

Preliminary study on phytonutrients constituents and the anti-oxidant potential of *Gracilaria opuntia* of Rameswaram Coast, India.

Mr. V.S. Chandrasekaran^{1,2}, Dr. M. Ganesh^{3*}, Dr. S. Latha⁴, Dr. Adlyne Reena Asirvatham⁵, Dr. M. Sivakumar⁶, Dr. K. Umasankar⁷,

¹ Research Scholar, Sri Ramachandra Faculty of Pharmacy, Sri Ramachandra Institute of Higher Education and Research (DU), Chennai. 600 116.

Email: chandrasekaranvs@sriramachandra.edu.in Mobile: +91 80721 07226

² Associate professor, Department of Pharmaceutical Biotechnology, Krishna Teja Pharmacy College, Tirupati. 517 506

Email: vschandru610@gmail.com Mobile: +91 80721 07226

³ *Professor and Head, Department of Biochemistry, Sri Ramachandra Medical College and Research Institute, Sri Ramachandra Institute of Higher Education and Research (DU), Chennai. 600 116

Email: ganeshm@sriramachandra.edu.in Mobile: +91 98841 99902

⁴ Assistant Professor, Department of Pharmacognosy, Sri Ramachandra Faculty of Pharmacy, Sri Ramachandra Institute of Higher Education and Research (DU), Chennai. 600 116

Email: latha.s@sriramachandra.edu.in Mobile: +91 99622 29445

⁵ Professor of Endocrinology and Consultant Endocrinologist, Sri Ramachandra Medical College and Research Institute, Sri Ramachandra Institute of Higher Education and Research (DU), Chennai. 600 116.

Email: adlyne@sriramachandra.edu.in Mobile: +91 98430 51977

⁶ Professor, HOD, Department of Pharmacognosy, Faculty of Pharmacy, Sree Balaji Medical College and Hospital, Bharat Institute of Science and Technology (DU), Chennai.

Email: sivampharma@gmail.com Mobile: +91 9884208394

⁷ Professor and Head, Department of Pharmaceutics, Krishna Teja Pharmacy College, Tirupati. 517 506.

Email: umasankar73@gmail.com Mobile: +91 94409 55960

*Corresponding Author; - Dr. M. Ganesh,

*Professor and Head, Department of Biochemistry, Sri Ramachandra Medical College and Research Institute, Sri Ramachandra Institute of Higher Education and Research (DU), Chennai. 600 116

Email: ganeshm@sriramachandra.edu.in

Mobile: +91 98841 99902

Abstract

Objective: Preliminary phytonutrient and invitro evaluation of *Gracilaria opuntia*.

Methods: Phytonutrient evaluation studied with various extracts like ethyl acetate, ethanol and purified water. The antioxidant potential was studied by DPPH, H₂O₂, NO and OH⁻ assays.

Results: The phytonutrient screening studies exhibits existence of various nutrient substituents. Also, a comprehensive anti-oxidant potential at 500 µg/ml in *Gracilaria opuntia* comparatively better than used standard.

Conclusion: Thus *Gracilaria opuntia* can be consider for curative disease from oxidative weakening in which *in-vivo* studies can be evaluated in advance.

Keywords: Gracilaria, Seaweed, Phytonutrient, Polyphenol compounds, Antioxidant.

INTRODUCTION

Natural ways stand as a golden mark to exemplify the outstanding phenomena of symbiosis. In India plants and herbal products in practice for health and medicinal purpose for several thousand years, there are nearly 1.5 million practitioners till using traditional medicine system such as Ayurveda, Siddha, Unani and Homeopathy. (Sheeta Verma *et al.*, 2008). The main benefits of herbal medicines

over allopathic are that it has least side effect, toxicity and cozy effective (Patil S Jolly *et al.*, 2003, Anggadireja J *et al.*, 1997, Bors W *et al.*, 1990). Antioxidants is a substance that is existing at low concentrations at which notably delays or inhibits oxidation of substrates. (Bhuvanewar *et al.*, 2013) They are preferred defense against free radical damage and maintain finest health for well -being. (Vishnu Kiran 2014, Velavan S 2011, Halliwell B *et al.*,

1997, Ramdani M *et al.*, 2017). The main background of the research work is to evaluate the *in vitro* antioxidant activity of *Gracilaria opuntia* of various extracts based on the comparison with standard.

MATERIALS & METHODS:

Collection of Algae Sample:

Gracilaria opuntia was collected from R.K. Algae project centre, Rameswaram Coast, India. The seaweed was authenticated under Botanical Survey of India, Calcutta.

Preparation of Seaweed Extracts:

Gracilaria opuntia seaweed saturated in running water and dried, powdered weighed accurately in the ratio of 1:3, accordingly volumes of ethyl acetate, ethanol and distilled water separately extraction process conducted. (Gajalakshmi *et al.*, 2018). The ethyl acetate, ethanol and aqueous extracts were freeze-dried, and kept in air-tight plastic containers. (Eun-sun Hwang *et al.*, 2014)

Phytonutrient screening

Phytonutrient analysis for the test of alkaloids, tannins, flavonoids, steroids, terpenoids, triterpenoids, anthraquinones, polyphenols, proteins, glycoside and carbohydrates of all the extracts was conducted as per Indian pharmacopoeia. (O' Sullivan AM *et al.*, 2011)

***In Vitro* Antioxidant Studies**

DPPH (1,1-DI Phenyl-2-Picryl Hydrazyl) Scavenging Activity:

To find free radical scavenging activity, test samples (0.5ml seaweed sample and standard amino acid) of different concentrations (100-500µg/ml), (Qi H Zheo *et al.*, 2005, Ciz M *et al.*, 2010) and 0.3 ml of DPPH solution (0.5 mM dissolved in ethanol solvent) were prepared. Control which contains only 3.5 ml of ethanol and 0.3 ml DPPH solution. After 100 minutes, absorbance was measured at 517 nm. (Ganesan P *et al.*, 2008).

Hydrogen Peroxide Scavenging Activity Test:

100-500µg/ml of test seaweed samples and standard ascorbic acid were prepared by dissolving in suitable solvent (Duh PD *et al.*, 1998) to each add 3.4 ml of phosphate buffer

(pH- 7.4) along with 0.6 ml of prepared 40nM hydrogen peroxide solution, incubate at 37°C for 5 minutes and absorbance was recorded at 230nm. (Lim SN *et al.*, 2002, Darcy Vrillon B 1993)

Hydroxyl radical scavenging activity:

500µL of seaweed sample each (100-500µg/ml) mixed with 200 µL 1.04nM EDTA and 200 µL FeCl₃ (1:1 v/v), 100 µL of H₂O₂ (1.0mM) and 100 µL ascorbic acid as standard, incubated at 37°C for 1 h. After mixture treated with 1.0mL of thiobarbituric acid (1%) and 1.0 mL of trichloroacetic acid (2.8%) incubated at 100°C for 20min. after cooling, absorbance is measured at 532 nm, against a blank sample. (Becka EM *et al.*, 2004)

Nitric oxide scavenging activity

0.5 mL of seaweed extract at various concentration (100-500µg/ml) with 2 mL of 10 mM sodium nitroprusside in 0.5 mL phosphate buffer saline (pH 7.4) incubated at 25°C. (Frankel *et al.*, 2008) 0.5 mL of the incubated solution is mixed with 0.5 mL of Griess reagent after 2hrs of incubation at room temperature for 30min and absorbance measured at 546 nm. (Martinez Tome M *et al.*, 2001, Takamatsu S *et al.*, 2003)

STATISTICAL ANALYSIS

All the Experimental findings were assessed by using linear regression statistical analysis and the IC₅₀ was calculated by plotting the graph as concentration versus percentage inhibition using MS Excel version MS OFFICE 2021.

RESULTS AND DISCUSSION

Phytonutrient screening studies has found that very high concentration of Proteins, steroids, phenols and high concentration of tannins, flavonoids, terpenoids, alkaloids, anthroquinones and reducing sugars were presence in ethyl acetate and ethanol extracts and in aqueous extract where very few phytonutrient constituents at low concentration found shown in table 1.

***In vitro* antioxidant studies**

DPPH scavenging activity:

Activity of all three extracts and ascorbic acid as standard on DPPH scavenging activity at

various concentration (100-500 µg/ml) exhibited better percentage of inhibition comparable to standard at same concentration respectively. Maximum DPPH scavenging activity of ethyl acetate (500 µg/ml) showed highest inhibitory effect when compared to ethanol and aqueous extracts of seaweed shown in figure 1. The IC₅₀ value also better to the standard comparison shown in table 2.

Hydrogen Peroxide assay:

The highest inhibitory activity in all three seaweed extracts showed at 500 µg/ml and IC₅₀ value shown in table 3. Ethyl acetate extract and ethanol extract shows better hydrogen peroxide scavenging activity than aqueous extract shown in figure 2.

Hydroxyl radical scavenging activity:

The effect of Hydroxyl radical was found and results shown as percentage of inhibition rate in figure 3. Ethyl acetate extract (500 µg/ml) exhibited the inhibition of about 86.27 is slightly better than standard shown 78.63 at same concentration respectively shown in table 4.

Nitric Oxide scavenging assay:

Nitric oxide scavenging activity on all three-seaweed extract shown in figure 4. The results show that ethyl acetate 69.52, ethanol extract shows 53.33 and aqueous extract 31.32 respectively. The IC₅₀ value also shows ethyl acetate seaweed extract having relatively good compared to standard were shown in table 5.

DISCUSSION

In preliminary phytonutrient analysis ethyl acetate extract of *Gracilaria opuntia* exhibits superior contrast to ethanol and distilled water extracts. In DPPH scavenging activity and Hydrogen Peroxide assay 500µg/ml ethyl acetate extract exhibited better with highest inhibitory effect then other two extracts and standard. The Hydroxyl assay activity of ethyl acetate and ethanol extracts expressed almost similar in inhibitory effect with respect to standard. At the same time Nitric oxide antioxidant activity of *Gracilaria opuntia* ethyl acetate extract is superior at the concentration of 500 µg/ml. The IC₅₀ comparatively better when compared to standard used.

CONCLUSION

The study on phytonutrient analysis of *Gracilaria opuntia* extract of ethyl acetate, ethanol and aqueous solvents having the beneficial secondary metabolites in seaweed. Thus, *Gracilaria opuntia* shows an abundant source of structurally novel and biochemical active metabolites compounds. From the results it was concluded that among three seaweed extracts ethyl acetate shows good antioxidant potential when compared to standard ascorbic acid in *in vitro* assay. The better antioxidant potential of *Gracilaria opuntia* extract have it made due to phenolic compounds presence. Thus, studied *Gracilaria opuntia* can be used as potential source of healthy food in diets with potential health effects like antioxidative.

REFERENCES

1. Verma S, Singh S. Current and future status of herbal medicines. *Vetinary World*. 2008;2(2):347. <http://dx.doi.org/10.5455/vetworld.2008.347-350>
2. Bhuvanewari S, Murugesan S, Subha TS, Dhamotharan R, Shettu N. *In vitro* antioxidant activity of marine red algae *Chondrococcus hornemanni* and *Spyridia fusiformis*. 2013.
3. Velavan S. Free radicals in health and diseases. *Pharmacol Online*. 2011;1(1): 1062–77.
4. Halliwell B. Antioxidants and human disease: a general introduction. *Nutritional Review* 1997;55(1Pt2):S44-9; discussion S49-52. <http://dx.doi.org/10.1111/j.1753-4887.1997.tb06100.x>
5. Patil S Jolly CI, Narayanan S. Free radical scavenging activity of *acacia catechu* and *Rotula aquatica*: implications in cancer therapy. *Indian Drugs*. 2003;40:328–32.
6. Anggadiredja J. Antioxidant activity of *Sargassum polycystum* (Phaeophyta) and *Laurencia obtusa* (Rodophyta) from Seribu Island. *Jounal Applied Phycology*. 1997;9:477–9.
7. Bors W, Heller W, Michel C, Saran M. Flavonoids as antioxidants: Determination of radical scavenging efficiencies. *Methods Enzymology*. 1990;186:343–55.

8. Hwang E-S, Thi ND. Effects of extraction and processing methods on antioxidant compound contents and radical scavenging activities of Laver (*Porphyratenera*). *Preventive Nutritional Food Science*. 2014; 19 (1) : 40 –8. <http://dx.doi.org/10.3746/pnf.2014.19.1.040>
9. O’Sullivan AM, O’Callaghan YC, O’Grady MN, Queguineur B, Hanniffy D, Troy DJ, et al. *In vitro* and cellular antioxidant activities of seaweed extracts prepared from five brown seaweeds harvested in spring from the west coast of Ireland. *Food Chemistry*. 2011;126(3): 1064–70. <http://dx.doi.org/10.1016/j.foodchem.2010.11.127>
10. Ganesan P, Kumar CS, Bhaskar N. Antioxidant properties of methanol extract and its solvent fractions obtained from selected Indian red seaweeds. *Bio-resource Technology*. 2008;99(8):2717–23. <http://dx.doi.org/10.1016/j.biortech.2007.07.005>
11. Lim SN, Cheung PCK, Ooi VEC, Ang PO. Evaluation of antioxidative activity of extracts from a brown seaweed, *Sargassum siliquastrum*. *Journal Agricultural Food Chemistry*. 2002;50 (13):3862–6. Available from: <http://dx.doi.org/10.1021/jf020096b>
12. Becker EM, Nissen LR, Skibsted LH. Antioxidant evaluation protocols: Food quality or health effects. *European Food Research and Technology*. 2004;219(6): 561–71. Available from: <http://dx.doi.org/10.1007/s00217-004-1012-4>
13. Frankel EN, Meyer AS. Review: The problems of using one-dimensional methods to evaluate multifunctional food and biological antioxidants. *Journal Science Food Agriculture*. 2000;80:1925–41
14. Martinez-Tome M, Carcia-Carmona F, Murcia MA. Comparison of the antioxidants and pro-oxidants activities of broccoli amino acids with those of common food additives. *Journal Science Food Agriculture*. 2001; 81: 1019–26.
15. Cíž M, Cížová H, Denev P, Kratchanova M, Slavov A, Lojek A. Different methods for control and comparison of the antioxidant properties of vegetables. *Food Control*. 2010;21:518–23.
16. Duh PD. Antioxidant activity of burdock (*Arctium lappa* Linne): Its scavenging effect on free radical and active oxygen. *Journal of Am Oil Chemistry Society*. 1998;75:455–61.
17. Gajalakshmi D. N1 and Murugesan, S2*(2018): Phytochemical screening of marine red alga *Botryocladialeptopoda*. (JAgardh) Kylin. 2018;463–70.
18. Vishnu Kiran M. *In vitro* Antioxidant activity of silver nano-particles from *Colpomeniasinuosa* and *Halymeniaporophyroides*. *World Journal Pharmaceutical Science*. 2014;2(8):817–20.
19. Ramdani M, Elasri O, Saidi N, Elkhiaati N, Taybi FA, Mostareh M, et al. Evaluation of antioxidant activity and total phenol content of *Gracilaria bursa-pastoris* harvested in Nador lagoon for an enhanced economic valorization. *Chemical Biotechnology Agriculture*. 2017;4(1). Available from: <http://dx.doi.org/10.1186/s40538-017-0110-z>
20. Qi H, Zhao T, Zhang Q, Li Z, Zhao Z, Xing R. Antioxidant activity of different molecular weight sulfated polysaccharides from *Ulva pertusa* Kjellm (Chlorophyta). *J Applied Phycology*. 2005;17(6):527–34. <http://dx.doi.org/10.1007/s10811-005-9003-9>
21. Darcy-Vrillon B. Nutritional aspects of the developing use of marine macroalgae for the human food industry. *International Journal Food Science Nutraceuticals*. 1993;
22. Takamatsu S, Hodges TW, Rajbhandari I, Gerwick WH, Hamann MT, Nagle DG. Marine natural products as novel antioxidant prototypes. *Journal Natural Products*. 2003;66(5):605–8. <http://dx.doi.org/10.1021/np0204038>

Table 1. Phytochemical constituents of Gracilaria opuntia

Phytochemical Test	EA Extract	ETOL Extract	DW Extract
Alkaloids	++	++	+
Tannins	++	+	+
Saponins	+	+	+
Steroids	+++	++	-
Terpenoids	++	+	-
Triterpenoids	+	+	-
Flavonoids	++	++	+
Reduced sugars	++	++	+
Anthraquinones	++	++	-
Proteins	+++	++	+
Polyphenols	+	+	-
Glycoside	+	+	-
Carbohydrates	++	++	+

Table 2. Effect of EA, ETOL, DW extracts of Gracilaria opuntia of DPPH Scavenging activity.

S.NO	Concentration (µg/ml)	Ascorbic acid (STD)	Ethyl Acetate	Ethanol	Aqueous
1	100	29.86	27.08	27	6.89
2	200	32.25	35.58	30.22	13.28
3	300	38.46	42.67	35.49	16.17
4	400	46.22	54.30	45.5	18.73
5	500	58.42	62.99	51.2	23.36
IC ₅₀		444.06	359.84	495.58	1193.51

Table 3. Effect of EA, ETOL, DW extracts of Gracilaria opuntia of Hydrogen Peroxide Scavenging activity.

S.NO	Concentration (µg/ml)	Ascorbic acid (STD)	Ethyl Acetate	Ethanol	Aqueous
1	100	18.09	11.45	13.28	2.4
2	200	22.46	18.28	18.14	3.02
3	300	28.99	25.26	20.29	8.21
4	400	34.14	32.06	21.25	10.12
5	500	38.26	41.89	29.78	12.19
IC ₅₀		715.00	872.71	1037.97	2016.84

Table 4. Effect of EA, ETOL, DW extracts of Gracilaria opuntia of Hydroxyl Scavenging activity.

S.NO	Concentration (µg/ml)	Ascorbic acid (STD)	Ethyl Acetate	Ethanol	Aqueous
1	100	26.43	42.17	31.84	9.43
2	200	32.31	49.17	42.31	16.17
3	300	56.45	62.13	51.45	23.37
4	400	64.83	74.03	63.15	30.22
5	500	78.63	86.27	71.25	41.05
IC ₅₀		277.66	187.12	279.81	653.69

Table 5. Effect of EA, ETOL, DW extracts of Gracilaria opuntia of Nitric Oxide Scavenging activity.

S.NO	Concentration (µg/ml)	Ascorbic acid (STD)	Ethyl Acetate	Ethanol	Aqueous
1	100	17.83	19.43	17.13	7.02
2	200	30.21	31.24	24.12	11.07
3	300	49.62	46.74	33.14	17.05
4	400	61.42	57.34	41.32	22.31

5	500	81.23	69.52	53.33	31.28
	IC ₅₀	312.28	342.83	480.71	843.77

Figure 1. Percentage of inhibition of EA, ETOL, DW extracts of *Gracilaria opuntia* of DPPH Scavenging activity.

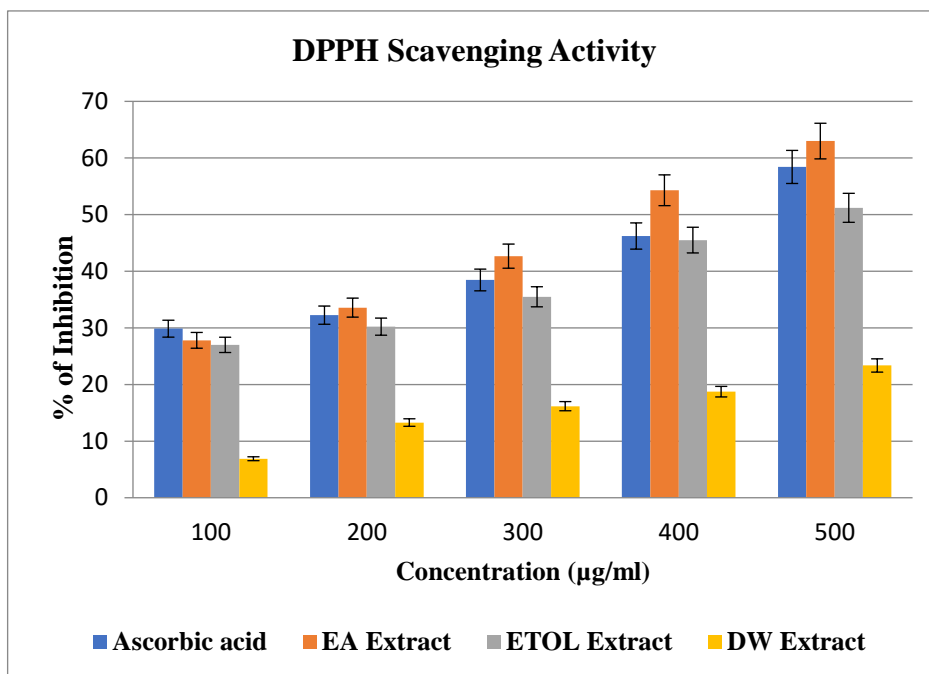


Figure 2. Percentage of inhibition of EA, ETOL, DW extracts of *Gracilaria opuntia* of Hydrogen Peroxide Scavenging activity.

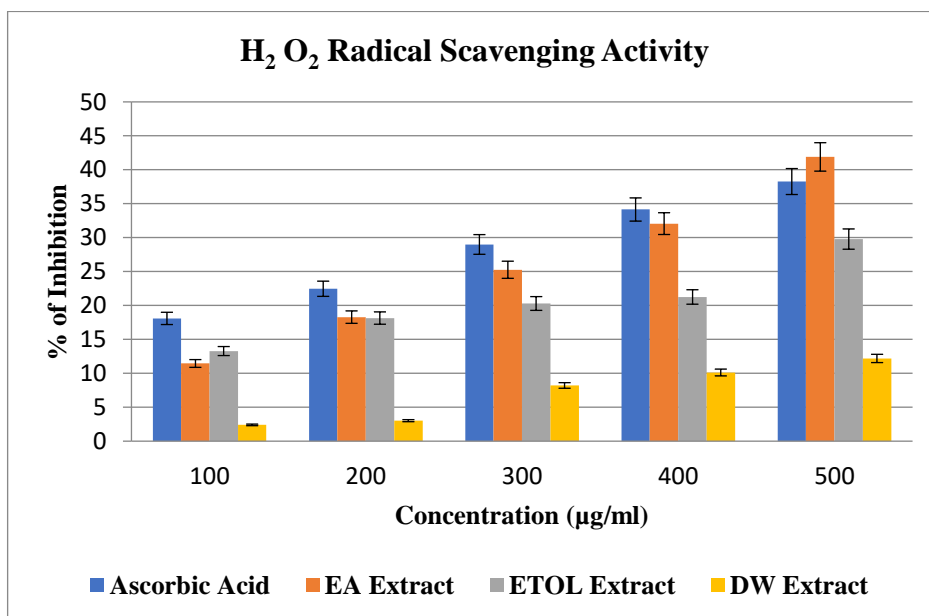


Figure 3. Percentage of inhibition of EA, ETOL, DW extracts of Gracilaria opuntia of Hydroxyl Scavenging activity.

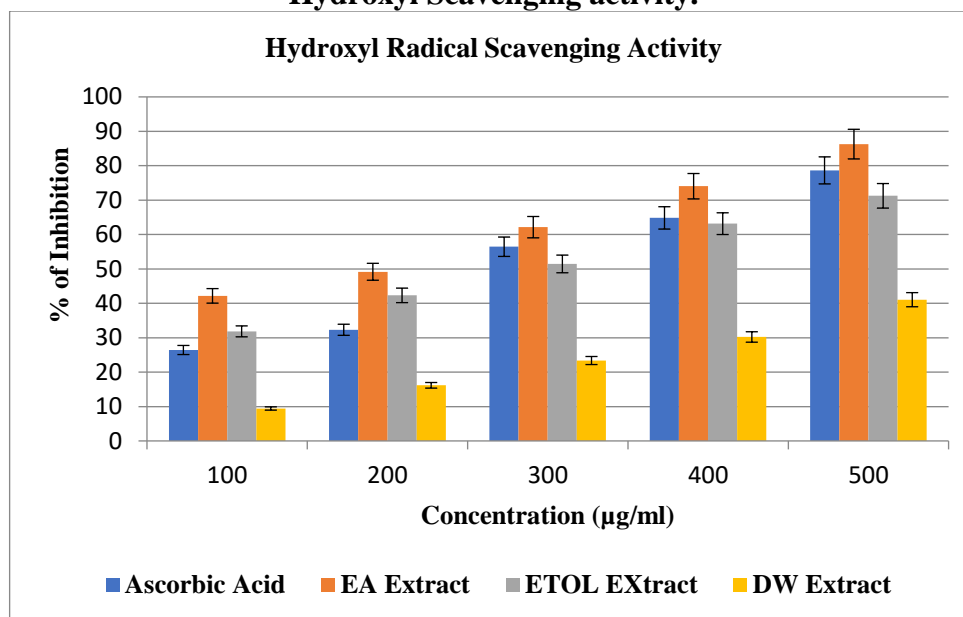
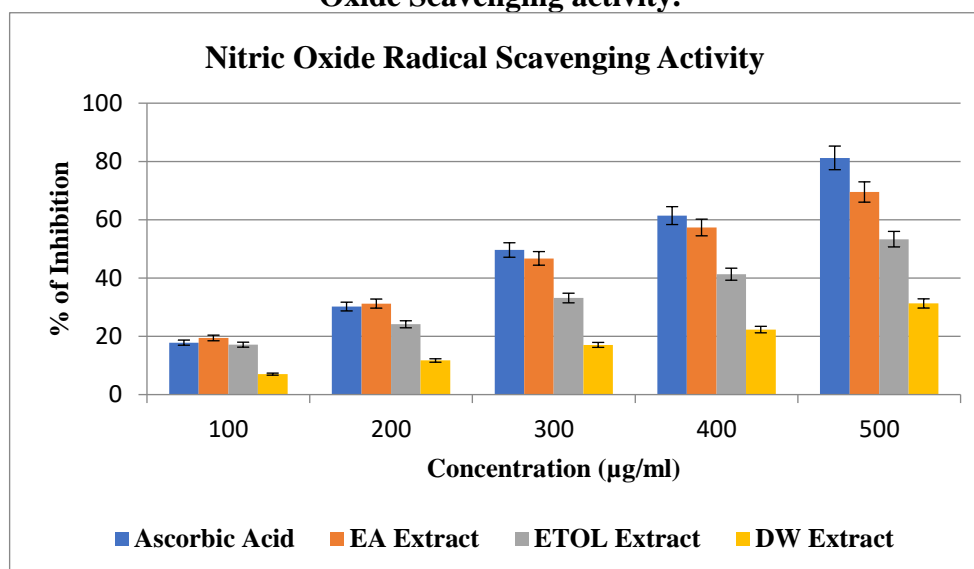


Figure 4. Percentage of inhibition of EA, ETOL, DW extracts of Gracilaria opuntia of Nitric Oxide Scavenging activity.



LIST OF ABBREVIATIONS

S.NO	CODE	ABBREVIATION
1.	ABR	Absorbance
2.	STD	Standard
3.	H2O2	Hydrogen peroxide
4.	DPPH	1,1-Diphenyl-2-picryl hydrazyl
5.	OH	Hydroxyl
6.	DNA	Deoxyribo nucleic acid
7.	ROS	Reactive oxygen species
8.	O2	Oxygen
9.	NOS	Nitric oxide synthase
10.	SOD	Super oxide dismutase
11.	BHT	Butylated hydroxyl toluene