^a ±0.34*

Represented data mean \pm SEM. * significant difference at p<0.05

The results in Table4 illustrated that MSG obese rats exhibited a significant lowering in total Testosterone (P<0.05) levels comparative to rats in control. The decrease in total testosterone is brought on by the breakdown of the neurones in the hypothalamus gland as well as the ensuing disruption of the hypothalamic-pituitary-testicular axis (50). The hypothalamic-pituitary-testis regulatory axis, which controls testicular Leydig cells' steroidogenesis can be disrupted as a result of neuronal losses in the hypothalamus, according to a previous study that found that administering MSG to rats and mice could cause damage to their neurons (51). According

to research by Fui et al. (52), overweight and moderate obesity are mostly linked to declines in total testosterone, especially in younger males. Total testosterone levels decline primarily as a result of decreased sex hormone binding globulin (SHBG) caused by obesity-related hyperinsulinemia. De Angelis et al. (53) reported that once obesity develops, in addition to insulin resistance, adiposity has been reported to have an impact on the hypothalamus-pituitary-gonadal axis, which may directly influence testosterone production. This suggests that the current decrease in plasma testosterone levels may be caused by insulin resistance.

Table 5. Pearson's correlation analysis of data

Correlations							
		HOMA-IR	HbA1c	FSH	LH	B_Weight	Testosterone
B_Weight	Pearson Correlation	.989**	.962**	.935**	.800*	1	-.930**
	Sig. (2-tailed)	.000	.000	.001	.017		.001
	N	8	8	8	8	8	8
HOMA-IR	Pearson Correlation	1	.965**	.951**	.860**	.989**	-.927**
	Sig. (2-tailed)		.000	.000	.006	.000	0.001

	N	8	8	8	8	8	8
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**Correlation is significant at the 0.01 level (2-tailed).

* Correlation is significant at the 0.05 level (2-tailed).

The current study demonstrated a significant rise in LH and FSH levels following MSG administration Table4. This could mean that MSG has disrupted or damaged the hypothalamus' hormonal secretion circuits (54). In response to hypothalamic gonadotropin releasing hormone, the pituitary releases both FSH and LH (GnRH). While FSH encourages Sertoli cells to assist spermatogenesis and inhibin B secretion, which inhibits FSH secretion, LH stimulates the secretion of testosterone from Leydig cells. In addition to promoting sperm production, testosterone acts as a feedback mechanism between the brain and pituitary to control GnRH secretion (55).

Because testosterone's negative feedback loop on the brain's and pituitary's secretory activities is reduced, higher levels of GnRH and FSH/LH secretion may be the cause of the high plasma concentrations of luteinizing hormone and follicle stimulating hormone after exposure to MSG in this study (56).

The observed increase in FSH level demonstrated that spermatogenesis is disrupted, resulting in an excess of signals to the brain to secrete FSH. Following MSG administration, both unstable levels of FSH and LH suggested that this substance may have damaged or disturbed the hormonal secretion pathways in the hypothalamus (54).

Table4 also showed that there is no significant change in circulating prolactin levels between MSG group and control group. These results correspond to the results obtained by Ernst et al. (57). They reported that prolactin directly promotes excess energy toward the visceral fat depot and found no evidence of a connection

between the level of blood prolactin and the degree of obesity or its associated metabolic disturbances or any systematic changes in basal concentrations of the hormone after massive weight loss.

Conclusion

Obesity is a collection of connected metabolic features that directly raising the risk of type 2 diabetes, insulin resistance, dyslipidemia and male infertility. One of the most often utilised food additives consumed with commercially processed meals is MSG. In addition to having a negative impact on reproductive health, prolonged, excessive use of MSG can cause obesity, kidney damage, hepatotoxicity, and metabolic disorders.

Conflict of Interests

The authors declare that they have no conflicts of interest regarding the publication of this paper.

Reference

- Dutta S, Biswas A, Sengupta P. Obesity, endocrine disruption and male infertility. Asian Pacific Journal of Reproduction 2019; 8: 5. 195-202.
- Du Plessis SS, Cabler S, McAlister DA, Sabanegh E, Agarwal A. The effect of obesity on sperm disorders and male infertility. Nat Rev Urol 2010; 7: 3. 153.
- Caballero AE, Bousquet-Santos K, Robles-Osorio L, Montagnani V, Soodini G, Porramatikul S, Hamdy O, Nobrega ACL, Horton ES. Overweight Latino children and adolescents have marked endothelial dysfunction and subclinical vascular inflammation in association with excess

- body fat and insulin resistance. *Diabetes Care*, 2008, 31.3: 576-582.
- Sethi JK, Vidal-Puig AJ. Adipocyte biology. Adipose tissue function and plasticity orchestrate nutritional adaptation. *Journal of lipid research*, 2007; 48: 6. 1253-1262.
- Giannessi D, Maltinti M, Del Ry S. Adiponectin circulating levels: a new emerging biomarker of cardiovascular risk. *Pharmacological research*, 2007; 56: 6. 459-467.
- Alarcon-Aguilar FJ, Almanza-Perez J, Blancas G, Angeles S, Garcia-Macedo R, Roman R, Cruz M. Glycine regulates the production of pro-inflammatory cytokines in lean and monosodium glutamate-obese mice. *European journal of pharmacology*, 2008, 599.1-3: 152-158.
- Blache D, Zhang S, Martin G, Fertility in male sheep. Modulators of the acute effects of nutrition on the reproductive axis of male sheep. *Reproduction*, 2003; 61: 387–402.
- Dutta, S., Sengupta, P., Muhamad, S., Male reproductive hormones and semen quality. *Asian Pacific Journal of Reproduction*, 2019; 8: 5. 189–194 .
- Stanska, K., Krzeski, A., The umami taste: From discovery to clinical use. *Otolaryngologia Polska*, 2016; 70: 4. 10–15.
- Helal EG, El-Sayed RA, El-Gamal MS. Assessment of the Physiological Changes Induced by Sodium Nitrite, Annatto or Mono Sodium Glutamate in Male Albino Rats. *Egyptian Journal of Hospital Medicine*, 2017; 67: 330-335.
- Oyeniran DO, Ojewale AO, Akingbade AM, Dare BJ, Adelaja MA, Oseni LO. The ameliorative potentials of aqueous leaf extract of *Amaranthus hybridus* on monosodium glutamate-induced testicular damage in Wistar rats. *JBRA Assisted Reproduction*, 2022; 26: 3. 469.
- Yousef M, El-Nassag D, Gasser M, Ibrahim A (2019). Potential Protective Effects of Propolis against Hepatotoxicity and Nephrotoxicity Induced by Monosodium Glutamate in Rabbits. *Alexandria Science Exchange Journal*, 2019, 40: 30-42.
- El-Sawy HBI, Soliman MM, El-Shazly SA, Ali HA. Protective effects of camel milk and vitamin E against monosodium glutamate induced biochemical and testicular dysfunctions. *Prog. Nutr*, 2018; 20: 1. 76-85.
- Nagata M, Suzuki W, Iizuka S, Tabuchi M, Maruyama H, Takeda S, Aburada M, Miyamoto K: Type 2 diabetes mellitus in obese mouse model induced by monosodium glutamate. *Exp Anim*, 2006; 55:109–115.
- Bölükbaş F, Öznurlu Y. The determination of the effect of in ovo administered monosodium glutamate on the embryonic development of thymus and bursa of Fabricius and percentages of alpha-naphthyl acetate esterase positive lymphocyte in chicken. *Environmental Science and Pollution Research*, 2022; 1-11.
- El Tabbal J. Monosodium glutamate in a type 2 diabetes context: A large scoping review. *Regulatory Toxicology and Pharmacology*, 2022; 133: 105223.
- Kayode OT, Rotimi DE, Kayode AAA, Olaolu TD, Adeyemi OS. Monosodium Glutamate (MSG)-induced male reproductive dysfunction: a mini review. *Toxics*. 2020; 8: 1. 7.
- Egbonu A, Osakwe O. Effects of high monosodium glutamate on some serum markers of lipid status in male Wistar rats.

- Journal of Medicine and Medical Sciences. 2011; 2: 1. 653-6.
- Allain CC, Poon LS, Chan CS, Richmond W, Fu PC. Enzymatic determination of total serum cholesterol. *Clinical chemistry*. 1974; 20: 4. 470-5.
- Siedel J, Schmuck R, Staepels J, Town M. Long term stable, liquid ready-to-use monoreagent for the enzymatic assay of serum or plasma triglycerides (GPO-PAP method). AACC meeting abstract 34. *Clin Chem*. 1993; 39: 1127.
- Tietz N. *Clinical guide to laboratory tests*. 4-th ed. Ed Wu ANB USA. 2006.
- Zander R, Lang W, Wolf HU. Alkaline haematin D-575, a new tool for the determination of haemoglobin as an alternative to the cyanhaemoglobin method. I. Description of the method. *Clinica chimica acta*. 1984; 136: 1:83-93.
- Sapin R. Insulin assays: previously known and new analytical features. *Clinical laboratory*. 2003; 49: 3-4. 113-21.
- Simental-Mendía LE, Rodríguez-Morán M, Guerrero-Romero F. The product of fasting glucose and triglycerides as surrogate for identifying insulin resistance in apparently healthy subjects. *Metabolic syndrome and related disorders*. 2008; 6: 4. 299-304.
- ECCLS. Determination of the catalytic activity concentration in serum of L-alanine aminotransferase (EC 2.6.1.2, ALAT)". 20:204-. *Klin Chem Mitt*. 1989; 211:20:204.
- Jaffé M. Ueber den Niederschlag, welchen Pikrinsäure in normalem Harn erzeugt und über eine neue Reaction des Kreatinins. *Zeitschrift für physiologische Chemie*. 1886; 10: 5. 391-400.
- Rock RCW, W.G.; Jennings, C.D. Nitrogen metabolites and renal function. In: Tietz NW, ed. *Fundamentals of Clinical Chemistry*. Philadelphia: WB Saunders. 1987;3rd ed:669–704.
- Landt M, Gingerich RL, Havel PJ, Mueller WM, Schoner B, Hale JE, Heiman ML. Radioimmunoassay of rat leptin: sexual dimorphism reversed from humans. *Clinical chemistry*, 1998; 44: 3. 565-570.
- Feldmann M, Maini RN. Anti-TNF α therapy of rheumatoid arthritis: what have we learned?. *Annual review of immunology*, 2001; 19: 1. 163-196 .
- Taga T, Kishimoto T. Gp130 and the interleukin-6 family of cytokines. *Annual review of immunology*, 1997; 15: 1. 797-819.
- Thienpont L, Verhseghe PG, Van Brussel KA, De Leenheer AP. Estradiol-17- β Quantified in Serum by Isotope Dilution-Gas Chromatography-Mass Spectrometry. *Clinical chemistry*, 1988; 34; 10. 2066-2069.
- Bablok W, Passing H, Bender R, Schneider B. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. *journal of clinical chemistry and clinical biochemistry*, 1988; 26: 11. 783-790.
- Swan SH, Elkin EP, Fenster L. The question of declining sperm density revisited: an analysis of 101 studies published 1934-1996. *Environmental health perspectives*, 2000; 108: 10. 961-966.
- Aitlhadj L, Ávila DS, Benedetto A, Aschner M, Stürzenbaum SR. Environmental exposure, obesity, and Parkinson's disease: lessons from fat and old worms.

- Environmental health perspectives, 2011; 119: 1. 20-28.
- Cohen P, Spiegelman BM. Cell biology of fat storage. *Molecular biology of the cell*, 2016; 27: 16. 2523-2527.
- Tu Q, Xiao LD, Ullah S, Fuller J, Du H. Hypertension management for community-dwelling older people with diabetes in Nanchang, China: study protocol for a cluster randomized controlled trial. *Trials*, 2018; 19: 1. 385.
- Olguin MC, Posadas MD, Revelant GC, Marinozzi D, Labourdette V, Venezia MR. Monosodium glutamate affects metabolic syndrome risk factors on obese adult rats: a preliminary study. *Journal of Obesity and Weight-Loss Medication*, 2018; 4:1. 23.
- Hermanussen M, Garcia AP, Sunder M, Voigt M, Salazar V, Tresguerres JA. Obesity, voracity, and short stature: the impact of glutamate on the regulation of appetite. *Eur. J. Clin. Nutr*, 2006; 60: 1. 25- 31.
- Wang CX, Zhang Y, Li QF, Sun HL, Chong HL, Jiang JX, Li QC. The Reproductive Toxicity of Monosodium Glutamate by Damaging GnRH Neurons Cannot Be Relieved Spontaneously Over Time. *Drug Design, Development and Therapy*, 2021; 15: 3499.
- Araujo TR, Freitas IN, Vettorazzi JF, Batista TM, Santos-Silva JC, Bonfleur ML. Benefits of L-alanine or L-arginine supplementation against adiposity and glucose intolerance in monosodium glutamate-induced obesity. *European journal of nutrition*, 2017; 56: 6. 2069-80.
- Abu Aita NA, Mohammed FF. Effect of marjoram oil on the clinicopathological, cytogenetic and histopathological alterations induced by sodium nitrite toxicity in rats. *Global Veter*, 2014; 12: 5. 606-616.
- Al-Shinnawy MS, Elkattan NA. Assessment of the changes in some diagnostic parameters in male albino rats fed on an Azo Dye. *International Journal of Environmental Science and Technology*, 2013; 4: 85-92.
- Okwudiri OO, Sylvanus AC, Peace IA. Monosodium glutamate induces oxidative stress and affects glucose metabolism in the kidney of rats. *International Journal of Biochemistry Research & Review*, 2012; 2: 1. 1.
- Hugues C, Eric R, Gyslaine B, Chevassus H, Renard E, Bertrand G, Mourand I, Puech R, Molinier N, Bringer J. Effects of oral monosodium (L)-glutamate on insulin secretion and glucose tolerance in healthy volunteers. *British journal of clinical pharmacology*, 2002; 53: 641-643.
- Tawfek N, Amin H, Abdalla A, Fargali S. Adverse effects of some food additives in adult male albino rats. *Current Science International*, 2015; 4: 525-537.
- Helal EG, El-Sayed RA, El-Gamal MS. Assessment of the Physiological Changes Induced by Sodium Nitrite, Annatto or Mono Sodium Glutamate in Male Albino Rats. *Egyptian Journal of Hospital Medicine*, 2017; 67: 330- 335.
- Tawfik SA, Al-Badr N. Adverse Effects of Monosodium glutamate on liver and kidney functions in adult rats and potential protective effect of vitamins C and E. *Food and Nutrition Sciences*, 2012; 3: 651-659.
- Perello M, Moreno G, Gaillard RC, Spinedi E. Glucocorticoid-dependency of increased adiposity in a model of hypothalamic obesity. *Neuroendocrinology Letters*, 2004; 25: 119–126.
- Mahmoud AM, Abd El-Twab SM, Abdel-Reheim ES. Consumption of polyphenol-rich *Morus alba* leaves extract attenuates early diabetic retinopathy: the underlying

- mechanism. *European journal of nutrition*, 2017; 56: 4. 1671-1684.
- Ochiogu I, Ogwu D, Uchendu C, Okoye C, Ihedioha J, Mbegbu E. Effects of monosodium-L-glutamate administration on serum levels of reproductive hormones and cholesterol, epididymal sperm reserves and testicular histomorphology of male albino rats. *Acta Veterinaria Hungarica*, 2015; 63: 1. 125-139.
- Igwebuike UM, Ochiogu IS, Ihedinihu BC, Ikokide JE, Idika IK. The effects of oral administration of monosodium glutamate (MSG) on the testicular morphology and cauda epididymal sperm reserves of young and adult male rats. *Veterinarski arhiv*, 2011; 81:4. 525-534.
- Fui MNT, Dupuis P, Grossmann M. Lowered testosterone in male obesity: mechanisms, morbidity and management. *Asian journal of andrology*, 2014; 16: 2. 223.
- de Angelis C, Garifalos F, Mazzella M, Menafrà D, Verde N, Castoro M, Pivonello R. Risk Factors Affecting Puberty: Environment, Obesity, and Lifestyles. In *Pediatric and Adolescent Andrology* (pp. 171-200). Springer, Cham, 2021.
- Jubaidi FF, Mathialagan RD, Noor MM, Taib IS, Budin SB. Monosodium glutamate daily oral supplementation: Study of its effects on male reproductive system on rat model. *Systems biology in reproductive medicine*, 2019; 65: 3. 194-204.
- Oduwole OO, Peltoketo H, Huhtaniemi IT. Role of follicle-stimulating hormone in spermatogenesis. *Frontiers in endocrinology*, 2018; 9: 763.
- Dimitriadis F, Adonakis G, Kaponis A, Mamoulakis C, Takenaka A, Sofikitis N. Pre-testicular, testicular, and post-testicular causes of male infertility. *Endocrinology of the testis and male reproduction*. Cham: Springer, 2017; 1-47.
- Ernst B, Thurnheer M, Schultes B. Basal serum prolactin levels in obesity--unrelated to parameters of the metabolic syndrome and unchanged after massive weight loss. *Obesity surgery*, 2009; 19: 1159-1162.