Quality of Agar Gracilaria verrucosa SeaWeed with Different Density in Polyculture System

Andi Rahmad Rahim

Department of Aquaculture, Faculty of Agriculture, Universitas Muhammadiyah Gresik, Indonesia, andirahmad@umg.ac.id

Dwi Retnaningtyas Utami

Department of Food Technology, Faculty of Agriculture, Universitas Muhammadiyah Gresik, Indonesia

Setyo Budi

Department of Agrotechnology, Faculty of Agriculture, Universitas Muhammadiyah Gresik, Indonesia

Abstract

The polyculture system was used to increase the productivity of extensive brackish water ponds to produce optimal agar with varying densities of three commodities: milkfish, Vannamei shrimp, and Gracilaria verrucosa. This study aims to obtain the optimal density of the three commodities in extensive brackishwater ponds with polyculture systems to produce the best agar quality for G. verrucosa. The research was conducted in the expanse of the Polyculture System Extensive brackishwater Pond in Lamongan Regency. The study used a Completely Randomized Design (CRD) with 3 density treatments (milkfish m-2 : Vannamei shrimp m-2 : G. verrucosa g m-2), and 3 replicates: A (10 : 10 : 250), B (20 : 20 : 500), and C (30 : 30 : 1000). Statistical analysis uses one way ANOVA (Analysis of Variance), while Tukey's HSD (Honestly Significant Difference) and Path Analysis use Pearson Correlation. The results showed that the best density obtained in treatment A was significantly different from treatments B and C in producing Specific Growth Rate, Absolute Weight, Absolute Length, Carbon Content, and quality of agar rendementing the best of seaweed G. verrucosa. From the path analysis, CNP nutrients and the growth of G. verrucosa seaweed have a strong and very strong influence to improve the quality of agar rendementing G. verrucosa seaweed.

Keywords: Agar, Extensive, Gracilaria verrucosa, Density, Polyculture.

INTRODUCTION

Government policy through the aquaculture revitalization program places shrimp, milkfish, and seaweed as superior commodities (Directorate General of Aquaculture, 2018). Polyculture is one system that is expected to increase pond production and revive the production of shrimp, milkfish, and seaweed as superior commodities. The polyculture system is a way of cultivating various fish species with different ecological niches to increase the productivity of traditionally managed ponds. The advantages of this system are it can minimize the risk of crop failure, improve the growth of cultivated commodities, produce quality seafood products, and provide added value to fish farmers through diversification of aquaculture products (Martínez-Porchas et al. 2010, Israel et al. 2017, Pantjara B. M and Mangampa. 2010). One of the problems faced in the polyculture system is determining the density of fishery commodities that are most effective in utilizing natural feed available in ponds. To utilize the natural food contained in the pond effectively, of course the combination of commodity species must be able to live together without causing competition for food or space (Kristanto A et al., 2013). The right density of 3 commodities-shrimp, milkfish, and seaweedin polyculture media are needed to produce optimal production. Gracilaria verrucosa seaweed utilizes the metabolism of milkfish and Vannamei shrimp as a source of nutrients to improve the quality of agar seaweed. The polyculture system is a beneficial system for seaweed because the waste and food residues from milkfish and Vannamei shrimp in the form of detritus are converted into nutrients through a diffusion process to accelerate the growth of seaweed (Samidjan et al., 2018).

Seaweed G. verrucosa is one of the agar producers that has been successfully cultivated in Indonesian ponds (Faturrahman et al., 2011). G. verrucosa contains agar with good gel strength in abundant quantities (Sornalakshm, 2017). Agar is a mixture of polysaccharides mainly found in red algae's matrix and cell walls and is usually extracted from species of algae belonging to the family Gracilariaceae (Painter 1983, Niu et al. 2013). In everyday life, gelatin is used as a food ingredient. Agar is thickening and gelling hydrocolloid used as a food additive and the demand for agar is increasing due to the increased consumption of processed foods (Ollando et al. 2019, Valderrama D. and Cai J. 2014). Whereas in industsry, agar is used as an additive in food canneries, pharmaceuticals, cosmetics, paints, and textiles (Istigomawati and Kusdarwati 2010. Marinho-Soriano E et al. 2002. Niu et al. 2013).

Agar quality is one of the important requirements to increase its selling value. Therefore, the factors that affect the agar content in Gracilaria need to be considered economically feasible (Sornalakshm, 2017). In order to achieve maximum production of seaweed, several important factors are needed, one of which is the density between seaweed, milkfish, and Vannamei shrimp. The right density between milkfish, Vannamei shrimp, and seaweed in a polyculture system will affect the growth of seaweed, where one of the efforts to improve the quality of seaweed is to increase its growth. Appropriate densities can increase business profits in polyculture systems and achieve sustainable cultivation.

One way to increase production is to adjust the density level (Isroni et al., 2020). However, information about the appropriate density in implementing polyculture cultivation of milkfish, Vannamei shrimp, and seaweed in ponds is still not widely known by the public. It is hoped that optimizing the density of the three commodities in the polyculture system can effectively utilize the ecological space of pond waters. So, it is necessary to determine the optimal density of the three leading commodities of milkfish, Vannamei shrimp, and seaweed in extensive ponds with a polyculture system to improve the quality of agar G. verrucosa seaweed.

Material and methods

Sampel Collection

Seedlings of Gracilaria verrucosa (red algae) from tissue culture were obtained from the Polyculture Pond of Pulokerto Village, Kraton District, Pasuruan Regency, East Java Province, Indonesia. Seaweed seeds were 14 days old, clean, fresh, and free from other types. The selected seaweed seeds were collected as much as 100 kg, packed in alkaline conditions, and avoided from the hot sun. Seaweed seeds were transported by motor boat to the research location with a distance of 125 km. After arriving at the research site, the seedlings were adapted for 48 hours before being stocked into the Research Media.

Research Place

This research was conducted in the Polyculture Extensive brackishwater Pond of Soko Village, Glagah District, Lamongan Regency, East Java Province, Indonesia, with an area of 21.3 ha. This research was carried out for 42 days in one of the polyculture ponds with an area of 0.5 ha, by installing a culture container made of tarpaulin inside the pond.

Fig. 1. Polyculture System Extensive Pond Expanse.



Research design

This study used a Completely Randomized Design (CRD) with 3 density treatments (milkfish m-2 : Vannamei shrimp m-2 : G. verrucosa g m-2) and 3 replicates, namely Treatment A (10 : 10 : 250), B (20 : 20 : 500), and C (30 : 30 : 1000) with a total of 9 treatments (Fig. 2a).

This study used 9 experimental units of 1 x 1 x 1 m tarpaulin (Fig. 2a), with a water level of 50 cm, and with the initial weight and length of G. verrucosa seaweed at 10 g and 8.0 - 11.5 cm (Fig. 2b). The source of brackishwater pond

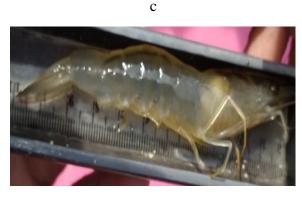
came from the estuary of the Solo River that was a source of brackish water with a salinity level of 10 - 20 g l-1. Shrimp and milkfish seeds came from brackishwater pond in Soko Village, Glagah District, Lamongan Regency. The seeds came from the artificial spawning process and were selected based on similar size, intact body shape, and active swimming. Every 500 seeds were collected in controlled media. The initial weight and length of milkfish and Vannamei shrimp stocked in the research media were milkfish 45.8 - 48.8 g and 17.14 – 17.48 cm, Vannamei shrimp 6.6 – 10.0 g and 10.4 - 10.6 cm (Fig. 2c and 2d).

Fig. 2. (a) Research Design (b) Seaweed Seeds of G. verrucosa (c) Milkfish (d) Vannamei shrimp.









d

Seaweed Nutrient Analysis

The measurement of the carbon level of G. verrucosa seaweed was using the Gravimetric method to determine of the ash level and water level converted to carbon level. The nitrogen content of G. verrucosa using the Kjeldahl method is destroyed with concentrated sulfuric acid with Zn granules as the catalyst, then collected and titrated with the help of an indicator (William Horwitz et al., 2006). The Phosphorus content of G. verrucosa using the 'UV-Vis spectrophotometry' method uses light passed through a container containing a solution, producing a spectrum (Lambert Beer's law).

Quality of Agar Seaweed Analysis

The agar level of the G. verrucosa seaweed rendement was measured using the weight of the raw material in the form of dry seaweed flour divided by the dry weight of the sample before being made into flour and expressed in a percent; the higher the rendement, the higher the output produced. Viscosity (thickness) of G. verrucosa was a processed agar-agar powder that had been heated at a temperature of 75oC, then its thickness was measured by using a Brookfield viscosimeter, the unit of viscosity was in the form of centipoises (cps). The gel strength of G. verrucosa is the maximum load required to break the polymer matrix in the loaded area, the seaweed gel solid formed from the heating process at the 75oC temperature and allowed to stand for one day until a gel solid is formed, the gel strength measurement is carried out using a Curd meter with units of g/cm2 (Wil liam Horwitz et al., 2006).

Growth Analysis

Measurement of the growth of G. verrucosa seaweed was carried out every week for 42 days of observation using an analytical balance measuring instrument with an accuracy of 0.0 g and a measuring instrument with an accuracy of 0.0 cm.

Measurement of absolute weight with the formula of (Fortes, 1989):

Absolute Weight (g) = Final Weight of Observation (g) – Initial Weight of Stocking (g).

Specific Growth Rate with the formula of (Dawes, C.J., Lluis, A.O. Trono, 1994):

Specific Growth Rate (% Day-1) =

Absolute Length Growth with the formula (Effendi, 1997):

Absolute Length Increase (cm) =

Final Length (cm) – Initial Length (cm).

Water Quality Analysis

The measurement of prawn farm water quality parameters is carried out by in situ and ex situ bases. In situ refers to temperature (oC) (thermometer), pH (digital pH meter), dissolved oxygen (ppm) (dissolved oxygen meter), salinity (g l-1) (hand refractometer), brightness (cm) (Secchi disk). Ex situ refers to the content of Carbon, Nitrogen, and Phosphorus (ppm) (spectrophotometer with nesslerization method) (Colman, 2010).

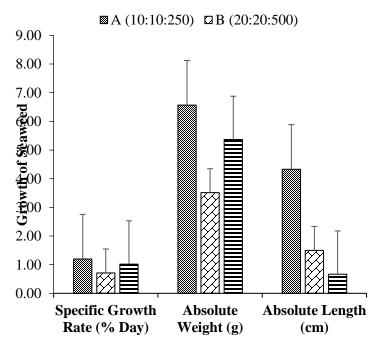
Statistical Analysis

Data analysis of this study used one way ANOVA (Analysis of Variance) to see the significant effect of different density treatments (milkfish m-2 : Vannamei shrimp m-2 : G. verrucosa g m-2) on absolute weight (g), specific growth rate (% day-1), absolute length (cm), carbon, nitrogen, and phosphorus content (%), agar rendement quality (%), viscosity (cps), gel strength (g cm-2) of G. verrucosa seaweed in extensive prawn farms with polyculture systems. If it gave a significant effect (p < 0.05), then it was proceeded with the Tukey's HSD test to see significant differences between treatments in each parameter, with a 95% confidence level. Path analysis was used to see how big the correlation between CNP nutrient content parameters and the growth of G. verrucosa seaweed with a polyculture system in increasing the rendement of G. verrucosa seaweed which is the final product of high-value agar products; the model was generated from Pearson analysis (Product Moment Correlation). The correlation value ranges from 0.0 to 1.0; the closer to number one, the stronger the relationship between the observed variables (Sugiyono, 2010).

RESULTS AND DISCUSSIONS

Growth of Seaweed

Figure 3. Growth of Seaweed (mean \pm SE, n = 9) of G. verrucosa for different densities (milkfish m-2 : Vannamei shrimp m-2 : G. verrucosa g m-2). Observation parameters were specific growth rate, absolute weight, and absolute length. Different notation/lower case letters indicate statistical significance (P < 0.05) between density treatments of culture.

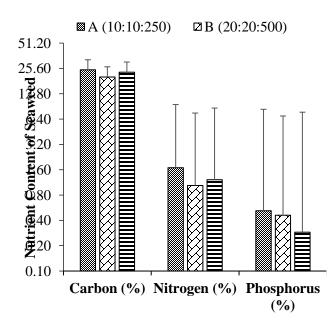


The average growth range of Gracilaria verrucosa seaweed was the specific growth rate 0.71 - 1.20 % day-1, absolute weight 3.51 -6.57 g, and absolute length 0.67 - 4.33 cm. From the Analysis of Variance (ANOVA), the provision of different densities had a significant effect on increasing the specific growth rate, absolute weight, and absolute length of G. verrucosa seaweed (p < 0.05). According to Matinfar et al (2013), the specific growth rate ranges from 3.5 to 3.7 % day-1 in Gracilaria persica. Seaweed growth in this study was influenced by different cultivation systems and initial weights. A high density of organisms will increase energy competition and affect growth. The density factor in polyculture media also influences the growth of seaweed. Shrimp

density has a significant effect on absolute weight and SGR of Gracilaria corticate. In the results of his research, the SGR range is 0.31 -1.23 % day-1 and the absolute weight is 14.92 - 73.67 g (Fourooghifard et al., 2018). The length of the thalus in G. verrucosa (Hudson) Papenfuss can reach 22.33 cm with an absolute weight of 65.91 g (Nana, S.S., 2008). The absolute length of G. verrucosa ranges from 2.5 to 3.8 cm (A.R. Rahim et al., 2016). The increase in the length of the thalus can be clearly seen from the shoots that begin to grow at the tip of the thalus, and it can reach an average length of 1.03 - 1.29 cm for 42 days (Muarif and Yala, Z.R., 2017).

Seaweed Nutrient Content

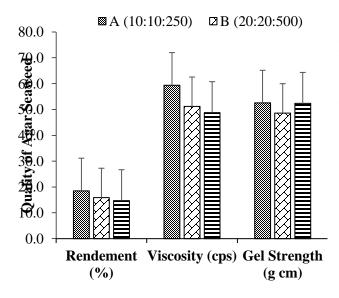
Figure 4. Nutrient Content of Seaweed (mean \pm SE, n = 9) of G. vertucosa for different densities (milkfish m-2 : Vannamei shrimp m-2 : G. verrucosa g m-2). **Observation** parameters were Carbon, Nitrogen, and Phosphorus. Different letters notation/lower case indicate statistical significance (P < 0.05) between density treatments of culture.



The average range of nutrients for G. verrucosa seaweed is Carbon 20.26 – 24.60 %, Nitrogen 1.04 - 1.69 %, and Phosphorus 0.29 - 0.52 %. From the Analysis of Variance (ANOVA), the provision of different densities had a significant effect on increasing the carbon content of G. verrucosa seaweed (p < 0.05). At the same time, the condition of different densities did not significantly affect the nitrogen and phosphorus content of G. verrucosa seaweed (p > 0.05). The carbon content of G. vertucosa is in the range of 23.53 - 29.47 % (A.R. Rahim, 2018b). The carbon content range in G. verrucosa cultivated on the coast is 21.38-24.57 % (Erlania et al., 2013). Carbon is the primary nutrient needed by seaweed in the photosynthesis process produce to carbohydrates which are the main components of seaweed (Stiger-Pouvreau et al., 2016). A.R. Rahim (2018a) reported the nitrogen range in G. verrucosa is between 0.85 - 2.02 %. The nitrogen content in the thalus tissue of G. verrucosa is 0.6 % (Rosyida et al., 2014). High nitrogen content in thalus tissue correlated with the growth of G. verrucosa seaweed (Bird et al. 1986, Rosyida et al. 2014). Nitrogen is utilized by seaweed to synthesize amino acids and proteins with the help of the enzyme nitrate reductase, which helps in the growth process (Klionsky et al., 2016). According to A.R. Rahim (2018b), the phosphorus content in G. verrucosa was 0.20 - 0.26 %. The phosphorus content of seaweed cultivated offshore ranges from 0.06 - 1.07 % (Yuniarsih et al., 2014). The phosphorus content of G. verrucosa seaweed on a laboratory scale ranged from 0.03 to 0.10 % (Mulatsih, 2015). High phosphorus levels in brackishwater ponds will support the growth of Gracilaria spp (Xu et al., 2008).

Quality of Agar Seaweed

Figure 5. Quality of Agar Seaweed (mean \pm SE, n = 9) of G. verrucosa for different densities (milkfish m-2 : Vannamei shrimp m-2 : G. verrucosa g m-2). Observation parameters were rendement, Viscosity, and gel strength. Different notation/lower case letters indicate showed statistical significance (P < 0.05) between density treatments of culture.



The results showed an average range of agar quality for G. verrucosa seaweed during this study, the rendement of 14.7 - 18.5 %, Viscosity of 48.8 - 59.4 cps, and gel strength of 48.6 - 52.4 g cm-2. Statistical test ANOVA (Analysis of Variance) giving different densities had a significant effect on improving the quality of agar rendement G. verrucosa seaweed (p < 0.05). In contrast, the provision of different densities did not significantly increase the quality of agar viscosity and gel strength of G. verrucosa seaweed (p > 0.05). According to

Mulyaningrum and Suwoyo (2018), Agar rendements obtained from G. verucossa seaweed in brackishwater pond ranged from 10.30 – 27.84 %. The polysaccharide rendement based on the mass of Gracilaria seaweed was 17.0 % (De Castro et al., 2018). The rendement of marine cultured G. verrucosa was 8.1-30 % and 14.7 %, respectively (Orosco et al. 1992, Oyieke 1993). Agar rendement obtained from Gracilaria sp. cultivated in brackishwater pond ranged from 5.768 % to 17.506 % (Yulistiana et al., 2020). Agar rendement from Gracilaria produced in brackishwater pond ranged from 24.6 - 30.6 % (Rahim, A.R., 2021). In the brackishwater pond, many nutrients are derived from the metabolic activity of polyculture organisms. It forms polysaccharides, such as agarose and agaropectin, acting as primary ingredients for creating agar (Anton, 2017). Rahim, A.R (2017), the Viscosity of G. verrucosa is 76.67 – 90.0 cps. Waluyo et al (2019), the Viscosity of Gracilaria seaweed is 201.6 cps. Wenno et al (2012), the nutrients in the waters produced from the cultivation process, the level of nutrients affect the viscosity value of seaweed. Gracilaria gel strength ranges from 50 - 300 g cm-2 and can reach 500 g cm-2 (Myco Supply, 2011). The gel strength of G. verrucosa from tissue culture started from 68.2 - 101.8 g cm-2 (Rahim, A.R et al., 2016). Waluyo et al. (2019), the power of the Gracilaria gel in brackishwater pond was 356.76 g cm-2. Gioele et al (2017), the gel strength of 3 Gracilaria species was 22.2 - 630 g cm-2. Rahim, A.R (2017), the gel strength of G. verrucosa was 40.0 - 56.6 g cm-2. This extreme difference in gel strength can be attributed to differences in location and physiological factors (Martín et al., 2013).

Water Quality

Table 1. Water Quality Parameters for different densities (Milkfish m-2 : Vannamei shrimpm-2: Seaweed G. verrucosa g m-2) during culture of G. verrucosa in PolycultureBrackishwater Pond.

	Water Quality Day 0							
Treatment	Temperature (°C)	Salinity (g l ⁻¹)	pН	Dissolved Oxygen (ppm)	Brightness (cm)	Carbon (ppm)	Nitrogen (ppm)	Phosphorus (ppm)
A (10 : 10 : 250)	31.5	15	6.54	5.5	45	4475.32	63.76	25.21
B (20:20:500)	31.5	15	6.54	5.5	45	4017.09	70.33	21.32
C (30 : 30 : 1000)	31.5	15	6.54	5.5	45	3190.24	65.32	24.51
	Water Quality Day 42							
Treatment	Temperature (°C)	Salinity (g l ⁻¹)	pН	Dissolved Oxygen (ppm)	Brightness (cm)	Carbon (ppm)	Nitrogen (ppm)	Phosphorus (ppm)
A (10 : 10 : 250)	31.6	18	6.93	4.2	43	627.43	13.89	18.30
B (20:20:500)	32.0	18	6.90	3.9	40	624.91	14.63	19.53
C (30:30: 	31.9	18	6.95	3.6	40	552.03	12.21	20.33

The average water quality range during the study was temperature 31.5 - 32.0 oC, salinity 15-18 g l-1, pH 6.54 - 6.95, dissolved oxygen 3.6 - 5.5 ppm, brightness 40 - 45 cm, carbon 552.03 - 4475.32 ppm, nitrogen 12.21 - 70.33 ppm, and phosphorus 18.30 - 25.21 ppm (Table 1).

Water quality is one of the most vital factors in seaweed cultivation activities because it can affect the growth and success of seaweed cultivation (Istiqomawati 2010, Susilowati T et al. 2012). Water quality parameters determine the development and distribution of macroalgae (Raikar et al., 2001). The growth of Gracilaria coronopifolia is positively correlated with temperatures between $15 - 35^{\circ}$ C and reaches its maximum production level at 30° C(Tsai et al. 2005, Yang et al. 2015). A suitable temperature for the development of Gracilaria lemaneiformis is between $12 - 23^{\circ}$ C. In contrast to G. lemaneiformis, the subtropical species Gracilaria tenuistipitata var. liui grows best at 20 - 30°C in brackishwater pond, but its growth rate decreases at temperatures below 15° C or above 32° C (Wu et al. 1994, Yang et al. 2015). In the study of Gracilaria fisheri, the optimum temperature of tropical seaweed in the Caribbean was found between $25 - 30^{\circ}$ C (Pakker et al., 1995). Most species of Gracilaria sp. grow well at temperatures of 20° C or above

(Bird et al. 1986, Yang et al. 2015). Water temperature controls the growth of seaweed, so it is one of the most important factors. In addition, the temperature can also affect several physiological processes in algae, such as the rate of diffusion and absorption of nutrients (Lapointe 1984, Yang et al. 2015).

The suitable salinity range for seaweed growth is 33 - 35 g l-1 with an optimal 33 g l-1. In the study of Gracilaria fisheri, the optimum salinity of seaweed in the Atlantic and Pacific oceans ranged from 15 - 30 g l-1 (Bird et al., 1986). Zhou et al (2013), studied the effect of salinity on the development and release of Gracilaria lemaneiformis carpospora, and found a range of 30 - 35 g l-1. Furthermore, Choi et al (2006), the effect of salinity on the growth of G. verrucosa and Gracilaria chorda, both species grow in a wide salinity range ranging from 5 -35 g l-1, with an optimum range of 15 - 30 g l-1. Bird et al (1986), Gracilaria spp. pale and die when the salinity is less than 15 g l-1, whereas Kumar et al (2010), Gracilaria corticata at salinity below 15 g l-1 causes the thalus to become weak. Sarkar et al (2019), Gracilaria tenuistipitata cultivation in brackishwater pond, pH 8.02 - 8.05 was obtained. Fourooghifard et al (2018), the pH obtained in the cultivation of Gracilaria corticata is s7.3 -8.7. Another study reported that a pH above 8 was optimal for Gracilaria growth (Jayasankar et al., 2006). Alkaline waters with a pH value of 7 - 9 are productive waters (Fourooghifard et al., 2018).

In line with this, the Gracilaria tenuistipitata study in brackishwater pond obtained DO 4.62 – 6.18 ppm (Sarkar et al., 2019). DO in Gracilaria corticata cultured with Vannamei shrimp ranged from 5.1 to 6.56 ppm (Fourooghifard et al., 2018). All living organisms need to be dissolved oxygen for respiration, metabolic processes, or the exchange of substances, producing energy for growth (Yulius et al., 2019). According to Amir (2019), the range of brightness values in seaweed cultivation Gracilaria sp. in the bracksishwater pond is 40 - 61 cm. The brightness in Gracilaria sp. ranged from 50 - 55 cm (Mapparimeng et al., 2019). Brightness is a variable related to the amount of light penetration into the waters the for photosynthesis process of seaweed.

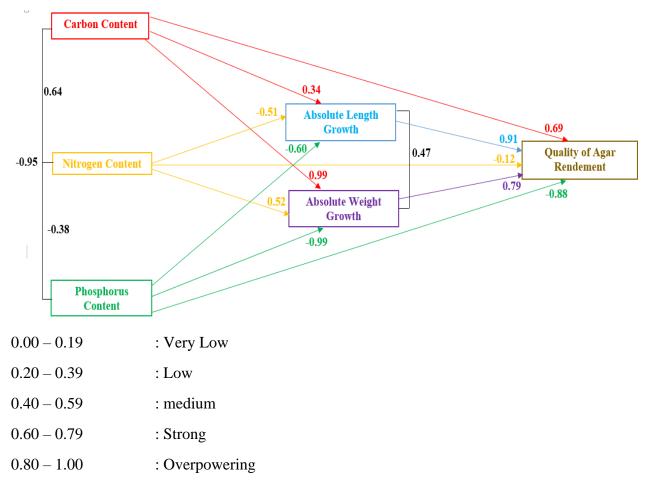
Dickson et al (2007) reported the range of Carbon in coastal waters is 1900 - 2090 ppm. A.R. Rahim (2018b), the range of seaweed carbon in brackishwater pond is 725.78 -4711.46 ppm. The high carbon content in brackishwater pond waters provides fertilizer, feeding, and metabolic processes. Takahashi et al (2006), carbon content is influenced by applying fertilizers and nutrients and carbonate material that enters coastal waters through rivers. A.R. Rahim (2018b), the range of brackishwater pond nitrogen is 14.61 – 94.99 ppm. Fourooghifard et al (2018) obtained the degree of nitrogen content in seaweed Gracilaria sp. of 7.63 - 16.70 ppm. Nitrogen deficiency is characterized by a change in the color of the thalus in red algae to pale (Moore, 1991). The range of phosphorus content obtained in brackishwater pond is 22.02 -24.22 ppm (A.R. Rahim, 2018a). Meanwhile, it was accepted by Tarigan and Edward (2003), phosphorus levels in sea waters ranged from 1.076 to 2.198 ppm. The high content of phosphorus in brackishwater pond is due to the addition of phosphorus fertilizer used to stimulate growth (Anam, 2007).

Path Analysis

	Content of Carbon (%)	Content of Nitrogen	Content of Phosphorus	Absolute Length	Absolute Weight	Quality of Agar Rendement
	(%)	(%)	(%)	(cm)	(g)	(%)
Content of						
Carbon	1.00					
(%)						
Content of						
Nitrogen	0.64	1.00				
(%)						
Content of						
Phosphorus	-0.95	-0.38	1.00			
(%)						
Absolute						
Length	0.34	-0.51	-0.60	1.00		
(cm)						
Absolute						
Weight	0.99	0.52	-0.99	0.47	1.00	
(g)						
Quality of						
Agar	0.69	-0.12	-0.88	0.91	0.79	1.00
Rendement	0.09	-0.12	-0.00	0.91	0.79	1.00
(%)						

Table 2. Pearson analysis (Product Moment Correlation) nutrient content, quality of agar
and growth of G. verrucosa for Different Density (Milkfish m-2 : Vannamei Shrimp m-2 :
Seaweed G. verrucosa g m-2) in Polyculture Brackishwater Pond.

Figure 6. Path Analysis Quality of Agar Rendement (%) of G. verrucosa for Different Density (Milkfish m-2 : Vannamei Shrimp m-2 : Seaweed G. verrucosa g m-2) in Polyculture Brackishwater Pond. Determining the model using correlation analysis, with a range of 0.00 - 1.00, getting closer to 1.00, strengthening the relationship between these parameters in producing a quality of agar rendement G. verrucosa.



From Path Analysis, the relationship between Carbon, Nitrogen, and Phosphorus content of G. verrucosa seaweed is an independent variable. The growth of absolute weight and absolute length of G. verrucosa seaweed is an intermediate variable. Agar quality of G. verrucosa seaweed rendement is the dependent variable. The independent and intermediary variables that affect the increase in the dependent variable are the quality of agar rendement G. verrucosa seaweed.

The carbon content with a strong category affects the quality of the rendement agar with a positive correlation value of 0.69, which is a 69% increase in the carbon content followed by the rise in the quality of the rendement agar.

Carbon is an essential factor in improving the quality of seaweed agar, the final product of red seaweed Gracilaria sp. Seaweed needs Carbon to produce carbohydrates in the photosynthesis process (Diniz et al. 2013, Chakraborty and Santra 2008). The most abundant substance in seaweed is found in cell walls, such as agarose. Then shallow nitrogen content affects the rendement agar quality with a negative correlation value of 0.12 or 12%, an increase in nitrogen content followed by a decrease in rendement agar quality. Nitrogen content can increase the growth of G. verrucosa but has a negative relationship with the formation of agarose (Ak et al., 2011). Phosphorus content strongly affects the rendement agar quality with a negative correlation value of 0.88, namely an

88% increase in phosphorus content followed by a decrease in rendement agar quality. Briggs and Funge-Smith (1993) state that phosphorus content is significant, but if the dose is excessive in water, it can inhibit growth. Seaweed growth that is less than perfect indirectly affects the gelatin content of seaweed rendement (Pong-masak et al., 2010).

The growth of absolute length very strongly affects the quality of agar rendement with a positive correlation value of 0.91 or 91%. An increase follows an increase in the whole distance in the quality of agar rendement. Erlania et al (2013), the morphology of seaweed Gracilaria sp. It has a long thalus; hence, it is more efficient in absorbing sunlight needed in photosynthesis. The process of photosynthesis will produce the final product of seaweed in the form of agarose. As the length of the thalus increases, the rate of photosynthesis will increase (Stewart, H.L and Carpenter, 2003). The growth of absolute weight strongly influences the quality of agar rendement; the positive correlation value is 0.79, which is 79% increase in growth of absolute weight followed by an increase in the quality of agar rendement. In the previous study by Andika Putra Syam and Suardi (2020), the agar content of seaweed is affected by the weight of the thalus.

The figure showed significant differences in the density of milkfish, Vannamei shrimp, and G. verrucosa seaweed in Polyculture Extensive brackishwater pond. They could produce nutrient content of Carbon, nitrogen, and phosphorus, which are used to increase absolute weight and absolute length growth in delivering the best final product from seaweed in the form of agar rendement content.

CONCLUSION

In conclusion, a significantly different density of milkfish, Vannamei shrimp, and Gracilaria verrucosa greatly affected the nutrient content of Carbon, absolute weight, specific growth rate, absolute length, and quality of agar rendement G. verrucosa seaweed. While the nutrient content of nitrogen, phosphorus, Viscosity, and gel strength, G. verrucosa seaweed did not have a significant effect. The supporting parameters in brackishwater pond water quality are Temperature, pH, DO, Salinity, Brightness, Carbon, Nitrogen, and Phosphorus. They are in the range that can meet the growth and quality of agar-rendement G. verrucosa seaweed. From the path analysis results, the content of carbon and absolute weight has a strong influence on improving the quality of agar-rendement. The absolute length has a very strong influence on improving the quality of of agar-rendement. The content of nitrogen and phosphorus has an inversely proportional effect on the quality of of agarrendement in an extensive brackishwater pond with polyculture systems. The density of the commodities three in an extensive brackishwater pond with the right polyculture system is needed to produce optimal growth and quality of agar seaweed.

ACKNOWLEDGEMENT

We gratefully acknowledge the help provided by the people of Soko Village, Glagah District, Lamongan Regency, East Java Province, Indonesia, who have provided support in the form of a brackishwater pond for this research. We also thank Muhammadiyah University of Gresik, Indonesia, for providing financial assistance for this research. This research did not receive any specific funding.

Reference

Ak, I., Çetin, Z., Cirik, Ş., & Göksan, T. (2011). Gracilaria verrucosa (Hudson) papenfuss culture using an agricultural organic fertilizer. Fresenius Environmental Bulletin, 20(8 A), 2156–2162.

- Amir, M. R. (2019). Studi Kelayakan Tambak Untuk Budidaya Rumput Laut (Gracilaria sp) Di Desa Panyiwi Kecamatan Cenrana Kabupaten Bone. Jurnal Environmental Science, 1(2). https://doi.org/10.35580/jes.v1i2.9061
- Anam, M. S. (2007). Guidelines for Polyculture Cultivation of Seaweed, Milkfish and Shrimp in Ponds. Food Security and Agricultural Extension Office District of Pasuruan.
- Andika Putra Syam, Suardi, M. S. (2020). Analisis pertumbuhan dan kandungan agar rumput laut Gracilaria sp. Dengan lokasi berbeda di perairan pesisir kabupaten luwu. Fisheries of Wallacea Journal, 1(1), 24–30.
- Anton, A. (2017). Pertumbuhan dan Kandungan Agar Rumput Laut (Gracilaria spp) Pada Beberapa Tingkat Salinitas. Jurnal Airaha, 6(2), 054–064. https://doi.org/10.15578/ja.v6i2.70
- Bird, C. J., McLachlan, J., & Oliveira, E. C. de. (1986). Gracilaria chilensis sp.nov. (Rhodophyta, Gigartinales), from Pacific South America. Canadian Journal of Botany, 64(12), 2928–2934. https://doi.org/10.1139/b86-387
- Briggs, M. R. P., & Funge-Smith, S. J. (1993).
 Macroalgae in aquaculture: an overview and their possible roles in shrimp culture.
 In Proceedings conference on marine biotechnology in the Asia Pacific (pp. 137–143).
- Chakraborty, S., & Santra, S. C. (2008). Biochemical composition of eight benthic algae collected from Sunderban. Indian Journal of Marine Sciences, 37(3), 329– 332.

- Choi, H. G., Kim, Y. S., Kim, J. H., Lee, S. J., Park, E. J., Ryu, J., & Nam, K. W. (2006). Effects of temperature and salinity on the growth of Gracilaria verrucosa and G. chorda, with the Potential for mariculture in Korea. Journal of Applied Phycology, 18(3-5), 269–277. https://doi.org/10.1007/s10811-006-9033y
- Colman, B. P. (2010). Understanding and eliminating iron interference in colorimetric nitrate and nitrite analysis. Environmental Monitoring and Assessment, 165(1-4), 633–641. https://doi.org/10.1007/s10661-009-0974x
- Dawes, C.J., Lluis, A.O. Trono, G. C. (1994). Laboratory and Field growth studies of commercial stains of Eucheuma denticulatus and Kappaphycus alvarezii in the Philippines. Applied Phycology, 6, 21– 24.
- De Castro, J. P. L., Costa, L. E. C., Pinheiro, M. P., Dos Santos Francisco, T., De Vasconcelos, P. H. M., Funari, L. M., Daudt, R. M., Dos Santos, G. R. C., Cardozo, N. S. M., & Freitas, A. L. P. (2018). Polysaccharides of red alga Gracilaria intermedia: Structure, antioxidant activity and rheological behavior. Polimeros, 28(2), 178–186. https://doi.org/10.1590/0104-1428.013116
- Dickson, A. G., Sabine, C. L., & Christian, J. R. (2007). Guide to Best Practices for Ocean CO2 measurements. PICES Special Publication. In Guide to Best Practices for Ocean CO2 measurements. PICES Special Publication (Vol. 3, Issue 8).
- Diniz, G. S., Barbarino, E., Oiano-Neto, J., Pacheco, S., & Lourenço, S. O. (2013). Perfil químico bruto y cálculo de los factores de conversión de nitrógeno a proteína en nueve especies de peces de aguas costeras de Brasil. Latin American

Journal of Aquatic Research, 41(2), 254-264. https://doi.org/10.3856/vol41-issue2fulltext-5

- Directorate General of Aquaculture. (2018). Center for Statistics and Information Data. Ministry of Marine Affairs and Fisheries.
- Effendi, M. I. (1997). Methods of Fisheries Biology. Gramedia Main Library.
- Erlania, E., Nirmala, K., & Soelistyowati, D. T. Penyerapan (2013). Karbon Pada Budidaya Rumput Laut Kappaphycus alvarezii dan Gracilaria gigas Di Perairan Teluk Gerupuk, Lombok Tengah, Nusa Tenggara Barat. Jurnal Riset Akuakultur, 8(2), 287. https://doi.org/10.15578/jra.8.2.2013.287-297
- Faturrahman, F., Meryandini, A., Junior, M. Z., I. (2011). Isolation Rusmana, and identification of an agar-liquefying marine bacterium and some properties of its extracellular agarases. **Biodiversitas** Journal of Biological Diversity, 12(4), 192–197.

https://doi.org/10.13057/biodiv/d120402

- Fortes, M. D. (1989). Seagrasses: A Resource Unknown in the ASEAN Region. ICLARM Education Series 5, 46p. (Issue November).
- Fourooghifard, H., Matinfar, A., Mortazavi, M. Roohani Ghadikolaee, K., & S.. Mirbakhsh, M. (2018). Nitrogen and phosphorous budgets for integrated culture of whiteleg shrimp Litopenaeus vannamei with red seaweed Gracilaria corticata in zero water exchange system. Iranian Journal of Fisheries Sciences, 17(3), 471-486. https://doi.org/10.22092/IJFS.2018.11638 2
- Gioele, C., Marilena, S., Valbona, A., Nunziacarla, S., Andrea, S., Antonio, M.

(2017). Gracilaria gracilis, Source of Agar: Short Review. Current Organic Α Chemistry, 380-386. 21(5). https://doi.org/10.2174/138527282066616 1017164605

10(3S) 2145-2162

- Israel, A., Guttman, L., Shpige, M., Neori, A. (2017). Development of Polyculture and Integrated Multi -Trophic Aquaculture (IMTA) in Israel: A Review. Israeli Journal of Aquaculture - Bamidgeh, January.
 - https://doi.org/10.46989/001c.21051
- Isroni, W., Bahri, A. S., & Amin, A. A. (2020). The effect of using the initial weight of seedlings by the floating method on the percentage of daily growth of seaweed eucheuma cottonii. IOP Conference Series: Earth and Environmental Science, 441(1). https://doi.org/10.1088/1755-1315/441/1/012132
- Istiqomawati, R. K. (2010). Technique of Seaweed Culture at Brackwish Water Aquaculture Development Center Situbondo of East Java. Jurnal Ilmiah Perikanan Dan Kelautan, 2(1), 44–51.
- Jayasankar, R., Seema, C., Leelabhai, K. S., & Kanagam, A. (2006). Pond Based Grow Out System of Gracilaria. Journal of Aquaculture in the Tropics, 21(3), 161-167.
- Klionsky, D. J., Abdelmohsen, K., Abe, A., Abedin, M. J., Abeliovich, H., Arozena, A. A., Adachi, H., Adams, C. M., Adams, P. D., Adeli, K., Adhihetty, P. J., Adler, S. G., Agam, G., Agarwal, R., Aghi, M. K., Agnello, M., Agostinis, P., Aguilar, P. V., Aguirre-Ghiso, J., Zughaier, S. M. (2016). Guidelines for the use and interpretation of assays for monitoring autophagy (3rd edition). Autophagy, 12(1), 1-222.https://doi.org/10.1080/15548627.2015.11 00356

- Kristanto, A., Pantjara, B., Insan, I. (2013). Polyculture of tiger prawns, Tilapia, Milkfish, and Seaweed in Idle Ponds, Tangerang Regency, Banten. Proceedings of the Aquaculture Technology Innovation Forum, 183–191.
- Kumar, M., Kumari, P., Gupta, V., Reddy, C.
 R. K., & Jha, B. (2010). Biochemical responses of red alga Gracilaria corticata (Gracilariales, Rhodophyta) to salinity induced oxidative stress. Journal of Experimental Marine Biology and Ecology, 391(1-2), 27–34. https://doi.org/10.1016/j.jembe.2010.06.0 01
- Lapointe, B. E. (1984). Biochemical Strategies For Growth Of Gracilaria tikvahiae (Rhodophyta) In Relation To Light Insentity And Nitrogen Availabity. J.Phycol, 20(1), 488–495.
- Mapparimeng, Liswahyuni, A., Permatasari, A., Fattah, N., A. (2019). Growth Rate of Seaweed (Gracilaria sp) With Tiered Shelf Pattern in Ponds, Samataring Village, East Sinjai District, Sinjai Regency. Jurnal Agrominansia, 4(1), 71–82.
- Marinho-Soriano, E., Morales, C., & Moreira,
 W. S. C. (2002). Cultivation of Gracilaria (Rhodophyta) in shrimp ponds effluents in Brazil. Aquaculture Research, 33, 1081– 1086.
- Martín, L. A., Rodríguez, M. C., Matulewicz, M. C., Fissore, E. N., Gerschenson, L. N., & Leonardi, P. I. (2013). Seasonal variation agar composition in and properties Gracilaria gracilis from (Gracilariales, Rhodophyta) of the Patagonian coast of Argentina. Phycological Research, 61(3), 163–171. https://doi.org/10.1111/pre.12000
- Martínez-Porchas, M., Martínez-Córdova, L. R., Porchas-Cornejo, M. A., & López-Elías, J. A. (2010). Shrimp polyculture: a

potentially profitable, sustainable, but uncommon aquacultural practice. Reviews in Aquaculture, 2(2), 73–85. https://doi.org/10.1111/j.1753-5131.2010.01023.x

- Matinfar, M., Rafiee, F., Nejatkhah Manavi, P., Joon Lee, I., & Hong, Y. K. (2013).
 Optimal conditions for tissue growth and branch induction of Gracilariopsis persica.
 Iranian Journal of Fisheries Sciences, 12(1), 24–33.
- Moore, J. W. (1991). Inorganic Contaminants of Surface Water Research and Monitoring Priorities. In Springer-Verlag (p. 334).
- Muarif, Yala, Z.R., R. (2017). Growth of Eucheuma cottonii Seaweed Cultured In Vitro with Different Numbers of Thallus. In Proceedings of the IV National Symposium on Maritime Affairs and Fisheries (pp. 251–259).
- Mulatsih, S. (2015). Model Optimasi Pengelolaan Kualitas Lingkungan melalui Peran Biofilter Rumput Laut (Gracilaria sp.) untuk Pengembangan Tambak yang Berkelanjutan. 9(01), 84–89.
- Mulyaningrum, S. R. H., Suwoyo, H. S. (2018). Growth, Agar Yield and Water Quality Variables Affecting Mass Propagation of Tissue Cultured Seaweed Gracilaria verrucosa in Pond. Ilmu Kelautan: Indonesian Journal of Marine Sciences, 23(1), 55. https://doi.org/10.14710/ik.ijms.23.1.55-62
- Myco Supply. (2011). Myco Supply COA Certificate of Analysis. Myco Supply Company, Inc. http://www.mycosupply.com/coa
- Nana, S.S., U. P. (2008). Manajemen kualitas tanah dan air dalam kegiatan perikanan budidaya. Balai Budidaya Air Payau, Takalar. In Dirjen Perikanan Budidaya

Departemen Kelautan dan Perikanan (p. 27). Dirjen Perikanan Budidaya Departemen Kelautan dan Perikanan Sulawesi Selatan.

- Niu, J., Xu, M., Wang, G., Zhang, K., & Peng, G. (2013). Comprehensive extraction of agar and R-phycoerythrin from gracilaria lemaneiformis (Bangiales, Rhodophyta). Indian Journal of Marine Sciences, 42(1), 21–28.
- Ollando, J. A., Mwakumanya, M. A., & Mindra, B. (2019). The viability of red alga (Gracilaria salicornia) seaweed farming for commercial extraction of agar at kibuyuni in kwale county South Coast Kenya. International Journal of Fisheries and Aquatic Studies, 7(2), 175–180.
- Orosco, C. A., Anong, C., Nukaya, M., Ohno, M., Sawamura, M., & Kusunose, H. (1992). Yield and Physical Characteristics of Agar from Gracilaria chorda Holmes: Comparison with Those from Southeast Asian Species. Nippon Suisan Gakkaishi, 58(9), 1711–1716. https://doi.org/10.2331/suisan.58.1711
- Oyieke, H. A. (1993). The yield, physical and chemical properties of agar gel from Gracilaria species (Gracilariales, Rhodophyta) of the Kenya coast. Hydrobiologia, 260-261(1), 613–620. https://doi.org/10.1007/BF00049079
- Painter, T. J. (1983). Algal polysaccharides. In: Aspinall GO (ed) The polysaccharides. Vol. II. Academic.
- Pakker, H., Breeman, A. M., Prud'homme van Reine, W. F., & Hock, C. (1995). A Comparative Study of Temperature Responses of Caribbean Seaweeds from Different Biogeographic Groups. Journal of Phycology, 31(4), 499–507. https://doi.org/10.1111/j.1529-8817.1995.tb02543.x

- Pantjara, B., M. Mangampa., R. (2010). Cultivation of tiger prawns, Penaeus monodon, in acid sulphate ponds in Tarakan, East Kalimantan. J.Perikanan, 12(1), 1–10.
- Pong-masak, P., Asaad, A. I., Hasnawi, H., Pirzan, A., Lanuru, M. (2010). Analisis Kesesuaian Lahan Untuk Pengembangan Budidaya Rumput Laut Di Gusung Batua , Pulau Badi. Ris. Akuakultur, 5(2), 299– 316.
- Sugiyono. (2010). Metode penelitian kuantitatif kualitatif. Bandung Alf (p. 143).
- Rahim, A.R., and Rosmarlinasiah. (2021). Productivity Improvement Of Seaweed (Gracilaria verrucosa) Fertilized With Vermicompost Made From Different Organic Waste. Indian Journal of Environmental Protection, 41(6), 613–620. https://www.e-ijep.co.in/41-6-613-620/
- Rahim, A. R. (2017). The Content of Agar Seaweed Gracilaria vertucosa Fertilized with Vermicompost. International Journal of Environment, Agriculture and Biotechnology, 2(4), 1879–1884. https://doi.org/10.22161/ijeab/2.4.51
- Rahim, A. R. (2018a). Application of seaweed gracilaria verrucosa tissue culture using different doses of vermicompost fertilizer. Nature Environment and Pollution Technology, 17(2).
- Rahim, A. R. (2018b). Utilization of organic wastes for vermicomposting using lumbricus rubellus in increasing quality and quantity of seaweed Gracilaria verrucosa. Asian Journal of Microbiology, Biotechnology and Environmental Sciences, 20(2).
- Rahim, A. R., Herawati, E. Y., Nursyam, H., & Hariati, A. M. (2016). Combination of vermicompost fertilizer, carbon, nitrogen

and phosphorus on cell characteristics, growth and quality of agar Seaweed Gracilaria verrucosa. Nature Environment and Pollution Technology, 15(4), 1153–1160.

- Raikar, S. V., Iima, M., Fujita, Y. (2001). Effect of temperature, salinity and light intensity on the growth of Gracilaria spp. (Gracilariales, Rhodophyta) from Japan, Malaysia and India. Indian Journal of Marine Sciences, 30(2), 98–104.
- Rosyida, E., Surawidjaja, E. H., Suseno, S. H., & Supriyono, E. (2014). Teknologi Pengkayaan Unsur-Unsur N, P, Fe Pada Rumput Laut Gracilaria verrucosa. Jurnal Kelautan Nasional, 8(3), 127. https://doi.org/10.15578/jkn.v8i3.6232
- Samidjan, I., Rachmawati, D., Indarji, A., & Hadi, P. (2018). Rekayasa Teknologi Polikultur Udang Vanamei dan Rumput Laut Pada Jarak Tanam Berbeda Terhadap Percepatan Pertumbuhan dan Kelulushidupan. Prosiding Seminar Nasional Kelautan Dan Perikanan IV 2018, 1(September), 249–255.
- Sarkar, S., Rekha, P. N., Biswas, G., Ghoshal, T. Ambasankar, K., K.. & Balasubramanian, C. P. (2019). Culture Potential of the Seaweed, Gracilaria (Rhodophyta) tenuistipitata in Brackishwater Tide Fed Pond System of Sundarban, India. Journal of Coastal Research. 86(sp1), 258-262. https://doi.org/10.2112/SI86-038.1
- Sornalakshm, V. (2017). Effects of Season on the Yield and Properties of Agar from gracilaria coticata. International Journal of Science, Engineering and Management (IJSEM), 2(12), 206–211.
- Stewart, H.L., Carpenter, R. C. (2003). The effecs of morphology and water flow on photosynthesis of marine macroalgae. Ecology, 84(11), 2,999–3,012.

- Stiger-Pouvreau, V., Bourgougnon, N., Deslandes, E. (2016). Carbohydrates from seaweeds (pp. 223–274). In: Fleurence J, Levine I (eds) Health and disease prevention. Academic.
- Susilowati, T., S. Rejeki, E. N. Dewi., Z. (2012). Effect of Depth on Growth Seaweed (Eucheuma cottonii) Cultivated Using the Longline Method at Mlonggo Beach, Jepara Regency. Jurnal Saintek Perikanan, 8(1), 7–12.
- Takahashi, T., Sutherland, S. C., Feely, R. A., & Wanninkhof, R. (2006). Decadal change of the surface water pCO2 in the North Pacific: A synthesis of 35 years of observations. Journal of Geophysical Research: Oceans,111(7).https://doi.org/10.1029/20 05JC003074
- Tarigan, M., & Edward. (2003). Kandungan Total Zat Padat Tersuspensi (Total suspenden Solid) Di Perairan Raha, Sulawesi Tenggara. Makara, 7(3), 109– 119.
- Tsai, C. C., Chang, J. S., Sheu, F., Shyu, Y. T., Yu, A. Y. C., Wong, S. L., Dai, C. F., & Lee, T. M. (2005). Seasonal growth dynamics of Laurencia papillosa and Gracilaria coronopifolia from a highly eutrophic reef in southern Taiwan: Temperature limitation and nutrient availability. Journal of Experimental Marine Biology and Ecology, 315(1), 49– 69. https://doi.org/10.1016/i.jembe.2004.08.0

https://doi.org/10.1016/j.jembe.2004.08.0 25

- Valderrama D, Cai J, H. N. (2014). Social and economic dimensions of carrageenan seaweed farming in Indonesia. In Social and economic dimensions of carrageenan seaweed farming.
- Waluyo, W., Permadi, A., Fanni, N. A., & Soedrijanto, A. (2019). Analisis Kualitas

Rumput Laut Gracilaria Verrucosa di Tambak Kabupaten Karawang, Jawa Barat. Grouper, 10(1), 32. https://doi.org/10.30736/grouper.v10i1.50

- Wenno, M. R., Thenu, J. L., & Cristina Lopulalan, C. G. (2012). Karakteristik Kappa Karaginan dari Kappaphycus alvarezii Pada Berbagai Umur Panen. Jurnal Pascapanen Dan Bioteknologi Kelautan Dan Perikanan, 7(1), 61.https://doi.org/10.15578/jpbkp.v7i1.69
- William Horwitz., George W. Latimer, J. (2006). Official Methods of Analysis of AOAC INTERNATIONAL. In AOAC INTERNATIONAL (Issue 18, p. 96).
- Wu, Chaoyuan, Li, Renzhi, Lin, G. (1994). Study on the optimum environmental parameters for the growth of Gracilaria tenuistipitata varliui in pond culture. Oceanologia et Limnologia Sinica, 25(1), 60–66.
- Xu, Y., Fang, J., & Wei, W. (2008). Application of Gracilaria lichenoides (Rhodophyta) for alleviating excess nutrients in aquaculture. Journal of Applied Phycology, 20(2), 199–203. https://doi.org/10.1007/s10811-007-9219y
- Yang, Y., Chai, Z., Wang, Q., Chen, W., He, Z., & Jiang, S. (2015). Cultivation of seaweed Gracilaria in Chinese coastal waters and its contribution to environmental improvements. Algal Research, 9, 236–244. ttps://doi.org/10.1016/j.algal.2015.03.017
- Yulistiana, U., Damayanti, A. A., & Cokrowati, N. (2020). Pertumbuhan Gracilaria sp. yang Dibudidayakan Pada Tambak di Bajo Baru Dompu. Rekayasa, 13(3), 212–218. https://doi.org/10.21107/rekayasa.v13i3.9 013

Yulius, Ramdhan, M., Prihantono, J., Pryambodo, D. G., Saepuloh, D., Salim, H. L., Rizaki, I., & Zahara, I. R. (2019).
Budidaya rumput laut dan pengelolaannya di pesisir Kabupaten Dompu, Provinsi Nusa Tenggara Barat berdasarkan analisa kesesuaian lahan dan daya dukung lingkungan. Segara, 15(1), 19–30.

10(3S) 2145-2162

- Yuniarsih, E., Nirmala, K., & Radiarta, I. N. (2014). Tingkat Penyerapan Nitrogen Dan Fosfor Pada Budidaya Rumput Laut Berbasis Imta (Integrated Multi-Trophic Aquaculture) Di Teluk Gerupuk, Lombok Tengah, Nusa Tenggara Barat. Jurnal Riset Akuakultur, 9(3), 487. https://doi.org/10.15578/jra.9.3.2014.487-500
- Zhou, W., Hu, Y., Sui, Z., Fu, F., Wang, J., Chang, L., Guo, W., & Li, B. (2013).
 Genome Survey Sequencing and Genetic Background Characterization of Gracilariopsis lemaneiformis (Rhodophyta) Based on Next-Generation Sequencing. PLoS ONE, 8(7). https://doi.org/10.1371/journal.pone.0069 909